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WORLD INTELLECTUAL  
PROPERTY ORGANIZATION



72

INTERNATIONAL APPLICATION PUBLISHED

(51) International Patent Classification 6 :

C12N 15/64, 15/67, 15/85, 9/72, 5/10

A1

WO 9604391A1

(43) International Publication Date: 15 February 1996 (15.02.96)

(21) International Application Number: PCT/US95/09576

(22) International Filing Date: 28 July 1995 (28.07.95)

(30) Priority Data:

08/286,740

5 August 1994 (05.08.94)

US

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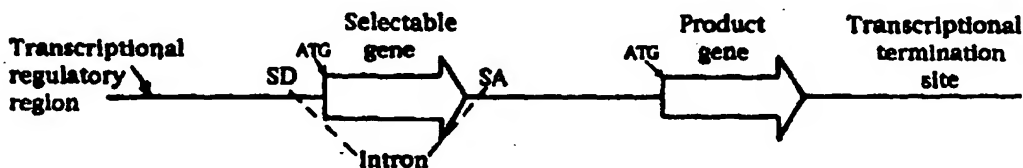
(81) Designated States: AU, CA, JP, MX, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE).

Published

*With international search report.*

*Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.*

(54) Title: METHOD FOR SELECTING HIGH-EXPRESSING HOST CELLS



(57) Abstract

A method for selecting recombinant host cells expressing high levels of a desired protein is described. This method utilizes eukaryotic host cells harboring a DNA construct comprising a selectable gene (preferably an amplifiable gene) and a product gene provided 3' to the selectable gene. The selectable gene is positioned within an intron defined by a splice donor site and a splice acceptor site and the selectable gene and product gene are under the transcriptional control of a single transcriptional regulatory region. The splice donor site is generally an efficient splice donor site and thereby regulates expression of the product gene using the transcriptional regulatory region. The transfected cells are cultured so as to express the gene encoding the product in a selective medium comprising an amplifying agent for sufficient time to allow amplification to occur, whereupon either the desired product is recovered or cells having multiple copies of the product gene are identified.

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METHOD FOR SELECTING HIGH-EXPRESSING HOST CELLSBACKGROUND OF THE INVENTIONField of the Invention

This invention relates to a method of selecting for high-expressing  
5 host cells, a method of producing a protein of interest in high yields and  
a method of producing eukaryotic cells having multiple copies of a sequence  
encoding a protein of interest.

Description of Background and Related Art

The discovery of methods for introducing DNA into living host cells  
10 in a functional form has provided the key to understanding many fundamental  
biological processes, and has made possible the production of important  
proteins and other molecules in commercially useful quantities.

Despite the general success of such gene transfer methods, several  
common problems exist that may limit the efficiency with which a gene  
15 encoding a desired protein can be introduced into and expressed in a host  
cell. One problem is knowing when the gene has been successfully  
transferred into recipient cells. A second problem is distinguishing  
between those cells that contain the gene and those that have survived the  
transfer procedures but do not contain the gene. A third problem is  
20 identifying and isolating those cells that contain the gene and that are  
expressing high levels of the protein encoded by the gene.

In general, the known methods for introducing genes into eukaryotic  
cells tend to be highly inefficient. Of the cells in a given culture, only  
a small proportion take up and express exogenously added DNA, and an even  
25 smaller proportion stably maintain that DNA.

Identification of those cells that have incorporated a product gene  
encoding a desired protein typically is achieved by introducing into the  
same cells another gene, commonly referred to as a selectable gene, that  
encodes a selectable marker. A selectable marker is a protein that is  
30 necessary for the growth or survival of a host cell under the particular  
culture conditions chosen, such as an enzyme that confers resistance to an  
antibiotic or other drug, or an enzyme that compensates for a metabolic or  
catabolic defect in the host cell. For example, selectable genes commonly  
used with eukaryotic cells include the genes for aminoglycoside  
35 phosphotransferase (APH), hygromycin phosphotransferase (hyg),  
dihydrofolate reductase (DHFR), thymidine kinase (tk), neomycin, puromycin,  
glutamine synthetase, and asparagine synthetase.

The method of identifying a host cell that has incorporated one gene  
on the basis of expression by the host cell of a second incorporated gene  
40 encoding a selectable marker is referred to as cotransfection (or  
cotransfection). In that method, a gene encoding a desired polypeptide and  
a selection gene typically are introduced into the host cell  
simultaneously, although they may be introduced sequentially. In the case  
of simultaneous cotransfection, the gene encoding the desired polypeptide

and the selectable gene may be present on a single DNA molecule or on separate DNA molecules prior to being introduced into the host cells. Wigler et al., Cell, 16:777 (1979). Cells that have incorporated the gene encoding the desired polypeptide then are identified or isolated by  
5 culturing the cells under conditions that preferentially allow for the growth or survival of those cells that synthesize the selectable marker encoded by the selectable gene.

The level of expression of a gene introduced into a eukaryotic host cell depends on multiple factors, including gene copy number, efficiency  
10 of transcription, messenger RNA (mRNA) processing, stability, and translation efficiency. Accordingly, high level expression of a desired polypeptide typically will involve optimizing one or more of those factors.

For example, the level of protein production may be increased by covalently joining the coding sequence of the gene to a "strong" promoter  
15 or enhancer that will give high levels of transcription. Promoters and enhancers are nucleotide sequences that interact specifically with proteins in a host cell that are involved in transcription. Kriegler, Meth. Enzymol., 185:512 (1990); Maniatis et al., Science, 236:1237 (1987). Promoters are located upstream of the coding sequence of a gene and  
20 facilitate transcription of the gene by RNA polymerase. Among the eukaryotic promoters that have been identified as strong promoters for high-level expression are the SV40 early promoter, adenovirus major late promoter, mouse metallothionein-I promoter, Rous sarcoma virus long terminal repeat, and human cytomegalovirus immediate early promoter (CMV).

25 Enhancers stimulate transcription from a linked promoter. Unlike promoters, enhancers are active when placed downstream from the transcription initiation site or at considerable distances from the promoter, although in practice enhancers may overlap physically and functionally with promoters. For example, all of the strong promoters  
30 listed above also contain strong enhancers. Bendig, Genetic Engineering, 7:91 (Academic Press, 1988).

The level of protein production also may be increased by increasing the gene copy number in the host cell. One method for obtaining high gene copy number is to directly introduce into the host cell multiple copies of  
35 the gene, for example, by using a large molar excess of the product gene relative to the selectable gene during cotransfection. Kaufman, Meth. Enzymol., 185:537 (1990). With this method, however, only a small proportion of the cotransfected cells will contain the product gene at high copy number. Furthermore, because no generally applicable, convenient  
40 method exists for distinguishing such cells from the majority of cells that contain fewer copies of the product gene, laborious and time-consuming screening methods typically are required to identify the desired high-copy number transfectants.

Another method for obtaining high gene copy number involves cloning  
45 the gene in a vector that is capable of replicating autonomously in the host cell. Examples of such vectors include mammalian expression vectors



derived from Epstein-Barr virus or bovine papilloma virus, and yeast 2-micron plasmid vectors. Stephens & Hentschel, Biochem. J., 248:1 (1987); Yates et al., Nature, 313:812 (1985); Beggs, Genetic Engineering, 2:175 (Academic Press, 1981).

5 Yet another method for obtaining high gene copy number involves gene amplification in the host cell. Gene amplification occurs naturally in eukaryotic cells at a relatively low frequency. Schimke, J. Biol. Chem., 263:5989 (1988). However, gene amplification also may be induced, or at least selected for, by exposing host cells to appropriate selective  
10 pressure. For example, in many cases it is possible to introduce a product gene together with an amplifiable gene into a host cell and subsequently select for amplification of the marker gene by exposing the cotransfected cells to sequentially increasing concentrations of a selective agent. Typically the product gene will be coamplified with the marker gene under  
15 such conditions.

The most widely used amplifiable gene for that purpose is a DHFR gene, which encodes a dihydrofolate reductase enzyme. The selection agent used in conjunction with a DHFR gene is methotrexate (Mtx). A host cell is cotransfected with a product gene encoding a desired protein and a DHFR  
20 gene, and transfectants are identified by first culturing the cells in culture medium that contains Mtx. A suitable host cell when a wild-type DHFR gene is used is the Chinese Hamster Ovary (CHO) cell line deficient in DHFR activity, prepared and propagated as described by Urlaub & Chasin, Proc. Nat. Acad. Sci. USA, 77:4216 (1980). The transfected cells then are  
25 exposed to successively higher amounts of Mtx. This leads to the synthesis of multiple copies of the DHFR gene, and concomitantly, multiple copies of the product gene. Schimke, J. Biol. Chem., 263:5989 (1988); Axel et al., U.S. Patent No. 4,399,216; Axel et al., U.S. Patent No. 4,634,665. Other references directed to co-transfection of a gene together with a genetic  
30 marker that allows for selection and subsequent amplification include Kaufman in Genetic Engineering, ed. J. Setlow (Plenum Press, New York), Vol. 9 (1987); Kaufman and Sharp, J. Mol. Biol., 159:601 (1982); Ringold et al., J. Mol. Appl. Genet., 1:165-175 (1981); Kaufman et al., Mol. Cell Biol., 5:1750-1759 (1985); Kaetzel and Nilson, J. Biol. Chem., 263:6244-  
35 6251 (1988); Hung et al., Proc. Natl. Acad. Sci. USA, 83:261-264 (1986); Kaufman et al., EMBO J., 6:87-93 (1987); Johnston and Kucey, Science, 242:1551-1554 (1988); Urlaub et al., Cell, 33:405-412 (1983).

To extend the DHFR amplification method to other cell types, a mutant DHFR gene that encodes a protein with reduced sensitivity to methotrexate  
40 may be used in conjunction with host cells that contain normal numbers of an endogenous wild-type DHFR gene. Simonsen and Levinson, Proc. Natl. Acad. Sci. USA, 80:2495 (1983); Wigler et al., Proc. Natl. Acad. Sci. USA, 77:3567-3570 (1980); Haber and Schimke, Somatic Cell Genetics, 8:499-508 (1982).

45 Alternatively, host cells may be co-transfected with the product gene, a DHFR gene, and a dominant selectable gene, such as a neo<sup>r</sup> gene. Kim

and Wold, Cell, 42:129 (1985); Capon et al., U.S. Pat. No. 4,965,199. Transfectants are identified by first culturing the cells in culture medium containing neomycin (or the related drug G418), and the transfectants so identified then are selected for amplification of the DHFR gene and the product gene by exposure to successively increasing amounts of Mtx.

As will be appreciated from this discussion, the selection of recombinant host cells that express high levels of a desired protein generally is a multi-step process. In the first step, initial transfectants are selected that have incorporated the product gene and the selectable gene. In subsequent steps, the initial transfectants are subject to further selection for high-level expression of the selectable gene and then random screening for high-level expression of the product gene. To identify cells expressing high levels of the desired protein, typically one must screen large numbers of transfectants. The majority of transfectants produce less than maximal levels of the desired protein. Further, Mtx resistance in DHFR transformants is at least partially conferred by varying degrees of gene amplification. Schimke, Cell, 37:705-713 (1984). The inadequacies of co-expression of the non-selected gene have been reported by Wold et al., Proc. Natl. Acad. Sci. USA, 76:5684-5688 (1979). Instability of the amplified DNA is reported by Kaufman and Schimke, Mol. Cell Biol., 1:1069-1076 (1981); Haber and Schimke, Cell, 26:355-362 (1981); and Fedespiel et al., J. Biol. Chem., 259:9127-9140 (1984).

Several methods have been described for directly selecting such recombinant host cells in a single step. One strategy involves co-transfecting host cells with a product gene and a DHFR gene, and selecting those cells that express high levels of DHFR by directly culturing in medium containing a high concentration of Mtx. Many of the cells selected in that manner also express the co-transfected product gene at high levels. Page and Sydenham, Bio/Technology, 9:64 (1991). This method for single-step selection suffers from certain drawbacks that limit its usefulness. High-expressing cells obtained by direct culturing in medium containing a high level of a selection agent may have poor growth and stability characteristics, thus limiting their usefulness for long-term production processes. Page and Snyderman, Bio/Technology, 9:64 (1991). Single-step selection for high-level resistance to Mtx may produce cells with an altered, Mtx-resistant DHFR enzyme, or cells that have altered Mtx transport properties, rather than cells containing amplified genes. Haber et al., J. Biol. Chem., 256:9501 (1981); Assaraf and Schimke, Proc. Natl. Acad. Sci. USA, 84:7154 (1987).

Another method involves the use of polycistronic mRNA expression vectors containing a product gene at the 5' end of the transcribed region and a selectable gene at the 3' end. Because translation of the selectable gene at the 3' end of the polycistronic mRNA is inefficient, such vectors exhibit preferential translation of the product gene and require high levels of polycistronic mRNA to survive selection. Kaufman, Meth.

Enzymol., 185:487 (1990); Kaufman, Meth. Enzymol., 185:537 (1990); Kaufman et al., EMBO J., 6:187 (1987). Accordingly, cells expressing high levels of the desired protein product may be obtained in a single step by culturing the initial transfectants in medium containing a selection agent appropriate for use with the particular selectable gene. However, the utility of these vectors is variable because of the unpredictable influence of the upstream product reading frame on selectable marker translation and because the upstream reading frame sometimes becomes deleted during methotrexate amplification (Kaufman et al., J. Mol. Biol., 159:601-621 [1982]; Levinson, Methods in Enzymology, San Diego: Academic Press, Inc. [1990]). Later vectors incorporated an internal translation initiation site derived from members of the picornavirus family which is positioned between the product gene and the selectable gene (Pelletier et al., Nature, 334:320 [1988]; Jang et al., J. Virol., 63:1651 [1989]).

A third method for single-step selection involves use of a DNA construct with a selectable gene containing an intron within which is located a gene encoding the protein of interest. See U.S. Patent No. 5,043,270 and Abrams et al., J. Biol. Chem., 264(24): 14016-14021 (1989). In yet another single-step selection method, host cells are co-transfected with an intron-modified selectable gene and a gene encoding the protein of interest. See WO 92/17566, published October 15, 1992. The intron-modified gene is prepared by inserting into the transcribed region of a selectable gene an intron of such length that the intron is correctly spliced from the corresponding mRNA precursor at low efficiency, so that the amount of selectable marker produced from the intron-modified selectable gene is substantially less than that produced from the starting selectable gene. These vectors help to insure the integrity of the integrated DNA construct, but transcriptional linkage is not achieved as selectable gene and the protein gene are driven by separate promoters.

Other mammalian expression vectors that have single transcription units have been described. Retroviral vectors have been constructed (Cepko et al., Cell, 37:1053-1062 [1984]) in which a cDNA is inserted between the endogenous Moloney murine leukemia virus (M-MuLV) splice donor and splice acceptor sites which are followed by a neomycin resistance gene. This vector has been used to express a variety of gene products following retroviral infection of several cell types.

With the above drawbacks in mind, it is one object of the present invention to increase the level of homogeneity with regard to expression levels of stable clones transfected with a product gene of interest, by expressing a selectable marker (DHFR) and the protein of interest from a single promoter.

It is another object to provide a method for selecting stable, recombinant host cells that express high levels of a desired protein product, which method is rapid and convenient to perform, and reduces the numbers of transfected cells which need to be screened. Furthermore, it is

an object to allow high levels of single and two unit polypeptides to be rapidly generated from clones or pools of stable host cell transfectants.

It is an additional object to provide expression vectors which bias for active integration events (i.e. have an increased tendency to generate transformants wherein the DNA construct is inserted into a region of the genome of the host cell which results in high level expression of the product gene) and can accommodate a variety of product genes without the need for modification.

10

#### SUMMARY OF THE INVENTION

Accordingly, the present invention is directed to a DNA construct (DNA molecule) alternative terminology comprising a 5' transcriptional initiation site and a 3' transcriptional termination site, a selectable gene (preferably an amplifiable gene) and a product gene provided 3' to the selectable gene, a transcriptional regulatory region regulating transcription of both the selectable gene and the product gene, the selectable gene positioned within an intron defined by a splice donor site and a splice acceptor site. The splice donor site preferably comprises an effective splice donor sequence as herein defined and thereby regulates expression of the product gene using the transcriptional regulatory region.

In another embodiment, the invention provides a method for producing a product of interest comprising culturing a eukaryotic cell which has been transfected with the DNA construct described above, so as to express the product gene and recovering the product.

In a further embodiment, the invention provides a method for producing eukaryotic cells having multiple copies of the product gene comprising transfecting eukaryotic cells with the DNA construct described above (where the selectable gene is an amplifiable gene), growing the cells in a selective medium comprising an amplifying agent for a sufficient time for amplification to occur, and selecting cells having multiple copies of the product gene. Preferably transfection of the cells is achieved using electroporation.

After transfection of the host cells, most of the transfectants fail to exhibit the selectable phenotype characteristic of the protein encoded by the selectable gene, but surprisingly a small proportion of the transfectants do exhibit the selectable phenotype, and among those transfectants, the majority are found to express high levels of the desired product encoded by the product gene. Thus, the invention provides an improved method for the selection of recombinant host cells expressing high levels of a desired product, which method is useful with a wide variety of eukaryotic host cells and avoids the problems inherent in existing cell selection technology.

### BRIEF DESCRIPTION OF THE DRAWINGS

Figures 1A-1D illustrate schematically various DNA constructs encompassed by the instant invention. The large arrows represent the selectable gene and the product gene, the V formed by the dashed lines shows the region of the precursor RNA internal to the 5' splice donor site (SD) and 3' splice acceptor site (SA) that is excised from vectors that contain a functional SD. The transcriptional regulatory region, selectable gene, product gene and transcriptional termination site are depicted in Figure 1A. Figure 1B depicts the DNA constructs of Example 1. The various splice donor sequences are depicted, i.e., wild type ras splice donor sequence (WT ras), mutant ras splice donor sequence (MUTANT ras) and non-functional splice donor sequence ( $\Delta$ GT). The probes used for Northern blot analysis in Example 1 are shown in Figure 1B. Figure 1C depicts the DNA constructs of Example 2 and Figure 1D depicts the DNA construct of Example 3 used for expression of anti-IgE V<sub>H</sub>.

Figure 2 depicts schematically the control DNA construct used in Example 1.

Figures 3A-Q depict the nucleotide sequence (SEQ ID NO: 1) of the DHFR/intron- (WT ras SD)-tPA expression vector of Example 1.

Figure 4 is a bar graph which shows the number of colonies that form in selective medium after electroporation of linearized duplicate miniprep DNA's prepared in parallel from the three vectors shown in Figure 1B (i.e. with wild type ras splice donor sequence [WT ras], mutant ras splice donor sequence [MUTANT ras] and non-functional splice donor sequence [ $\Delta$ GT]) and from the control vector that has DHFR under control of SV40 promoter and tPA under control of CMV promoter (see Figure 2). Cells were selected in nucleoside free medium and counted with an automated colony counter.

Figures 5A-C are bar graphs depicting expression of tPA from stable pools and clones generated from the vectors shown in Figure 1B. In Figure 5A greater than 100 clones from each vector transfection were mixed, plated in 24 well plates, and assayed by tPA ELISA at "saturation". In Figure 5B, twenty clones chosen at random derived from each of the vectors were assayed by tPA ELISA at "saturation". In Figure 5C, the pools mentioned in Figure 5A (except the  $\Delta$ GT pool) were exposed to 200nM Mtx to select for DHFR amplification and then pooled and assayed for tPA expression.

Figures 6A-P depict the nucleotide sequence (SEQ ID NO: 2) of the DHFR/intron- (WT ras SD)-TNFr-IgG expression vector of Example 2.

Figures 7A-B are bar graphs depicting expression of TNFr-IgG using dicistronic or control vectors (see Example 2). Vectors containing TNFr-IgG (but otherwise identical to those described for tPA expression in Example 1) were constructed (see Figure 1C), introduced into dp12.CHO cells by electroporation, pooled, and assayed for product expression before (Figure 7A) and after (Figure 7B) being subjected to amplification in 200nM Mtx.

Figure 8 depicts schematically the DNA construct used for expression of the V<sub>L</sub> of anti-IgE in Example 3.

Figures 9A-O depict the nucleotide sequence (SEQ ID NO: 3) of the anti-IgE V<sub>H</sub> expression vector of Example 3.

Figures 10A-Q depict the nucleotide sequence (SEQ ID NO: 4) of the anti-IgE V<sub>L</sub> expression vector of Example 3.

5 Figure 11 is a bar graph depicting anti-IgE expression in Example 3. Heavy (V<sub>H</sub>) and light (V<sub>L</sub>) chain expression vectors were constructed, co-electroporated into CHO cells, clones were selected and assayed for antibody expression. Additionally, pools were established and assessed with regard to expression before and after Mtx selection at 200nM and 1μM.

#### 10 DESCRIPTION OF THE PREFERRED EMBODIMENTS

##### Definitions:

The "DNA construct" disclosed herein comprises a non-naturally occurring DNA molecule which can either be provided as an isolate or integrated in another DNA molecule e.g. in an expression vector or the  
15 chromosome of an eukaryotic host cell.

The term "selectable gene" as used herein refers to a DNA that encodes a selectable marker necessary for the growth or survival of a host cell under the particular cell culture conditions chosen. Accordingly, a host cell that is transformed with a selectable gene will be capable of  
20 growth or survival under certain cell culture conditions wherein a non-transfected host cell is not capable of growth or survival. Typically, a selectable gene will confer resistance to a drug or compensate for a metabolic or catabolic defect in the host cell. Examples of selectable genes are provided in the following table. See also Kaufman, Methods in  
25 Enzymology, 185: 537-566 (1990), for a review of these.

**TABLE 1**  
**Selectable Genes and their Selection Agents**

Selection Agent	Selectable Gene
Methotrexate	Dihydrofolate reductase
Cadmium	Metallothionein
PALA	CAD
Xyl-A-or adenosine and 2'- deoxycoformycin	Adenosine deaminase
Adenine, azaserine, and coformycin	Adenylate deaminase
6-Azaauridine, pyrazofuran	UMP Synthetase
Mycophenolic acid	IMP 5'-dehydrogenase

	Mycophenolic acid with limiting xanthine	Xanthine-guanine phosphoribosyltransferase
	Hypoxanthine, aminopterin, and thymidine (HAT)	Mutant HGPRTase or mutant thymidine kinase
5	5-Fluorodeoxyuridine	Thymidylate synthetase
	Multiple drugs e.g. adriamycin, vincristine or colchicine	P-glycoprotein 170
	Aphidicolin	Ribonucleotide reductase
10	Methionine sulfoximine	Glutamine synthetase
	$\beta$ -Aspartyl hydroxamate or Albizziin	Asparagine synthetase
	Canavanine	Arginosuccinate synthetase
	$\alpha$ -Difluoromethylornithine	Ornithine decarboxylase
15	Compactin	HMG-CoA reductase
	Tunicamycin	N-Acetylglucosaminyl transferase
	Borrelidin	Threonyl-tRNA synthetase
	Ouabain	Na <sup>+</sup> K <sup>+</sup> -ATPase

The preferred selectable gene is an amplifiable gene. As used herein, the term "amplifiable gene" refers to a gene which is amplified (i.e. additional copies of the gene are generated which survive in intrachromosomal or extrachromosomal form) under certain conditions. The amplifiable gene usually encodes an enzyme (i.e. an amplifiable marker) which is required for growth of eukaryotic cells under those conditions. For example, the gene may encode DHFR which is amplified when a host cell transformed therewith is grown in Mtx. According to Kaufman, the selectable genes in Table 1 above can also be considered amplifiable genes. An example of a selectable gene which is generally not considered to be an amplifiable gene is the neomycin resistance gene (Cepko et al., supra).

As used herein, "selective medium" refers to nutrient solution used for growing eukaryotic cells which have the selectable gene and therefore includes a "selection agent". Commercially available media such as Ham's F10 (Sigma), Minimal Essential Medium ([MEM], Sigma), RPMI-1640 (Sigma), and Dulbecco's Modified Eagle's Medium ([DMEM], Sigma) are exemplary nutrient solutions. In addition, any of the media described in Ham and Wallace, Meth. Enz., 58:44 (1979), Barnes and Sato, Anal. Biochem., 102:255

(1980), U.S. Patent Nos. 4,767,704; 4,657,866; 4,927,762; or 4,560,655; WO 90/03430; WO 87/00195; U.S. Patent Re. 30,985; or U.S. Patent No. 5,122,469, the disclosures of all of which are incorporated herein by reference, may be used as culture media. Any of these media may be  
5 supplemented as necessary with hormones and/or other growth factors (such as insulin, transferrin, or epidermal growth factor), salts (such as sodium chloride, calcium, magnesium, and phosphate), buffers (such as HEPES), nucleosides (such as adenosine and thymidine), antibiotics (such as Gentamycin™ drug), trace elements (defined as inorganic compounds usually  
10 present at final concentrations in the micromolar range), and glucose or an equivalent energy source. Any other necessary supplements may also be included at appropriate concentrations that would be known to those skilled in the art. The preferred nutrient solution comprises fetal bovine serum.

The term "selection agent" refers to a substance that interferes with  
15 the growth or survival of a host cell that is deficient in a particular selectable gene. Examples of selection agents are presented in Table 1 above. The selection agent preferably comprises an "amplifying agent" which is defined for purposes herein as an agent for amplifying copies of the amplifiable gene, such as Mtx if the amplifiable gene is DHFR. See Table  
20 1 for examples of amplifying agents.

As used herein, the term "transcriptional initiation site" refers to the nucleic acid in the DNA construct corresponding to the first nucleic acid incorporated into the primary transcript, i.e., the mRNA precursor, which site is generally provided at, or adjacent to, the 5' end of the DNA  
25 construct.

The term "transcriptional termination site" refers to a sequence of DNA, normally represented at the 3' end of the DNA construct, that causes RNA polymerase to terminate transcription.

As used herein, "transcriptional regulatory region" refers to a  
30 region of the DNA construct that regulates transcription of the selectable gene and the product gene. The transcriptional regulatory region normally refers to a promoter sequence (i.e. a region of DNA involved in binding of RNA polymerase to initiate transcription) which can be constitutive or inducible and, optionally, an enhancer (i.e. a *cis*-acting DNA element,  
35 usually from about 10-300 bp, that acts on a promoter to increase its transcription).

As used herein, "product gene" refers to DNA that encodes a desired protein or polypeptide product. Any product gene that is capable of expression in a host cell may be used, although the methods of the  
40 invention are particularly suited for obtaining high-level expression of a product gene that is not also a selectable or amplifiable gene. Accordingly, the protein or polypeptide encoded by a product gene typically will be one that is not necessary for the growth or survival of a host cell under the particular cell culture conditions chosen. For example, product  
45 genes suitably encode a peptide, or may encode a polypeptide sequence of



amino acids for which the chain length is sufficient to produce higher levels of tertiary and/or quaternary structure.

Examples of bacterial polypeptides or proteins include, e.g., alkaline phosphatase and  $\beta$ -lactamase. Examples of mammalian polypeptides or proteins include molecules such as renin; a growth hormone, including human growth hormone, and bovine growth hormone; growth hormone releasing factor; parathyroid hormone; thyroid stimulating hormone; lipoproteins; alpha-1-antitrypsin; insulin A-chain; insulin B-chain; proinsulin; follicle stimulating hormone; calcitonin; luteinizing hormone; glucagon; clotting factors such as factor VIIIC, factor IX, tissue factor, and von Willebrands factor; anti-clotting factors such as Protein C; atrial natriuretic factor; lung surfactant; a plasminogen activator, such as urokinase or human urine or tissue-type plasminogen activator (t-PA); bombesin; thrombin; hemopoietic growth factor; tumor necrosis factor-alpha and -beta; enkephalinase; RANTES (regulated on activation normally T-cell expressed and secreted); human macrophage inflammatory protein (MIP-1-alpha); a serum albumin such as human serum albumin; mullerian-inhibiting substance; relaxin A-chain; relaxin B-chain; prorelaxin; mouse gonadotropin-associated peptide; a microbial protein, such as beta-lactamase; DNase; inhibin; activin; vascular endothelial growth factor (VEGF); receptors for hormones or growth factors; integrin; protein A or D; rheumatoid factors; a neurotrophic factor such as bone-derived neurotrophic factor (BDNF), neurotrophin-3, -4, -5, or -6 (NT-3, NT-4, NT-5, or NT-6), or a nerve growth factor such as NGF- $\beta$ ; platelet-derived growth factor (PDGF); fibroblast growth factor such as aFGF and bFGF; epidermal growth factor (EGF); transforming growth factor (TGF) such as TGF-alpha and TGF-beta, including TGF- $\beta$ 1, TGF- $\beta$ 2, TGF- $\beta$ 3, TGF- $\beta$ 4, or TGF- $\beta$ 5; insulin-like growth factor-I and -II (IGF-I and IGF-II); des(1-3)-IGF-I (brain IGF-I), insulin-like growth factor binding proteins; CD proteins such as CD-3, CD-4, CD-8, and CD-19; erythropoietin; osteoinductive factors; immunotoxins; a bone morphogenetic protein (BMP); an interferon such as interferon-alpha, -beta, and -gamma; colony stimulating factors (CSFs), e.g., M-CSF, GM-CSF, and G-CSF; interleukins (ILs), e.g., IL-1 to IL-10; superoxide dismutase; T-cell receptors; surface membrane proteins; decay accelerating factor; viral antigen such as, for example, a portion of the AIDS envelope; transport proteins; homing receptors; addressins; regulatory proteins; antibodies; chimeric proteins such as immunoadhesins and fragments of any of the above-listed polypeptides.

The product gene preferably does not consist of an anti-sense sequence for inhibiting the expression of a gene present in the host. Preferred proteins herein are therapeutic proteins such as TGF- $\beta$ , TGF- $\alpha$ , PDGF, EGF, FGF, IGF-I, DNase, plasminogen activators such as t-PA, clotting factors such as tissue factor and factor VIII, hormones such as relaxin and insulin, cytokines such as IFN- $\gamma$ , chimeric proteins such as TNF receptor IgG immunoadhesin (TNFr-IgG) or antibodies such as anti-IgE.

The term "intron" as used herein refers to a nucleotide sequence present within the transcribed region of a gene or within a messenger RNA precursor, which nucleotide sequence is capable of being excised, or spliced, from the messenger RNA precursor by a host cell prior to translation. Introns suitable for use in the present invention are suitably prepared by any of several methods that are well known in the art, such as purification from a naturally occurring nucleic acid or de novo synthesis. The introns present in many naturally occurring eukaryotic genes have been identified and characterized. Mount, Nuc. Acids Res., 10:459 (1982). Artificial introns comprising functional splice sites also have been described. Winey et al., Mol. Cell Biol., 9:329 (1989); Gattermann et al., Mol. Cell Biol., 9:1526 (1989). Introns may be obtained from naturally occurring nucleic acids, for example, by digestion of a naturally occurring nucleic acid with a suitable restriction endonuclease, or by PCR cloning using primers complementary to sequences at the 5' and 3' ends of the intron. Alternatively, introns of defined sequence and length may be prepared synthetically using various methods in organic chemistry. Narang et al., Meth. Enzymol., 68:90 (1979); Caruthers et al., Meth. Enzymol., 154:287 (1985); Froehler et al., Nuc. Acids Res., 14:5399 (1986).

As used herein "splice donor site" or "SD" refers to the DNA sequence immediately surrounding the exon-intron boundary at the 5' end of the intron, where the "exon" comprises the nucleic acid 5' to the intron. Many splice donor sites have been characterized and Ohshima et al., J. Mol. Biol., 195:247-259 (1987) provides a review of these. An "efficient splice donor sequence" refers to a nucleic acid sequence encoding a splice donor site wherein the efficiency of splicing of messenger RNA precursors having the splice donor sequence is between about 80 to 99% and preferably 90 to 95% as determined by quantitative PCR. Examples of efficient splice donor sequences include the wild type (WT) ras splice donor sequence and the GAC:GTAAGT sequence of Example 3. Other efficient splice donor sequences can be readily selected using the techniques for measuring the efficiency of splicing disclosed herein.

The terms "PCR" and "polymerase chain reaction" as used herein refer to the *in vitro* amplification method described in US Patent No. 4,683,195 (issued July 28, 1987). In general, the PCR method involves repeated cycles of primer extension synthesis, using two DNA primers capable of hybridizing preferentially to a template nucleic acid comprising the nucleotide sequence to be amplified. The PCR method can be used to clone specific DNA sequences from total genomic DNA, cDNA transcribed from cellular RNA, viral or plasmid DNAs. Wang & Mark, in PCR Protocols, pp. 70-75 (Academic Press, 1990); Scharf, in PCR Protocols, pp. 84-98; Kawasaki & Wang, in PCR Technology, pp. 89-97 (Stockton Press, 1989). Reverse transcription-polymerase chain reaction (RT-PCR) can be used to analyze RNA samples containing mixtures of spliced and unspliced mRNA transcripts. Fluorescently tagged primers designed to span the intron are used to

amplify both spliced and unspliced targets. The resultant amplification products are then separated by gel electrophoresis and quantitated by measuring the fluorescent emission of the appropriate band(s). A comparison is made to determine the amount of spliced and unspliced transcripts present in the RNA sample.

One preferred splice donor sequence is a "consensus splice donor sequence". The nucleotide sequences surrounding intron splice sites, which sequences are evolutionarily highly conserved, are referred to as "consensus splice donor sequences". In the mRNAs of higher eukaryotes, the 5' splice site occurs within the consensus sequence AG:GUAAGU (wherein the colon denotes the site of cleavage and ligation). In the mRNAs of yeast, the 5' splice site is bounded by the consensus sequence :GUAUGU. Padgett, et al., Ann. Rev. Biochem., 55:1119 (1986).

The expression "splice acceptor site" or "SA" refers to the sequence immediately surrounding the intron-exon boundary at the 3' end of the intron, where the "exon" comprises the nucleic acid 3' to the intron. Many splice acceptor sites have been characterized and Ohshima et al., J. Mol. Biol., 195:247-259 (1987) provides a review of these. The preferred splice acceptor site is an efficient splice acceptor site which refers to a nucleic acid sequence encoding a splice acceptor site wherein the efficiency of splicing of messenger RNA precursors having the splice acceptor site is between about 80 to 99% and preferably 90 to 95% as determined by quantitative PCR. The splice acceptor site may comprise a consensus sequence. In the mRNAs of higher eukaryotes, the 3' splice acceptor site occurs within the consensus sequence (U/C)<sub>11</sub>NCAG:G. In the mRNAs of yeast, the 3' acceptor splice site is bounded by the consensus sequence (C/U)AG:. Padgett, et al., *supra*.

As used herein "culturing for sufficient time to allow amplification to occur" refers to the act of physically culturing the eukaryotic host cells which have been transformed with the DNA construct in cell culture media containing the amplifying agent, until the copy number of the amplifiable gene (and preferably also the copy number of the product gene) in the host cells has increased relative to the transformed cells prior to this culturing.

The term "expression" as used herein refers to transcription or translation occurring within a host cell. The level of expression of a product gene in a host cell may be determined on the basis of either the amount of corresponding mRNA that is present in the cell or the amount of the protein encoded by the product gene that is produced by the cell. For example, mRNA transcribed from a product gene is desirably quantitated by northern hybridization. Sambrook, et al., Molecular Cloning: A Laboratory Manual, pp. 7.3-7.57 (Cold Spring Harbor Laboratory Press, 1989). Protein encoded by a product gene can be quantitated either by assaying for the biological activity of the protein or by employing assays that are independent of such activity, such as western blotting or radioimmunoassay using antibodies that are capable of reacting with the protein. Sambrook,

et al., Molecular Cloning: A Laboratory Manual, pp. 18.1-18.88 (Cold Spring Harbor Laboratory Press, 1989).

#### Modes for Carrying Out the Invention

Methods and compositions are provided for enhancing the stability and/or copy number of a transcribed sequence in order to allow for elevated levels of a RNA sequence of interest. In general, the methods of the present invention involve transfecting a eukaryotic host cell with an expression vector comprising both a product gene encoding a desired polypeptide and a selectable gene (preferably an amplifiable gene).

Selectable genes and product genes may be obtained from genomic DNA, cDNA transcribed from cellular RNA, or by in vitro synthesis. For example, libraries are screened with probes (such as antibodies or oligonucleotides of about 20-80 bases) designed to identify the selectable gene or the product gene (or the protein(s) encoded thereby). Screening the cDNA or genomic library with the selected probe may be conducted using standard procedures as described in chapters 10-12 of Sambrook et al., Molecular Cloning: A Laboratory Manual (New York: Cold Spring Harbor Laboratory Press, 1989). An alternative means to isolate the selectable gene or product gene is to use PCR methodology as described in section 14 of Sambrook et al., *supra*.

A preferred method of practicing this invention is to use carefully selected oligonucleotide sequences to screen cDNA libraries from various tissues known to contain the selectable gene or product gene. The oligonucleotide sequences selected as probes should be of sufficient length and sufficiently unambiguous that false positives are minimized.

The oligonucleotide generally is labeled such that it can be detected upon hybridization to DNA in the library being screened. The preferred method of labeling is to use <sup>32</sup>P- labeled ATP with polynucleotide kinase, as is well known in the art, to radiolabel the oligonucleotide. However, other methods may be used to label the oligonucleotide, including, but not limited to, biotinylation or enzyme labeling.

Sometimes, the DNA encoding the selectable gene and product gene is preceded by DNA encoding a signal sequence having a specific cleavage site at the N-terminus of the mature protein or polypeptide. In general, the signal sequence may be a component of the expression vector, or it may be a part of the selectable gene or product gene that is inserted into the expression vector. If a heterologous signal sequence is used, it preferably is one that is recognized and processed (i.e., cleaved by a signal peptidase) by the host cell. For yeast secretion the native signal sequence may be substituted by, e.g., the yeast invertase leader, alpha factor leader (including *Saccharomyces* and *Kluyveromyces*  $\alpha$ -factor leaders, the latter described in U.S. Pat. No. 5,010,182 issued 23 April 1991), or acid phosphatase leader, the *C. albicans* glucoamylase leader (EP 362,179 published 4 April 1990), or the signal described in WO 90/13646 published 15 November 1990. In mammalian cell expression the native signal sequence

of the protein of interest is satisfactory, although other mammalian signal sequences may be suitable, such as signal sequences from secreted polypeptides of the same or related species, as well as viral secretory leaders, for example, the herpes simplex gD signal. The DNA for such precursor region is ligated in reading frame to the selectable gene or product gene.

As shown in Figure 1A, the selectable gene is generally provided at the 5' end of the DNA construct and this selectable gene is followed by the product gene. Therefore, the full length (non-spliced) message will contain DHFR as the first open reading frame and will therefore generate DHFR protein to allow selection of stable transfectants. The full length message is not expected to generate appreciable amounts of the protein of interest as the second AUG in a dicistronic message is an inefficient initiator of translation in mammalian cells (Kozak, J. Cell Biol., 115: 887-903 [1991]).

The selectable gene is positioned within an intron. Introns are noncoding nucleotide sequences, normally present within many eukaryotic genes, which are removed from newly transcribed mRNA precursors in a multiple-step process collectively referred to as splicing.

A single mechanism is thought to be responsible for the splicing of mRNA precursors in mammalian, plant, and yeast cells. In general, the process of splicing requires that the 5' and 3' ends of the intron be correctly cleaved and the resulting ends of the mRNA be accurately joined, such that a mature mRNA having the proper reading frame for protein synthesis is produced. Analysis of a variety of naturally occurring and synthetically constructed mutant genes has shown that nucleotide changes at many of the positions within the consensus sequences at the 5' and 3' splice sites have the effect of reducing or abolishing the synthesis of mature mRNA. Sharp, Science, 235:766 (1987); Padgett, et al., Ann. Rev. Biochem., 55:1119 (1986); Green, Ann. Rev. Genet., 20:671 (1986). Mutational studies also have shown that RNA secondary structures involving splicing sites can affect the efficiency of splicing. Solnick, Cell, 43:667 (1985); Konarska, et al., Cell, 42:165 (1985).

The length of the intron may also affect the efficiency of splicing. By making deletion mutations of different sizes within the large intron of the rabbit beta-globin gene, Wieringa, et al. determined that the minimum intron length necessary for correct splicing is about 69 nucleotides. Cell, 37:915 (1984). Similar studies of the intron of the adenovirus E1A region have shown that an intron length of about 78 nucleotides allows correct splicing to occur, but at reduced efficiency. Increasing the length of the intron to 91 nucleotides restores normal splicing efficiency, whereas truncating the intron to 63 nucleotides abolishes correct splicing. Ulfendahl, et al., Nuc. Acids Res., 13:6299 (1985).

To be useful in the invention, the intron must have a length such that splicing of the intron from the mRNA is efficient. The preparation of introns of differing lengths is a routine matter, involving methods well known in the art, such as de novo synthesis or in vitro deletion

mutagenesis of an existing intron. Typically, the intron will have a length of at least about 150 nucleotides, since introns which are shorter than this tend to be spliced less efficiently. The upper limit for the length of the intron can be up to 30 kB or more. However, as a general proposition, the intron is generally less than about 10 kB in length.

The intron is modified to contain the selectable gene not normally present within the intron using any of the various known methods for modifying a nucleic acid *in vitro*. Typically, a selectable gene will be introduced into an intron by first cleaving the intron with a restriction endonuclease, and then covalently joining the resulting restriction fragments to the selectable gene in the correct orientation for host cell expression, for example by ligation with a DNA ligase enzyme.

The DNA construct is dicistronic, i.e. the selectable gene and product gene are both under the transcriptional control of a single transcriptional regulatory region. As mentioned above, the transcriptional regulatory region comprises a promoter. Suitable promoting sequences for use with yeast hosts include the promoters for 3-phosphoglycerate kinase (Hitzeman et al., J. Biol. Chem., 255:2073 [1980]) or other glycolytic enzymes (Hess et al., J. Adv. Enzyme Reg., 7:149 [1968]; and Holland, Biochemistry, 17:4900 [1978]), such as enolase, glyceraldehyde-3-phosphate dehydrogenase, hexokinase, pyruvate decarboxylase, phosphofructokinase, glucose-6-phosphate isomerase, 3-phosphoglycerate mutase, pyruvate kinase, triosephosphate isomerase, phosphoglucose isomerase, and glucokinase.

Other yeast promoters, which are inducible promoters having the additional advantage of transcription controlled by growth conditions, are the promoter regions for alcohol dehydrogenase 2, isocytochrome C, acid phosphatase, degradative enzymes associated with nitrogen metabolism, metallothionein, glyceraldehyde-3-phosphate dehydrogenase, and enzymes responsible for maltose and galactose utilization. Suitable vectors and promoters for use in yeast expression are further described in Hitzeman et al., EP 73,657A. Yeast enhancers also are advantageously used with yeast promoters.

Expression control sequences are known for eukaryotes. Virtually all eukaryotic genes have an AT-rich region located approximately 25 to 30 bases upstream from the site where transcription is initiated. Another sequence found 70 to 80 bases upstream from the start of transcription of many genes is a CXCAAT region where X may be any nucleotide.

Product gene transcription from vectors in mammalian host cells is controlled by promoters obtained from the genomes of viruses such as polyoma virus, fowlpox virus (UK 2,211,504 published 5 July 1989), adenovirus (such as Adenovirus 2), bovine papilloma virus, avian sarcoma virus, cytomegalovirus, a retrovirus, hepatitis-B virus and most preferably Simian Virus 40 (SV40), from heterologous mammalian promoters, e.g. the actin promoter or an immunoglobulin promoter, from heat-shock promoters, and from the promoter normally associated with the product gene, provided such promoters are compatible with the host cell systems.

The early and late promoters of the SV40 virus are conveniently obtained as an SV40 restriction fragment that also contains the SV40 viral origin of replication. Fiers et al., Nature, 273:113 (1978); Mulligan and Berg, Science, 209:1422-1427 (1980); Pavlakis et al., Proc. Natl. Acad. Sci. USA, 78:7398-7402 (1981). The immediate early promoter of the human cytomegalovirus (CMV) is conveniently obtained as a *HindIII* E restriction fragment. Greenaway et al., Gene, 18:355-360 (1982). A system for expressing DNA in mammalian hosts using the bovine papilloma virus as a vector is disclosed in U.S. 4,419,446. A modification of this system is described in U.S. 4,601,978. See also Gray et al., Nature, 295:503-508 (1982) on expressing cDNA encoding immune interferon in monkey cells; , Reyes et al., Nature, 297:598-601 (1982) on expression of human  $\beta$ -interferon cDNA in mouse cells under the control of a thymidine kinase promoter from herpes simplex virus, Canaani and Berg, Proc. Natl. Acad. Sci. USA, 79:5166-5170 (1982) on expression of the human interferon  $\beta$ 1 gene in cultured mouse and rabbit cells, and Gorman et al., Proc. Natl. Acad. Sci. USA, 79:6777-6781 (1982) on expression of bacterial CAT sequences in CV-1 monkey kidney cells, chicken embryo fibroblasts, Chinese hamster ovary cells, HeLa cells, and mouse NIH-3T3 cells using the Rous sarcoma virus long terminal repeat as a promoter.

Preferably the transcriptional regulatory region in higher eukaryotes comprises an enhancer sequence. Enhancers are relatively orientation and position independent having been found 5' (Lainins et al., Proc. Natl. Acad. Sci. USA, 78:993 [1981]) and 3' (Lusky et al., Mol. Cell Bio., 3:1108 [1983]) to the transcription unit, within an intron (Banerji et al., Cell, 33:729 [1983]) as well as within the coding sequence itself (Osborne et al., Mol. Cell Bio., 4:1293 [1984]). Many enhancer sequences are now known from mammalian genes (globin, elastase, albumin,  $\alpha$ -fetoprotein and insulin). Typically, however, one will use an enhancer from a eukaryotic cell virus. Examples include the SV40 enhancer on the late side of the replication origin (bp 100-270), the cytomegalovirus early promoter enhancer (CMV), the polyoma enhancer on the late side of the replication origin, and adenovirus enhancers. See also Yaniv, Nature, 297:17-18 (1982) on enhancing elements for activation of eukaryotic promoters. The enhancer may be spliced into the vector at a position 5' or 3' to the product gene, but is preferably located at a site 5' from the promoter.

The DNA construct has a transcriptional initiation site following the transcriptional regulatory region and a transcriptional termination region following the product gene (see Figure 1A). These sequences are provided in the DNA construct using techniques which are well known in the art.

The DNA construct normally forms part of an expression vector which may have other components such as an origin of replication (i.e., a nucleic acid sequence that enables the vector to replicate in one or more selected host cells) and, if desired, one or more additional selectable gene(s). Construction of suitable vectors containing the desired coding and control sequences employs standard ligation techniques. Isolated plasmids or DNA

fragments are cleaved, tailored, and religated in the form desired to generate the plasmids required.

Generally, in cloning vectors the origin of replication is one that enables the vector to replicate independently of the host chromosomal DNA, and includes origins of replication or autonomously replicating sequences. Such sequences are well known. The 2 $\mu$  plasmid origin of replication is suitable for yeast, and various viral origins (SV40, polyoma, adenovirus, VSV or BPV) are useful for cloning vectors in mammalian cells. Generally, the origin of replication component is not needed for mammalian expression vectors (the SV40 origin may typically be used only because it contains the early promoter).

Most expression vectors are "shuttle" vectors, i.e., they are capable of replication in at least one class of organisms but can be transfected into another organism for expression. For example, a vector is cloned in *E. coli* and then the same vector is transfected into yeast or mammalian cells for expression even though it is not capable of replicating independently of the host cell chromosome.

For analysis to confirm correct sequences in plasmids constructed, plasmids from the transformants are prepared, analyzed by restriction, and/or sequenced by the method of Messing et al., Nucleic Acids Res., 9:309 (1981) or by the method of Maxam et al., Methods in Enzymology, 65:499 (1980).

The expression vector having the DNA construct prepared as discussed above is transformed into a eukaryotic host cell. Suitable host cells for cloning or expressing the vectors herein are yeast or higher eukaryote cells.

Eukaryotic microbes such as filamentous fungi or yeast are suitable hosts for vectors containing the product gene. *Saccharomyces cerevisiae*, or common baker's yeast, is the most commonly used among lower eukaryotic host microorganisms. However, a number of other genera, species, and strains are commonly available and useful herein, such as *S. pombe* [Beach and Nurse, Nature, 290:140 (1981)], *Kluyveromyces lactis* [Louvencourt et al., J. Bacteriol., 737 (1983)], *Yarrowia* [EP 402,226], *Pichia pastoris* [EP 183,070], *Trichoderma reesia* [EP 244,234], *Neurospora crassa* [Case et al., Proc. Natl. Acad. Sci. USA, 76:5259-5263 (1979)], and *Aspergillus* hosts such as *A. nidulans* [Ballance et al., Biochem. Biophys. Res. Commun., 112:284-289 (1983); Tilburn et al., Gene, 26:205-221 (1983); Yelton et al., Proc. Natl. Acad. Sci. USA, 81:1470-1474 (1984)] and *A. niger* [Kelly and Hynes, EMBO J., 4:475-479 (1985)].

Suitable host cells for the expression of the product gene are derived from multicellular organisms. Such host cells are capable of complex processing and glycosylation activities. In principle, any higher eukaryotic cell culture is workable, whether from vertebrate or invertebrate culture. Examples of invertebrate cells include plant and insect cells. Numerous baculoviral strains and variants and corresponding permissive insect host cells from hosts such as *Spodoptera frugiperda*



(caterpillar), *Aedes aegypti* (mosquito), *Aedes albopictus* (mosquito), *Drosophila melanogaster* (fruitfly), and *Bombyx mori* host cells have been identified. See, e.g., Luckow et al., Bio/Technology, 6:47-55 (1988); Miller et al., in Genetic Engineering, Setlow, J.K. et al., eds., Vol. 8  
5 (Plenum Publishing, 1986), pp. 277-279; and Maeda et al., Nature, 315:592-594 (1985). A variety of such viral strains are publicly available, e.g., the L-1 variant of *Autographa californica* NPV and the Bm-5 strain of *Bombyx mori* NPV, and such viruses may be used as the virus herein according to the present invention, particularly for transfection of *Spodoptera frugiperda*  
10 cells.

Plant cell cultures of cotton, corn, potato, soybean, petunia, tomato, and tobacco can be utilized as hosts. Typically, plant cells are transfected by incubation with certain strains of the bacterium *Agrobacterium tumefaciens*, which has been previously manipulated to contain  
15 the product gene. During incubation of the plant cell culture with *A. tumefaciens*, the product gene is transferred to the plant cell host such that it is transfected, and will, under appropriate conditions, express the product gene. In addition, regulatory and signal sequences compatible with plant cells are available, such as the nopaline synthase promoter and  
20 polyadenylation signal sequences. Depicker et al., J. Mol. Appl. Gen., 1:561 (1982). In addition, DNA segments isolated from the upstream region of the T-DNA 780 gene are capable of activating or increasing transcription levels of plant-expressible genes in recombinant DNA-containing plant tissue. EP 321,196 published 21 June 1989.

25 However, interest has been greatest in vertebrate cells, and propagation of vertebrate cells in culture (tissue culture) has become a routine procedure in recent years [Tissue Culture, Academic Press, Kruse and Patterson, editors (1973)]. Examples of useful mammalian host cell lines are monkey kidney CV1 line transformed by SV40 (COS-7, ATCC CRL  
30 1651); human embryonic kidney line (293 or 293 cells subcloned for growth in suspension culture, Graham et al., J. Gen Virol., 36:59 [1977]); baby hamster kidney cells (BHK, ATCC CCL 10); Chinese hamster ovary cells/-DHFR (CHO, Urlaub and Chasin, Proc. Natl. Acad. Sci. USA, 77:4216 [1980]); dp12.CHO cells (EP 307,247 published 15 March 1989); mouse sertoli cells  
35 (TM4, Mather, Biol. Reprod., 23:243-251 [1980]); monkey kidney cells (CV1 ATCC CCL 70); African green monkey kidney cells (VERO-76, ATCC CRL-1587); human cervical carcinoma cells (HELA, ATCC CCL 2); canine kidney cells (MDCK, ATCC CCL 34); buffalo rat liver cells (BRL 3A, ATCC CRL 1442); human lung cells (W138, ATCC CCL 75); human liver cells (Hep G2, HB 8065); mouse  
40 mammary tumor (MMT 060562, ATCC CCL51); TRI cells (Mather et al., Annals N.Y. Acad. Sci., 383:44-68 [1982]); MRC 5 cells; FS4 cells; and a human hepatoma line (Hep G2).

Host cells are transformed with the above-described expression or cloning vectors of this invention and cultured in conventional nutrient  
45 media modified as appropriate for inducing promoters, selecting transformants, or amplifying the genes encoding the desired sequences.

Infection with *Agrobacterium tumefaciens* is used for transformation of certain plant cells, as described by Shaw et al., Gene, 23:315 (1983) and WO 89/05859 published 29 June 1989. For mammalian cells without such cell walls, the calcium phosphate precipitation method of Graham and van der Eb, Virology, 52:456-457 (1978) may be used. General aspects of mammalian cell host system transformations have been described by Axel in U.S. 4,399,216 issued 16 August 1983. Transformations into yeast are typically carried out according to the method of Van Solingen et al., J. Bact., 130:946 (1977) and Hsiao et al., Proc. Natl. Acad. Sci. (USA), 76:3829 (1979). However, other methods for introducing DNA into cells such as by nuclear injection or by protoplast fusion may also be used.

In the preferred embodiment the DNA is introduced into the host cells using electroporation. See Andreason, J. Tiss. Cult. Meth., 15:56-62 (1993), for a review of electroporation techniques useful for practicing the instantly claimed invention. It was discovered that electroporation techniques for introducing the DNA construct into the host cells were preferable over calcium phosphate precipitation techniques insofar as the latter could cause the DNA to break up and forming concatemers.

The mammalian host cells used to express the product gene herein may be cultured in a variety of media as discussed in the definitions section above. The media contains the selection agent used for selecting transformed host cells which have taken up the DNA construct (either as an intra- or extra-chromosomal element). To achieve selection of the transformed eukaryotic cells, the host cells may be grown in cell culture plates and individual colonies expressing the selectable gene (and thus the product gene) can be isolated and grown in growth medium until the nutrients are depleted. The host cells are then analyzed for transcription and/or transformation as discussed below. The culture conditions, such as temperature, pH, and the like, are those previously used with the host cell selected for expression, and will be apparent to the ordinarily skilled artisan.

Gene amplification and/or expression may be measured in a sample directly, for example, by conventional Southern blotting, Northern blotting to quantitate the transcription of mRNA (Thomas, Proc. Natl. Acad. Sci. USA, 77:5201-5205 [1980]), dot blotting (DNA analysis), or in situ hybridization, using an appropriately labeled probe, based on the sequences provided herein. Various labels may be employed, most commonly radioisotopes, particularly <sup>32</sup>P. However, other techniques may also be employed, such as using biotin-modified nucleotides for introduction into a polynucleotide. The biotin then serves as the site for binding to avidin or antibodies, which may be labeled with a wide variety of labels, such as radionuclides, fluorescens, enzymes, or the like. Alternatively, antibodies may be employed that can recognize specific duplexes, including DNA duplexes, RNA duplexes, and DNA-RNA hybrid duplexes or DNA-protein duplexes. The antibodies in turn may be labeled and the assay may be carried out where the duplex is bound to a surface, so that upon the

formation of duplex on the surface, the presence of antibody bound to the duplex can be detected.

Gene expression, alternatively, may be measured by immunological methods, such as immunohistochemical staining of tissue sections and assay  
5 of cell culture or body fluids, to quantitate directly the expression of gene product. With immunohistochemical staining techniques, a cell sample is prepared, typically by dehydration and fixation, followed by reaction with labeled antibodies specific for the gene product coupled, where the labels are usually visually detectable, such as enzymatic labels,  
10 fluorescent labels, luminescent labels, and the like. A particularly sensitive staining technique suitable for use in the present invention is described by Hsu et al., Am. J. Clin. Path., 75:734-738 (1980).

In the preferred embodiment, the mRNA is analyzed by quantitative PCR (to determine the efficiency of splicing) and protein expression is  
15 measured using ELISA as described in Example 1 herein.

The product of interest preferably is recovered from the culture medium as a secreted polypeptide, although it also may be recovered from host cell lysates when directly expressed without a secretory signal. When the product gene is expressed in a recombinant cell other than one of human  
20 origin, the product of interest is completely free of proteins or polypeptides of human origin. However, it is necessary to purify the product of interest from recombinant cell proteins or polypeptides to obtain preparations that are substantially homogeneous as to the product of interest. As a first step, the culture medium or lysate is centrifuged  
25 to remove particulate cell debris. The product of interest thereafter is purified from contaminant soluble proteins and polypeptides, for example, by fractionation on immunoaffinity or ion-exchange columns; ethanol precipitation; reverse phase HPLC; chromatography on silica or on a cation exchange resin such as DEAE; chromatofocusing; SDS-PAGE; ammonium sulfate  
30 precipitation; gel electrophoresis using, for example, Sephadex G-75; chromatography on plasminogen columns to bind the product of interest and protein A Sepharose columns to remove contaminants such as IgG.

The following examples are offered by way of illustration only and are not intended to limit the invention in any manner. All patent and  
35 literature references cited herein are expressly incorporated by reference.

#### EXAMPLE 1

##### tPA production using the dicistronic expression vectors

It was sought to increase the level of homogeneity with regard to expression levels of stable clones by expressing a selectable marker (such  
40 as DHFR) and the protein of interest from a single promoter. These vectors divert most of the transcript to product expression while linking it at a fixed ratio to DHFR expression via differential splicing.

Vectors were constructed which were derived from the vector pRK (Suva et al., Science, 237:893-896 [1987]) which contains an intron between the  
45 cytomegalovirus immediate early promoter (CMV) and the cDNA that encodes

the polypeptide of interest. The intron of pRK is 139 nucleotides in length, has a splice donor site derived from cytomegalovirus immediate early gene (CMVIE), and a splice acceptor site from an IgG heavy chain variable region (V<sub>H</sub>) gene (Eaton et al., Biochem., 25:8343 [1986]).

- 5 DHFR/intron vectors were constructed by inserting an EcoRV linker into the BSTX1 site present in the intron of pRK7. An 830 base-pair fragment containing a mouse DHFR coding fragment was inserted to obtain DHFR intron expression vectors which differ only in the sequence that comprises the splice donor site. Those sequences were altered by  
10 overlapping PCR mutagenesis to obtain sequences that match splice donor sites found between exons 3 and 4 of normal and mutant Ras genes. PCR was also used to destroy the splice donor site.

- A mouse DHFR cDNA fragment (Simonsen et al., Proc. Natl. Acad. Sci. USA, 80:2495-2499 [1983]) was inserted into the intron of this vector 59  
15 nucleotides downstream of the splice donor site. The splice donor site of this vector was altered by mutagenesis to change the ratio of spliced to non-spliced message in transfected cells. It has previously been shown that a single nucleotide change (G to A) converted a relatively efficient splice donor site found in the normal ras gene into an inefficient splice  
20 site (Cohen et al., Nature, 334:119-124 [1988]). This effect has been demonstrated in the context of the ras gene and confirmed when these sequences were transferred to human growth hormone constructs (Cohen et al., Cell, 58:461-472 [1989]). Additionally, a non functional 5' splice site (GT to CA) was constructed as a control ( $\Delta$ GT). A polylinker was  
25 inserted 35 nucleotides downstream of the 3' splice site to accept the cDNA of interest. A vector containing tPA (Pennica et al., Nature, 301:214-221 [1983]) was linearized downstream of the polyadenylation site before it was introduced into CHO cells (Potter et al., Proc. Natl. Acad. Sci. USA, 81:7161 [1984]).

- 30 Plasmid DNA's that contained DHFR/intron, tPA and (a) wild type ras (WT ras), i.e. Figure 3 (SEQ ID NO: 1), (b) mutant ras, or (c) non-functional splice donor site ( $\Delta$ GT) were introduced into CHO DHFR minus cells by electroporation. The intron vectors were each linearized downstream of the polyadenylation site by restriction endonuclease  
35 treatment. The control vector was linearized downstream of the second polyadenylation site. The DNA's were ethanol precipitated after phenol/chloroform extraction and were resuspended in 20 $\mu$ l 1/10 Tris EDTA. Then, 10 $\mu$ g of DNA was incubated with 10<sup>7</sup> CHO.dpl2 cells (EP 307,247 published 15 March 1989) in 1 ml of PBS on ice for 10 min. before  
40 electroporation at 400 volts and 330 $\mu$ f using a BRL Cell Porator.

Cells were returned to ice for 10 min. before being plated into non-selective medium. After 24 hours cells were fed nucleoside-free medium to select for stable DHFR+ clones which were pooled. The pooled DHFR+ clones were lysed and mRNA's were prepared.

- 45 To prepare the mRNA, RNA was extracted from 5 x 10<sup>7</sup> cells which were grown from pools of more than 200 clones derived from the stable

transfection of the three vectors, the essential construction of which is shown in Figure 1B and from non-transfected CHO cells. RNA was purified over oligo-DT cellulase (Collaborative Biomedical Products). 10µg of mRNA was then subjected to Northern blotting which involved running the mRNA on a 1.2% agarose, 6.6% formaldehyde gel, and transferring it to a nylon filter (Stratagene Duralon-UV membrane), prehybridized, probed and washed according to the manufacturer's instructions.

The filter was probed sequentially using probes (shown in Figure 1B) that would detect (a) the full length message, (b) both full length and spliced message, or (c) beta actin. Probing with the long probe showed that the vector that contains the efficient splice donor site (i.e. WT ras) generates predominately a mRNA of the size predicted for the spliced product while the other two vectors gave rise primarily to a mRNA that corresponds in size to non-spliced message. The DHFR probe detected only full length message and demonstrated that the WT ras splice donor derived vector generates very little full length message with which to confer a DHFR positive phenotype.

Figure 4 shows the number of DHFR positive colonies obtained after duplicate electroporations with the three intron vectors described above and from a conventional vector that has a CMV promoter driving tPA and a SV40 promoter driving DHFR (see Figure 2). The increase in colony number parallels the increase in full length message that accumulates with the modification of the splice donor sites. The conventional vector efficiently generates colonies and does not vary significantly from the ΔGT construct.

The level of tPA expression was determined by seeding cells in 1 ml of F12:DMEM (50:50, with 5% FBS) in 24 well dishes to near confluency. Growth of the cells continued until the media was exhausted. Media was then assayed by ELISA for tPA production. Briefly, anti-tPA antibody was coated onto the wells of an ELISA microtiter plate, media samples were added to the wells followed by washing. Binding of the antigen (tPA) was then quantified using horse radish peroxidase (HRPO) labelled anti-tPA antibody.

Figure 5A depicts the titers of secreted tPA protein after pooling the clones of each group shown in Figure 4. While the number of colonies increased with a weakening of splice donor function, the inverse was seen with respect to tPA expression. The expression levels are consistent with the RNA products that are observed; as more of the dicistronic message is spliced an increased amount of message will contain tPA as the first open reading frame resulting in increased tPA expression. A mutation of GT to CA in the splice donor site results in an abundance of DHFR positive colonies which express undetectable levels of tPA, possibly resulting from inefficient utilization of the second AUG. Importantly, Figure 5A also shows that expression levels obtained from one of the dicistronic vectors (with WT ras SD) was about threefold higher than that obtained with the control vector containing a CMV promoter/enhancer driving tPA, SV40

promoter/enhancer controlling DHFR and SV40 polyadenylation signals controlling the expression of tPA and DHFR.

Additionally, the homogeneity of expression in the pools was investigated. Figure 5B shows that all 20 clones generated by the WT ras splice donor site derived dicistronic vectors express detectable levels of tPA while only 4 of 20 clones generated by the control vector express tPA. None of the clones transfected with the non-splicing ( $\Delta$ GT) vector expressed tPA levels detectable by ELISA. This finding is consistent with previous observations that relatively few clones generated by conventional vectors make useful levels of protein.

Expression of tPA was increased following methotrexate amplification of pools. Figure 5C shows that 2 of the dicistronic vector derived pools (i.e. with WT ras and MUTANT ras SD sites) increased in expression markedly (8.4 and 7.7 fold), while the pool generated by the conventional vector increased only slightly (2.8 fold) when each was subjected to 200 nM Mtx. An overall increase of 9 fold was obtained using the best dicistronic (WT ras SD) versus the conventional vector following amplification. Growth of the highest expressing amplified pool in nutrient rich production medium yielded titers of 4.2  $\mu$ g/ml tPA.

It was shown that manipulation of the splice donor sequence alters the ratio of spliced to full length message and the number of colonies that form in selective medium. It was also shown that dicistronic expression vectors generate clones that express high levels of recombinant proteins. Surprisingly, it was possible to isolate high expressors which had the efficient WT ras splice donor site by selection for DHFR<sup>r</sup> cells despite the efficiency with which the DHFR gene was spliced from the RNA precursors formed in these cells.

## EXAMPLE 2

### TNFr-IgG production using the dicistronic expression vectors

To prove the general applicability of this approach, a second product was evaluated in the dicistronic vector system containing, as the DNA of interest, an immunoadhesin (TNFr-IgG) capable of binding tumor necrosis factor (TNF) (Ashkenazi et al., Proc. Natl. Acad. Sci. USA, 88:10535-10539 [1991]). The experiments described in Example 1 above were essentially repeated except that the product gene encoded the immunoadhesin TNFr-IgG. Plasmid DNA's that contained a TNFr-IgG cDNA and (a) WT ras, i.e. Figure 6 (SEQ ID NO: 2), (b) mutant ras or (c) nonfunctional splice donor site ( $\Delta$ GT) were introduced into the dp12.CHO cells as discussed for Example 1. See Figure 1C for an illustration of the DNA constructs.

It was discovered that the number of DHFR positive colonies generated by three of these vectors was similar to that seen with the tPA constructs. Expression of TNFr-IgG also paralleled that seen with the tPA constructs (Figure 7A). Amplification of pools from two of the constructs showed a marked increase in expression of immunoadhesin (9.6 and 6.8 fold) (Figure

7B). The best of these amplified pools expressed 9.5 µg/ml when grown in nutrient rich production medium.

Thus, it was again shown that dicistronic expression vectors generate clones that express high levels of recombinant proteins. Furthermore, 5 contrary to expectations, it was discovered that isolation of high product expressing host DHFR<sup>+</sup> cells was possible using an efficient splice donor site (i.e. the WT ras splice donor site).

### EXAMPLE 3

#### Antibody production using a dicistronic expression vector

10 The usefulness of this system for antibody expression was evaluated by testing production of an antibody directed against IgE (Presta et al., Journal of Immunology, 151:2623-2632 [1993]). Further, the flexibility of the system with regard to transcription initiation was tested by replacing the CMV promoter/enhancer present in the previous vectors with the 15 promoter/ enhancer derived from the early region of SV40 virus (Griffin, B., Structure and Genomic Organization of SV40 and Polyoma Virus, In J. Tooze [Ed] DNA Tumor Viruses, Cold Spring Harbor Laboratory, Cold Spring Harbor, New York). The heavy chain of the antibody was inserted downstream of DHFR as described in the earlier tPA and TNFr-IgG constructs. 20 Additionally, a new splice donor site sequence (GAC:GTAAGT) was engineered into the vector which matches the consensus splice donor site more closely than did the splice donor sites present in the vectors tested in Examples 1 and 2. The resultant expression vector is shown in Figures 1D and 9.

It was discovered that this vector produced fewer colonies than the 25 vectors previously tested, and produced predominantly a spliced RNA product. A second vector was constructed to have the light chain of the antibody under control of the SV40 promoter/enhancer and poly-A and the hygromycin B resistance gene under control of the CMV promoter/enhancer and SV40 poly-A. These vectors were linearized at unique HpaI sites downstream 30 of the poly-A signal, mixed at a ratio of light chain vector to heavy chain vector of 10:3 and electroporated into CHO cells using an optimized protocol (as discussed in Examples 1 and 2).

Figure 11 shows the levels of antibody expressed by clones and pools after selection in hygromycin B followed by selection for DHFR expression. 35 All 20 of the clones analyzed expressed high levels of antibody when grown in rich medium and varied from one another by only a factor of four. A pool of antibody producing clones was generated and assayed shortly after it was established. That pool was grown continuously for 6 weeks without a significant decrease in productivity demonstrating that its stability was 40 sufficient to generate gram quantities of protein from its large scale culture.

The pool was subjected to methotrexate amplification at 200nM and 1µM and achieved a greater than 2 fold increase in antibody titer. The 1µM Mtx resistant pool achieved a titer of 41 mg/L when grown under optimal 45 conditions in suspension culture.

The structure of the expressed antibody was examined. Proteins expressed by the 200nM methotrexate resistant pool and by a well characterized expression clone generated by conventional vectors (Presta et al. [1993], supra) were metabolically labeled with S<sup>35</sup> cysteine and methionine. In particular, confluent 35mm plates of cells were metabolically labeled with 50μCi each S-35 methionine and S-35 cysteine (Amersham) in serum free cysteine and methionine free F12:DMEM. After one hour, nutrient rich production media was added and labeled proteins were allowed to "chase" into the medium for six more hours. Proteins were run on a 12% SDS/PAGE gel (NOVEX) non-reduced or following reduction with B-mercaptoethanol. Dried gels were exposed to film for 16 hours. CHO control cells were also labeled.

The majority of the antibody protein is secreted with a molecular weight of about 155 kilodaltons, consistent with a properly disulfide-linked antibody molecule with 2 light and 2 heavy chains. Upon reduction the molecular weight shifts to 2 approximately equally abundant proteins of 22.5 and 55 kilodaltons. The protein generated from the pool is indistinguishable from the antibody produced by the well characterized expression clone, with no apparent increase of free heavy or light chain expressed by the pool.

#### CONCLUSION

The efficient expression system described herein utilizes vectors consisting of promoter/enhancer elements followed by an intron containing the selectable marker coding sequence, followed by the cDNA of interest and a polyadenylation signal.

Several splice donor site sequences were tested for their effect on colony number and expression of the cDNA of interest. A non-functional splice donor site, splice donor sites found in an intron between exons 3 and 4 of mutant (mutant ras) and normal (WT ras) forms of the Harvey Ras gene and another efficient SD site (see Example 3) were used. The vectors were designed to direct expression of dicistronic primary transcripts. Within a transfected cell some of the transcripts remain full length while the remainder are spliced to excise the DHFR coding sequence. When the splice donor site is weakened or destroyed an increase in colony number is observed.

Expression levels show the inverse pattern, with the most efficient splice donor sites generating the highest levels of tPA, TNF $\alpha$  immunoadhesin or anti-IgE V $\alpha$ .

The homogeneity of expression of clones generated by the ras splice donor site intron DHFR vectors was compared to clones generated from a conventional vector with a separate promoter/enhancer and polyadenylation signal for each DHFR and tPA. The DHFR intron vector gives rise to colonies that are much more homogeneous with regard to expression than those generated by the conventional vector. Non-expressing clones derived from the conventional vector may be the result of breaks in the tPA or



TNFr-IgG domain of the plasmid during integration into the genome or the result of methylation of promoter elements (Busslinger et al., Cell, 34:197-206 [1983]; Watt et al., Genes and Development, 2:1136-1143 [1988]) driving tPA or TNFr-IgG expression. Promoter silencing by methylation or  
5 breaks in the DHFR-intron vectors would very likely render them incapable of conferring a DHFR positive phenotype.

It was found that pools generated by the DHFR-intron vectors could be amplified in methotrexate and would increase in expression by a factor of 8.4 (tPA), or 9.8 (TNFr-IgG). Pools from conventional vectors increased  
10 by only 2.8 and 3.0 fold for tPA and TNFr-IgG when amplified similarly. Amplified pools resulted in 9 fold higher tPA levels and 15 fold higher TNFr-IgG levels when compared to the conventional vector amplified pools.

Without being limited to any theory, the increase in expression of methotrexate resistant pools derived from the dicistronic vectors is likely  
15 due to the transcriptional linkage of DHFR and the product; when cells are selected for increased DHFR expression they consistently over-express product. Conventional approaches lack selectable marker and cDNA expression linkage and therefore methotrexate amplification often generates DHFR overexpression without the concomitant increase in product expression.

20 A further increase of 4 and 6.3 fold in expression were obtained when amplified tPA and TNFr-IgG pools were transferred from the media used for the selections and amplifications to a nutrient rich production medium.

In Example 3, the expression vector had a splice donor site that more closely matches the consensus splice donor sequence and had the heavy chain  
25 of a humanized anti-IgE antibody inserted downstream. This vector was linearized and co-electroporated with a second linearized vector that expresses the hygromycin resistance gene and the light chain of the antibody each under the control of its own promoter/enhancer and poly-A signals. An excess of light chain expression vector over the heavy chain  
30 dicistronic expression vector was used to bias in favor of light chain expression. Clones and a pool were generated after hygromycin B and DHFR selections. The clones were found to express relatively consistent, high levels of antibody, as did the pool. The 1 $\mu$ M pool achieved a titer of 41mg/L when grown under optimal conditions in suspension culture.

35 The anti-IgE antibody was assessed by metabolic labeling followed by SDS/PAGE under reducing and non reducing conditions and found to be indistinguishable from the protein expressed by a highly characterized clonal cell line. Of particular importance is the finding that no free light chain is observed in the pool relative to the clone.

40 A stable expression system for CHO cells has been developed that produces high levels of recombinant proteins rapidly and with less effort than that required by other expression systems. The vector system generates stable clones that express consistently high levels thereby reducing the number of clones that must be screened to obtain a highly  
45 productive clonal line. Alternatively, pools have been used to conveniently generate moderate to high levels of protein. This approach

may be particularly useful when a number of related proteins are to be expressed and compared.

Without being limited to this theory, it is possible the vectors that have very efficient splice donor sites generate very productive clones  
5 because so little transcript remains non spliced that only integration events that lead to the generation of high levels of RNA produce enough DHFR protein to give rise to colonies in selective medium. The high level of spliced message from such clones is then translated into abundant amounts of the protein of interest. Pools of clones made concurrently by  
10 introducing conventional vectors expressed lower levels of protein, and were unstable with regard to long term expression, and expression could not be appreciably increased when the cells were subjected to methotrexate amplification.

The system developed herein is versatile in that it allows high  
15 levels of single and multiple subunit polypeptides to be rapidly generated from clones or pools of stable transfectants. This expression system combines the advantages of transient expression systems (rapid and labor non intensive generation of research amounts of protein) with the concurrent development of highly productive stable production cell lines.

## SEQUENCE LISTING

## (1) GENERAL INFORMATION:

- 5 (i) APPLICANT: GENENTECH, INC.
- (ii) TITLE OF INVENTION: METHOD FOR SELECTING HIGH-EXPRESSING HOST CELLS
- 10 (iii) NUMBER OF SEQUENCES: 4
- (iv) CORRESPONDENCE ADDRESS:
- (A) ADDRESSEE: Genentech, Inc.
- (B) STREET: 460 Point San Bruno Blvd
- 15 (C) CITY: South San Francisco
- (D) STATE: California
- (E) COUNTRY: USA
- (F) ZIP: 94080
- (v) COMPUTER READABLE FORM:
- 20 (A) MEDIUM TYPE: 5.25 inch, 360 Kb floppy disk
- (B) COMPUTER: IBM PC compatible
- (C) OPERATING SYSTEM: PC-DOS/MS-DOS
- (D) SOFTWARE: patin (Genentech)
- 25 (vi) CURRENT APPLICATION DATA:
- (A) APPLICATION NUMBER:
- (B) FILING DATE:
- (C) CLASSIFICATION:
- 30 (vii) PRIOR APPLICATION DATA:
- (A) APPLICATION NUMBER: 08/286740
- (B) FILING DATE: 05-AUG-1994
- (viii) ATTORNEY/AGENT INFORMATION:
- 35 (A) NAME: Lee, Wendy M.
- (B) REGISTRATION NUMBER: 00,000
- (C) REFERENCE/DOCKET NUMBER: 798PCT
- (ix) TELECOMMUNICATION INFORMATION:
- 40 (A) TELEPHONE: 415/225-1994
- (B) TELEFAX: 415/952-9881
- (C) TELEX: 910/371-7168

## (2) INFORMATION FOR SEQ ID NO:1:

- 45 (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 7360 bases
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- 50 (D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:
- 55 TTCGAGCTCG CCCGACATTG ATTATTGACT AGTTATTAAT AGTAATCAAT 50
- TACGGGGTCA TTAGTTCATA GCCCATATAT GGAGTTCGCG GTTACATAAC 100
- 60 TTACGGTAAA TGGCCCGCCT GGCTGACCGC CCAACGACCC CCGCCCATTG 150
- ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA 200
- 65 TTGACGTCAA TGGGTGGAGT ATTTACGGTA AACTGCCCAC TTGGCAGTAC 250

ATCAAGTGTA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT 300  
5 AAATGGCCCG CCTGGCATTG TGCCAGTAC ATGACCTTAT GGGACTTTCC 350  
TACTTGGCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTGATGC 400  
10 GGTTTTGGCA GTACATCAAT GGGCGTGGAT AGCGGTTTGA CTCACGGGGA 450  
TTTCCAAGTC TCCACCCCAT TGACGTCAAT GGGAGTTTGT TTTGGCACCA 500  
15 AAATCAACGG GACTTTCCAA AATGTCGTAA CAACTCCGCC CCATTGACGC 550  
AAATGGGCGG TAGGCGTGTA CGGTGGGAGG TCTATATAAG CAGAGCTCGT 600  
20 TTAGTGAACC GTCAGATCGC CTGGAGACGC CATCCACGCT GTTTTGACCT 650  
25 CCATAGAAGA CACCGGGACC GATCCAGCCT CCGCGGCCCG GAACGGTGCA 700  
TTGGAACGCG GATTCCCCGT GCCAAGAGTG CTGTAAGTAC CGCCTATAGA 750  
30 GCGATAAGAG GATTTTATCC CCGCTGCCAT CATGGTTCGA CCATTGAACT 800  
GCATCGTCGC CGTGTCCTCAA AATATGGGGA TTGGCAAGAA CGGAGACCTA 850  
35 CCCTGCCCTC CGCTCAGGAA CGCGTTCAAG TACTTCCAAA GAATGACCAC 900  
40 AACCTCTTCA GTGGAAGGTA AACAGAATCT GGTGATTATG GGTAGGAAAA 950  
CCTGGTTCTC CATTCCTGAG AAGAATCGAC CTTTAAAGGA CAGAATTAAT 1000  
45 ATAGTTCTCA GTAGAGAACT CAAAGAACCA CCACGAGGAG CTCATTTTCT 1050  
TGCCAAAAGT TTGGATGATG CCTTAAGACT TATTGAACAA CCGGAATTGG 1100  
50 CAAGTAAAGT AGACATGGTT TGGATAGTCG GAGGCAGTTC TGTTTACCAG 1150  
55 GAAGCCATGA ATCAACCAGG CCACCTTAGA CTCTTTGTGA CAAGGATCAT 1200  
GCAGGAATTT GAAAGTGACA CGTTTTTCCC AGAAATTGAT TTGGGGAAAT 1250  
60 ATAAACCTCT CCCAGAATAC CCAGGCGTCC TCTCTGAGGT CCAGGAGGAA 1300  
AAAGGCATCA AGTATAAGTT TGAAGTCTAC GAGAAGAAAG ACTAACAGGA 1350  
65 AGATGCTTTC AAGTTCTCTG CTCCCCTCCT AAAGCTATGC ATTTTATATA 1400

GACCATGGGA CTTTGTCTGG CTTTAGACCC CTTGGCTTC GTTAGAACGC 1450

5 GGCTACAATT AATACATAAC CTTATGTATC ATACACATAG ATTTAGGTGA 1500

CACTATAGAA TAACATCCAC TTTGCCTTTC TCTCCACAGG TGTCACTCCA 1550

10 GGTCAACTGC ACCTCGGTTC TAAGCTTGGG CTGCAGGTCG CCGTGAATTT 1600

AAGGGACGCT GTGAAGCAAT CATGGATGCA ATGAAGAGAG GGCTCTGCTG 1650

15 TGTGCTGCTG CTGTGTGGAG CAGTCTTCGT TTCGCCCAGC CAGGAAATCC 1700

ATGCCCCGATT CAGAAGAGGA GCCAGATCTT ACCAAGTGAT CTGCAGAGAT 1750

20 GAAAAACGC AGATGATATA CCAGCAACAT CAGTCATGGC TCGCCCTGT 1800

25 GCTCAGAAGC AACC GG GTGG AATATTGCTG GTGCAACAGT GGCAGGGCAC 1850

AGTGCCACTC AGTGCCTGTC AAAAGTTGCA GCGAGCCAAG GTGTTTCAAC 1900

30 GGGGGCACCT GCCAGCAGGC CCTGTACTTC TCAGATTTCTG TGTGCCAGTG 1950

CCCCGAAGGA TTTGCTGGGA AGTGCTGTGA AATAGATACC AGGGCCACGT 2000

35 GCTACGAGGA CCAGGGCATC AGCTACAGGG GCACGTGGAG CACAGCGGAG 2050

40 AGTGGCGCCG AGTGCACCAA CTGGAACAGC AGCGCGTTGG CCCAGAAGCC 2100

CTACAGCGGG CGGAGGCCAG ACGCCATCAG GCTGGGCCTG GGAACCACA 2150

45 ACTACTGCAG AAACCCAGAT CGAGACTCAA AGCCCTGGTG CTACGTCTTT 2200

AAGGCGGGGA AGTACAGCTC AGAGTTCTGC AGCACCCTG CCTGCTCTGA 2250

50 GGGAAACAGT GACTGCTACT TTGGAATGG GTCAGCCTAC CGTGGCACGC 2300

55 ACAGCCTCAC CGAGTCGGGT GCCTCCTGCC TCCCGTGGA TTCCATGATC 2350

CTGATAGGCA AGGTTTACAC AGCACAGAAC CCCAGTGCCC AGGCACTGGG 2400

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65 CCCTCCTGCT CCACCTGCGG CCTGAGACAG TACAGCCAGC CTCAGTTTCTG 2550

CATCAAAGGA GGGCTCTTCG CCGACATCGC CTCCCACCCC TGGCAGGCTG 2600

5 CCATCTTTGC CAAGCACAGG AGGTCGCCCC GAGAGCGGTT CCTGTGCGGG 2650

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20 GCACTGTGTG CCTTCCCCCG GCGGACCTGC AGCTGCCGGA CTGGACGGAG 2950

25 TGTGAGCTCT CCGGCTACGG CAAGCATGAG GCCTTGTCTC CTTTCTATTC 3000

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15 CGCGCGAGGC AGTATTCTTG AAGACGAAAG GGCCTCGTGA TACGCCTATT 5150

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20 CTTTTCGGGG AAATGTGCGC GGAACCCCTA TTTGTTTATT TTTCTAAATA 5250

CATTCAAATA TGTATCCGCT CATGAGACAA TAACCCTGAT AAATGCTTCA 5300

ATAATATTGA AAAAGGAAGA GTATGAGTAT TCAACATTTC CGTGTGCGCC 5350

30 TTATTCCTT TTTGCGGCA TTTGCCTTC CTGTTTTTGC TCACCCAGAA 5400

ACGCTGGTGA AAGTAAAAGA TGCTGAAGAT CAGTTGGGTG CACGAGTGGG 5450

35 TTACATCGAA CTGGATCTCA ACAGCGGTAA GATCCTTGAG AGTTTTCGCC 5500

CCGAAGAACG TTTTCCAATG ATGAGCACTT TTAAAGTTCT GCTATGTGGC 5550

GCGGTATTAT CCCGTGATGA CGCCGGGCAA GAGCAACTCG GTCGCCGCAT 5600

45 ACACTATTCT CAGAATGACT TGGTTGAGTA CTCACCAGTC ACAGAAAAGC 5650

ATCTTACGGA TGGCATGACA GTAAGAGAAT TATGCAGTGC TGCCATAACC 5700

50 ATGAGTGATA AACTGCGGC CAACTTACTT CTGACAACGA TCGGAGGACC 5750

GAAGGAGCTA ACCGCTTTTT TGCAACAT GGGGGATCAT GTAACGCGC 5800

TTGATCGTTG GGAACCGGAG CTGAATGAAG CCATACCAA CGACGAGCGT 5850

60 GACACCACGA TGCCAGCAGC AATGGCAACA ACGTTGCGCA AACTATTAAC 5900

TGGCGAACTA CTTACTCTAG CTTCCCGGCA ACAATTAATA GACTGGATGG 5950

65 AGGCGGATAA AGTTGCAGGA CCACTTCTGC GCTCGGCCCT TCCGGCTGGC 6000



TGGTTTATTG CTGATAAATC TGGAGCCGGT GAGCGTGGGT CTCGCGGTAT 6050  
5 CATTGCAGCA CTGGGGCCAG ATGGTAAGCC CTCCCGTATC GTAGTTATCT 6100  
ACACGACGGG GAGTCAGGCA ACTATGGATG AACGAAATAG ACAGATCGCT 6150  
10 GAGATAGGTG CCTCACTGAT TAAGCATTGG TAACTGTCAG ACCAAGTTTA 6200  
CTCATATATA CTTTAGATTG ATTTAAAACT TCATTTTAA TTTAAAAGGA 6250  
15 TCTAGGTGAA GATCCTTTTT GATAATCTCA TGACCAAAT CCCTTAACGT 6300  
GAGTTTTCGT TCCACTGAGC GTCAGACCCC GTAGAAAAGA TCAAAGGATC 6350  
20 TTCTTGAGAT CTTTTTTTTC TGCGCGTAAT CTGCTGCTTG CAAACAAAAA 6400  
25 AACCACCGCT ACCAGCGGTG GTTTGTTTGC CGGATCAAGA GCTACCAACT 6450  
CTTTTCCGA AGGTAAGTGG CTTCAAGCAGA GCGCAGATAC CAAATACTGT 6500  
30 CCTTCTAGTG TAGCCGTAGT TAGGCCACCA CTTCAAGAAC TCTGTAGCAC 6550  
CGCCTACATA CCTCGCTCTG CTAATCCTGT TACCAGTGGC TGCTGCCAGT 6600  
35 GCGGATAAGT CGTGTCTTAC CGGGTTGGAC TCAAGACGAT AGTTACCGGA 6650  
40 TAAGGCGCAG CGGTCGGGCT GAACGGGGGG TTCGTGCACA CAGCCCAGCT 6700  
TGGAGCGAAC GACCTACACC GAACTGAGAT ACCTACAGCG TGAGCATTGA 6750  
45 GAAAGCGCCA CGCTTCCCGA AGGGAGAAAG GCGGACAGGT ATCCGGTAAG 6800  
CGGCAGGGTC GGAACAGGAG AGCGCACGAG GGAGCTTCCA GGGGGAAACG 6850  
50 CCTGGTATCT TTATAGTCCT GTCGGGTTTC GCCACCTCTG ACTTGAGCGT 6900  
CGATTTTGT GATGCTCGTC AGGGGGGCGG AGCCTATGGA AAAACGCCAG 6950  
CAACGCGGCC TTTTACGGT TCCTGGCCTT TTGCTGGCCT TTTGCTCACA 7000  
60 TGTCTTTCC TGCCTTATCC CCTGATTCTG TGGATAACCG TATTACCGCC 7050  
TTTGAGTGAG CTGATACCGC TCGCCGACG CGAACGACCG AGCGCAGCGA 7100  
65 GTCAGTGAGC GAGGAAGCGG AAGAGCGCCC AATACGCAA CCGCCTCTCC 7150

CCGCGCGTTG GCCGATTCAT TAATCCAGCT GGCACGACAG GTTTCCTCGAC 7200  
5 TGGAAAGCGG GCAGTGAGCG CAACGCAATT AATGTGAGTT ACCTCACTCA 7250  
TTAGGCACCC CAGGCTTTAC ACTTTATGCT TCCGGCTCGT ATGTTGTGTG 7300  
10 GAATTGTGAG CGGATAACAA TTTCACACAG GAAACAGCTA TGACCATGAT 7350  
TACGAATTAA 7360

15

## (2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:  
20 (A) LENGTH: 6889 bases  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: double  
(D) TOPOLOGY: linear

25 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

TTTCGAGCTCG CCCGACATTG ATTATTGACT AGTTATTAAT AGTAATCAAT 50  
30 TACGGGGTCA TTAGTTCATA GCCCATATAT GGAGTTCCGC GTTACATAAC 100  
TTACGGTAAA TGGCCCGCCT GGCTGACCGC CCAACGACCC CCGCCCATTG 150  
35 ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA 200  
40 TTGACGTCAA TGGGTGGAGT ATTTACGGTA AACTGCCCCAC TTGGCAGTAC 250  
ATCAAGTGTA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT 300  
45 AAATGGCCCG CCTGGCATTG TGCCAGTAC ATGACCTTAT GGGACTTTCC 350  
TACTTGGCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTGATGC 400  
50 GGTTTTGGCA GTACATCAAT GGGCGTGGAT AGCGGTTTGA CTCACGGGGA 450  
55 TTTCCAAGTC TCCACCCCAT TGACGTCAAT GGGAGTTTGT TTTGGCACCA 500  
AAATCAACGG GACTTTCCAA AATGTCGTAA CAACTCCGCC CCATTGACGC 550  
60 AAATGGGCGG TAGGCGTGTA CGGTGGGAGG TCTATATAAG CAGAGCTCGT 600  
TTAGTGAACC GTCAGATCGC CTGGAGACGC CATCCACGCT GTTTTGACCT 650  
65 CCATAGAAGA CACCGGGACC GATCCAGCCT CCGCGGCCCG GAACGGTGCA 700

TTGGAACGCG GATTCCCCGT GCCAAGAGTG CTGTAAGTAC CGCCTATAGA 750

5 GCGATAAGAG GATTTTATCC CCGCTGCCAT CATGGTTCGA CCATTGAACT 800

GCATCGTCGC CGTGTCCCAA AATATGGGGA TTGGCAAGAA CGGAGACCTA 850

10 CCCTGCCCTC CGCTCAGGAA CGCGTTCAAG TACTTCCAAA GAATGACCAC 900

AACCTCTTCA GTGGAAGGTA AACAGAATCT GGTGATTATG GGTAGGAAAA 950

15 CCTGGTTCTC CATTCTGAG AAGAATCGAC CTTTAAAGGA CAGAATTAAT 1000

ATAGTTCTCA GTAGAGAACT CAAAGAACCA CCACGAGGAG CTCATTTTCT 1050

20 TGCCAAAAGT TTGGATGATG CTTAAGACT TATTGAACAA CCGGAATTGG 1100

25 CAAGTAAAGT AGACATGGTT TGGATAGTCG GAGGCAGTTC TGTTTACCAG 1150

GAAGCCATGA ATCAACCAGG CCACCTTAGA CTCTTTGTGA CAAGGATCAT 1200

30 GCAGGAATTT GAAAGTGACA CGTTTTTCCC AGAAATTGAT TTGGGGAAAT 1250

ATAAACCTCT CCCAGAATAC CCAGGCGTCC TCTCTGAGGT CCAGGAGGAA 1300

35 AAAGGCATCA AGTATAAGTT TGAAGTCTAC GAGAAGAAAG ACTAACAGGA 1350

40 AGATGCTTTC AAGTTCTCTG CTCCCCTCCT AAAGCTATGC ATTTTATAA 1400

GACCATGGGA CTTTGTCTGG CTTTAGACCC CTTGGCTTC GTTAGAACGC 1450

45 GGCTACAATT AATACATAAC CTTATGTATC ATACACATAG ATTTAGGTGA 1500

CACTATAGAA TAACATCCAC TTTGCCTTTC TCTCCACAGG TGTCACTCCA 1550

50 GGTCAACTGC ACCTCGGTTC TATCGATTGA ATTCCCCGGC CATAGCTGTC 1600

55 TGGCATGGGC CTCTCCACCG TGCCTGACCT GCTGCTGCCG CTGGTGCTCC 1650

TGGAGCTGTT GGTGGGAATA TACCCCTCAG GGGTTATTGG ACTGGTCCCT 1700

60 CACCTAGGGG ACAGGGAGAA GAGAGATAGT GTGTGTCCCC AAGGAAAATA 1750

TATCCACCCT CAAAATAATT CGATTTGCTG TACCAAGTGC CACAAAGGAA 1800

65 CCTACTTGTA CAATGACTGT CCAGGCCCGG GGCAGGATAC GGAAGTGCAGG 1850

GAGTGTGAGA GCGGCTCCTT CACCGCTTCA GAAAACCACC TCAGACACTG 1900  
CCTCAGCTGC TCCAAATGCC GAAAGGAAAT GGGTCAGGTG GAGATCTCTT 1950  
5 CTTGCACAGT GGACCGGGAC ACCGTGTGTG GCTGCAGGAA GAACCAGTAC 2000  
CGGCATTATT GGAGTGAAAA CCTTTTCCAG TGCTTCAATT GCAGCCTCTG 2050  
CCTCAATGGG ACCGTGCACC TCTCCTGCCA GGAGAAACAG AACACCGTGT 2100  
15 GCACCTGCCA TGCAGGTTTC TTTCTAAGAG AAAACGAGTG TGTCTCCTGT 2150  
AGTAACTGTA AGAAAAGCCT GGAGTGCACG AAGTTGTGCC TACCCCAGAT 2200  
20 TGAGAATGTT AAGGGCACTG AGGACTCAGG CACCACAGAC AAGAGAGTTG 2250  
AGCTCAAAAC CCCACTTGGT GACACAATC ACACATGCCC ACGGTGCCCA 2300  
GAGCCCAAAT CTTGTGACAC ACCTCCCCCG TGCCCACGGT GCCCAGAGCC 2350  
30 CAAATCTTGT GACACACCTC CCCCATGCCC ACGGTGCCCA GAGCCCAAAT 2400  
CTTGTGACAC ACCTCCCCCA TGCCCACGGT GCCCAGCACC TGAACTCCTG 2450  
35 GGAGGACCGT CAGTCTTCCT CTTCCCCCA AAACCAAGG ATACCCTTAT 2500  
GATTTCCCGG ACCCCTGAGG TCACGTGCGT GGTGGTGGAC GTGAGCCACG 2550  
AAGACCCCGA GGTCCAGTTC AAGTGGTACG TGGACGGCGT GGAGGTGCAT 2600  
45 AATGCCAAGA CAAAGCCGCG GGAGGAGCAG TTCAACAGCA CGTTCCGTGT 2650  
GGTCAGCGTC CTCACCGTCC TGCACCAGGA CTGGCTGAAC GGCAAGGAGT 2700  
50 ACAAGTGCAA GGTCTCCAAC AAAGCCCTCC CAGCCCCCAT CGAGAAAACC 2750  
ATCTCCAAA CCAAGGACA GCCCGAGAA CCACAGGTGT ACACCCTGCC 2800  
CCCATCCCGG GAGGAGATGA CCAAGAACCA GGTCAGCCTG ACCTGCCTGG 2850  
60 TCAAAGGCTT CTACCCAGC GACATCGCCG TGGAGTGGGA GAGCAGCGGG 2900  
CAGCCGGAGA ACAACTACAA CACCACGCCT CCCATGCTGG ACTCCGACGG 2950  
65 CTCCTTCTT CTCTACAGCA AGCTACCGT GGACAAGAGC AGGTGGCAGC 3000

AGGGGAACAT CTTCTCATGC TCCGTGATGC ATGAGGCTCT GCACAACCGC 3050  
5 TTCACGCAGA AGAGCCTCTC CCTGTCTCCG GGTAAATGAG TGCGACGGCC 3100  
GGGGATCCTC TAGAGTCGAC CTGCAGAAGC TTGGCCGCCA TGGCCCAACT 3150  
10 TGTTTATTGC AGCTTATAAT GGTACAAAT AAAGCAATAG CATCACAAAT 3200  
TTCACAAATA AAGCATTTTT TCACTGCAT TCTAGTTGTG GTTTGTCCAA 3250  
15 ACTCATCAAT GTATCTTATC ATGTCTGGAT CGATCGGGAA TTAATTCGGC 3300  
GCAGCACCAT GGCCTGAAAT AACCTCTGAA AGAGGAACTT GGTTAGGTAC 3350  
20 CTTCTGAGGC GGAAAGAACC AGCTGTGGAA TGTGTGTCAG TTAGGGTGTG 3400  
GAAAGTCCCC AGGCTCCCCA GCAGGCAGAA GTATGCAAAG CATGCATCTC 3450  
AATTAGTCAG CAACCAGGTG TGGAAAGTCC CCAGGCTCCC CAGCAGGCAG 3500  
30 AAGTATGCAA AGCATGCATC TCAATTAGTC AGCAACCATA GTCCCGCCCC 3550  
TAACTCCGCC CATCCCGCCC CTAACTCCGC CCAGTTCCGC CCATTCTCCG 3600  
35 CCCCATGGCT GACTAATTTT TTTTATTTAT GCAGAGGCCG AGGCCGCCTC 3650  
GGCCTCTGAG CTATTCCAGA AGTAGTGAGG AGGCTTTTTT GGAGGCCTAG 3700  
GCTTTTGCAA AAAGCTGTTA ACAGCTTGGC ACTGGCCGTC GTTTTACAAC 3750  
45 GTCGTGACTG GGAAAACCCT GCGGTTACCC AACTTAATCG CCTTGCAGCA 3800  
CATCCCCCCT TCGCCAGCTG GCGTAATAGC GAAGAGGCCC GCACCGATCG 3850  
50 CCCTTCCCAA CAGTTGCGTA GCCTGAATGG CGAATGGCGC CTGATGCGGT 3900  
ATTTTCTCCT TACGCATCTG TGCGGTATTT CACACCGCAT ACGTCAAAGC 3950  
AACCATAGTA CGCGCCCTGT AGCGGCGCAT TAAGCGCGGC GGGTGTGGTG 4000  
60 GTTACGCGCA GCGTGACCGC TACACTTGCC AGCGCCCTAG CGCCCGCTCC 4050  
TTTCGCTTTC TTCCCTTCCT TTCTCGCCAC GTTCGCCGGC TTTCCCGTC 4100  
65 AAGCTCTAAA TCGGGGGCTC CCTTTAGGGT TCCGATTAG TGCTTTACGG 4150

CACCTCGACC CCAAAAAACT TGATTGGGT GATGGTTCAC GTAGTGGGCC 4200  
ATCGCCCTGA TAGACGGTTT TTCGCCCTTT GACGTTGGAG TCCACGTTCT 4250  
5 TTAATAGTGG ACTCTGTTC CAACTGGAA CAACACTCAA CCCTATCTCG 4300  
GGCTATTCTT TTGATTATA AGGGATTTTG CCGATTTCGG CCTATTGGTT 4350  
10 AAAAAATGAG CTGATTAAAC AAAAATTAA CGCGAATTTT AACAAAATAT 4400  
15 TAACGTTTAC AATTTTATGG TGCACTCTCA GTACAATCTG CTCTGATGCC 4450  
GCATAGTTAA GCCAACTCCG CTATCGCTAC GTGACTGGGT CATGGCTGCG 4500  
20 CCCGACACC CGCCAACACC CGCTGACGCG CCCTGACGGG CTTGTCTGCT 4550  
CCCGGCATCC GCTTACAGAC AAGCTGTGAC CGTCTCCGGG AGCTGCATGT 4600  
25 GTCAGAGGTT TTCACCGTCA TCACCGAAAC GCGCGAGGCA GTATTCTTGA 4650  
30 AGACGAAAGG GCCTCGTGAT ACGCCTATTT TTATAGGTTA ATGTCATGAT 4700  
AATAATGGTT TCTTAGACGT CAGGTGGCAC TTTTCGGGGA AATGTGCGCG 4750  
35 GAACCCCTAT TTGTTTATTT TTCTAAATAC ATTCAAATAT GTATCCGCTC 4800  
ATGAGACAAT AACCTGATA AATGCTTCAA TAATATTGAA AAAGGAAGAG 4850  
TATGAGTATT CAACATTTCG GTGTCGCCCT TATTCCCTTT TTTGCGGCAT 4900  
45 TTTGCCTTCC TGTTTTTGCT CACCCAGAAA CGCTGGTGAA AGTAAAAGAT 4950  
GCTGAAGATC AGTTGGGTGC ACGAGTGGGT TACATCGAAC TGGATCTCAA 5000  
50 CAGCGGTAAG ATCCTTGAGA GTTTTCGCCC CGAAGAACGT TTTCCAATGA 5050  
TGAGCACTTT TAAAGTTCTG CTATGTGGCG CGGTATTATC CCGTGATGAC 5100  
GCCGGGCAAG AGCAACTCGG TCGCCGCATA CACTATTCTC AGAATGACTT 5150  
60 GGTTGAGTAC TCACCACTCA CAGAAAAGCA TCTTACGGAT GGCATGACAG 5200  
TAAGAGAATT ATGCAGTGCT GCCATAACCA TGAGTGATAA CACTGCGGCC 5250  
65 AACTTACTTC TGACAACGAT CGGAGGACCG AAGGAGCTAA CCGCTTTTTT 5300

GCACAACATG GGGGATCATG TAACTCGCCT TGATCGTTGG GAACCGGAGC 5350

5 TGAATGAAGC CATACCAAAC GACGAGCGTG ACACCACGAT GCCAGCAGCA 5400

ATGGCAACAA CGTTGCGCAA ACTATTAAGT GGCAGAACTAC TTACTCTAGC 5450

10 TTCCCGGCAA CAATTAATAG ACTGGATGGA GCGGATAAA GTTGCAGGAC 5500

CACTTCTGCG CTCGGCCCTT CCGGCTGGCT GGTTTATTGC TGATAAATCT 5550

15 GGAGCCGGTG AGCGTGGGTC TCGCGGTATC ATTGCAGCAC TGGGGCCAGA 5600

TGGTAAGCCC TCCCGTATCG TAGTTATCTA CACGACGGGG AGTCAGGCAA 5650

20 CTATGGATGA ACGAAATAGA CAGATCGCTG AGATAGGTGC CTCACTGATT 5700

25 AAGCATTGGT AACTGTCAGA CCAAGTTTAC TCATATATAC TTTAGATTGA 5750

TTTAAACTT CATTTTAAAT TAAAAGGAT CTAGGTGAAG ATCCTTTTTG 5800

30 ATAATCTCAT GACCAAATC CCTTAACGTG AGTTTTCGTT CCACTGAGCG 5850

TCAGACCCCG TAGAAAAGAT CAAAGGATCT TCTTGAGATC CTTTTTTTCT 5900

35 GCGCGTAATC TGCTGCTTGC AAACAAAAAA ACCACCGCTA CCAGCGGTGG 5950

40 TTTGTTTGCC GGATCAAGAG CTACCAACTC TTTTCCGAA GGTAAGTGGC 6000

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45 AGGCCACCAC TTCAAGAACT CTGTAGCACC GCCTACATAC CTCGCTCTGC 6100

TAATCCTGTT ACCAGTGGCT GCTGCCAGTG GCGATAAGTC GTGTCTTACC 6150

50 GGGTTGGACT CAAGACGATA GTTACCGGAT AAGGCGCAGC GGTGGGGCTG 6200

55 AACGGGGGGT TCGTGACAC AGCCAGCTT GGAGCGAACG ACCTACACCG 6250

AACTGAGATA CCTACAGCGT GAGCATTGAG AAAGCGCCAC GCTTCCCGAA 6300

60 GGGAGAAAGG CGGACAGGTA TCCGGTAAGC GGCAGGGTCG GAACAGGAGA 6350

GCGCACGAGG GAGCTTCCAG GGGGAAACGC CTGGTATCTT TATAGTCCTG 6400

65 TCGGGTTTCG CCACCTCTGA CTTGAGCGTC GATTTTGTG ATGCTCGTCA 6450

GGGGGGCGGA GCCTATGGAA AAACGCCAGC AACCGGCCT TTTTACGGTT 6500  
5 CCTGGCCTTT TGCTGGCCTT TTGCTCACAT GTTCTTTCCT GCGTTATCCC 6550  
CTGATTCTGT GGATAACCGT ATTACCGCCT TTGAGTGAGC TGATACCGCT 6600  
10 CGCCGCAGCC GAACGACCGA GCGCAGCGAG TCAGTGAGCG AGGAAGCGGA 6650  
AGAGCGCCCA ATACGCAAAC CGCCTCTCCC CGCGCGTTGG CCGATTCAAT 6700  
15 AATCCAGCTG GCACGACAGG TTTCCCGACT GGAAAGCGGG CAGTGAGCGC 6750  
AACGCAATTA ATGTGAGTTA CCTCACTCAT TAGGCACCCC AGGCTTTACA 6800  
20 CTTTATGCTT CCGGCTCGTA TGTGTGTGG AATTGTGAGC GGATAACAAT 6850  
25 TTCACACAGG AAACAGCTAT GACCATGATT ACGAATTAA 6889

(2) INFORMATION FOR SEQ ID NO:3:

30 (i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 6557 bases  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: double  
35 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

40 TTCGAGCTCG CCCGACATTG ATTATTGACT AGAGTCGATC GACAGCTGTG 50  
GAATGTGTGT CAGTTAGGGT GTGGAAAGTC CCCAGGCTCC CCAGCAGGCA 100  
45 GAAGTATGCA AAGCATGCAT CTCAATTAGT CAGCAACCAG GTGTGGAAAG 150  
TCCCCAGGCT CCCCAGCAGG CAGAAGTATG CAAAGCATGC ATCTCAATTA 200  
50 GTCAGCAACC ATAGTCCCGC CCCTAACTCC GCCCATCCCG CCCCTAACTC 250  
55 CGCCCAAGTTC CGCCCATCT CCGCCCCATG GCTGACTAAT TTTTTTTATT 300  
TATGCAGAGG CCGAGGCCGC CTCGGCCTCT GAGCTATTCC AGAAGTAGTG 350  
60 AGGAGGCTTT TTTGGAGGCC TAGGCTTTTG CAAAAGCTA GCTTATCCGG 400  
CCGGGAACGG TGCATTGGAA CGCGGATTCC CCGTGCCAAG AGTGACGTAA 450  
65 GTACCGCCTA TAGAGCGATA AGAGGATTTT ATCCCCGCTG CCATCATGGT 500



TCGACCATTG AACTGCATCG TCGCCGTGTC CCAAAATATG GGGATTGGCA 550  
5 AGAACGGAGA CCTACCCTGG CCTCCGCTCA GGAACGAGTT CAAGTACTTC 600  
CAAAGAATGA CCACAACCTC TTCAGTGGAA GGTAACAGA ATCTGGTGAT 650  
10 TATGGGTAGG AAAACCTGGT TCTCCATTCC TGAGAAGAAT CGACCTTTAA 700  
AGGACAGAAT TAATATAGTT CTCAGTAGAG AACTCAAAGA ACCACCACGA 750  
15 GGAGCTCATT TTCTTGCCAA AAGTTTGGAT GATGCCTTAA GACTTATTGA 800  
ACAACCGGAA TTGGCAAGTA AAGTAGACAT GGTTTGGATA GTCGGAGGCA 850  
20 GTTCTGTTTA CCAGGAAGCC ATGAATCAAC CAGGCCACCT TAGACTCTTT 900  
25 GTGACAAGGA TCATGCAGGA ATTTGAAAGT GACACGTTTT TCCCAGAAAT 950  
TGATTGGGG AAATATAAAC CTCTCCCAGA ATACCCAGGC GTCCTCTCTG 1000  
30 AGGTCCAGGA GGAAAAGGC ATCAAGTATA AGTTTGAAGT CTACGAGAAG 1050  
AAAGACTAAC AGGAAGATGC TTTCAAGTTC TCTGCTCCCC TCCTAAAGCT 1100  
35 ATGCATTTTT ATAAGACCAT GGGACTTTTG CTGGCTTTAG ATCCCCTTGG 1150  
40 CTTCGTTAGA ACGCAGCTAC AATTAATACA TAACCTTATG TATCATACAC 1200  
ATACGATTTA GGTGACACTA TAGATAACAT CCACTTTGCC TTTCTCTCCA 1250  
45 CAGGTGTCCA CTCCCAGGTC CAACTGCACC TCGTTCTAT CGATTGAATT 1300  
CCACCATGGG ATGGTCATGT ATCATCCTTT TTCTAGTAGC AACTGCAACT 1350  
50 GGAGTACATT CAGAAGTTCA GCTGGTGGAG TCTGGCGGTG GCCTGGTGCA 1400  
GCCAGGGGGC TCACTCCGTT TGTCCTGTGC AGTTTCTGGC TACTCCATCA 1450  
CCTCCGATA TAGCTGGAAC TGGATCCGTC AGGCCCCGGG TAAGGGCCTG 1500  
60 GAATGGGTTG CATCGATTAC GTATGCCGA TCGACTAACT ATAACCCTAG 1550  
CGTCAAGGGC CGTATCACTA TAAGTCGCGA CGATTCCAAA AACACATTCT 1600  
65 ACCTGCAGAT GAACAGCCTG CGTGCTGAGG AACTGCCGT CTATTATTGT 1650

GCTCGAGGCA GCCACTATTT CGGCGCCTGG CACTTCGCCC TGTGGGGTCA 1700  
5 AGGAACCCCTG GTCACCGTCT CCTCGGCCTC CACCAAGGGC CCATCGGTCT 1750  
TCCCCCTGGC ACCCTCCTCC AAGAGCACCT CTGGGGGCAC AGCGGCCCTG 1800  
10 GGCTGCCTGG TCAAGGACTA CTTCCCCGAA CCGGTGACGG TGTCGTGGAA 1850  
CTCAGGCGCC CTGACCAGCG GCGTGCACAC CTTCCCGGCT GTCCTACAGT 1900  
15 CCTCAGGACT CTACTCCCTC AGCAGCGTGG TGA CTGTGCC CTCTAGCAGC 1950  
TTGGGCACCC AGACCTACAT CTGCAACGTG AATCACAAGC CCAGCAACAC 2000  
20 CAAGGTGGAC AAGAAAGTTG AGCCCAAATC TTGTGACAAA ACTCACACAT 2050  
25 GCCCACCGTG CCCAGCACCT GAACTCCTGG GGGGACCGTC AGTCTTCCTC 2100  
TTCCCCCAA AACCCAAGGA CACCCTCATG ATCTCCCGGA CCCCTGAGGT 2150  
30 CACATGCGTG GTGGTGGACG TGAGCCACGA AGACCCTGAG GTCAAGTTCA 2200  
ACTGGTACGT GGACGGCGTG GAGGTGCATA ATGCCAAGAC AAAGCCGCGG 2250  
35 GAGGAGCAGT ACAACAGCAC GTACCGTGTG GTCAGCGTCC TCACCGTCCT 2300  
GCACCAGGAC TGGCTGAATG GCAAGGAGTA CAAGTGCAAG GTCTCCAACA 2350  
AAGCCCTCCC AGCCCCCATC GAGAAAACCA TCTCAAAGC CAAAGGGCAG 2400  
45 CCCCAGAAC CACAGGTGTA CACCCTGCCC CCATCCCGG AAGAGATGAC 2450  
CAAGAACCAG GTCAGCCTGA CCTGCCTGGT CAAAGGCTTC TATCCCAGCG 2500  
50 ACATCGCCGT GGAGTGGGAG AGCAATGGGC AGCCGAGAA CAACTACAAG 2550  
ACCACGCCTC CCGTGCTGGA CTCCGACGGC TCCTTCTTCC TCTACAGCAA 2600  
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60 CCGTGATGCA TGAGGCTCTG CACAACCACT ACACGCAGAA GAGCCTCTCC 2700  
CTGTCTCCGG GTAAATGAGT GCGACGGCCC TAGAGTCGAC CTGCAGAAGC 2750  
65 TTGGCCGCCA TGGCCCAACT TGTTTATTGC AGCTTATAAT GGTACAAAT 2800

AAAGCAATAG CATCACAAAT TTCACAAATA AAGCATTTTT TTTACTGCAT 2850

5 TCTAGTTGTG GTTTGTCCAA ACTCATCAAT GTATCTTATC ATGTCTGGAT 2900

CGATCGGGAA TTAATTCGGC GCAGCACCAT GGCCTGAAAT AACCTCTGAA 2950

10 AGAGGAACTT GGTTAGGTAC CTTCTGAGGC GGAAAGAACC AGCTGTGGAA 3000

TGTGTGTCAG TTAGGGTGTG GAAAGTCCCC AGGCTCCCCA GCAGGCAGAA 3050

15 GTATGCAAAG CATGCATCTC AATTAGTCAG CAACCAGGTG TGGAAAGTCC 3100

CCAGGCTCCC CAGCAGGCAG AAGTATGCAA AGCATGCATC TCAATTAGTC 3150

20 AGCAACCATA GTCCCGCCCC TAACTCCGCC CATCCCGCCC CTAACCTCCG 3200

25 CCAGTTCCGC CCATTCTCCG CCCCATGGCT GACTAATTTT TTTTATTTAT 3250

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30 AGGCTTTTTT GGAGGCCTAG GCTTTTGCAA AAAGCTGTTA CCTCGAGCGG 3350

CCGCTTAATT AAGGCGCGCC ATTTAAATCC TGCAGGTAAC AGCTTGGCAC 3400

35 TGGCCGTCGT TTTACAACGT CGTGA CTGGG AAAACCCTGG CGTTACCCAA 3450

CTTAATCGCC TTGCAGCACA TCCCCCTTC GCCAGCTGGC GTAATAGCGA 3500

AGAGGCCCGC ACCGATCGCC CTTCCCAACA GTTGCCTAGC CTGAATGGCG 3550

45 AATGGCGCCT GATGCGGTAT TTTCTCCTTA CGCATCTGTG CGGTATTTCA 3600

CACCGCATAC GTCAAAGCAA CCATAGTACG CGCCCTGTAG CGGCGCATT 3650

50 AGCGCGGCGG GTGTGGTGGT TACGCGCAGC GTGACCGCTA CACTTGCCAG 3700

55 CGCCCTAGCG CCCGCTCCTT TCGCTTTCTT CCCTTCCTTT CTCGCCACGT 3750

TCGCCGGCTT TCCCCGTCAA GCTCTAAATC GGGGGCTCCC TTTAGGGTTC 3800

60 CGATTTAGTG CTTTACGGCA CCTCGACCCC AAAAACTTG ATTTGGGTGA 3850

TGGTTCACGT AGTGGGCCAT CGCCCTGATA GACGGTTTTT CGCCCTTTGA 3900

65 CGTTGGAGTC CACGTTCTTT AATAGTGGAC TCTTGTCCA AACTGGAACA 3950

ACACTCAACC CTATCTCGGG CTATTCTTTT GATTTATAAG GGATTTTGCC 4000  
5 GATTTCGGCC TATTGGTTAA AAAATGAGCT GATTTAACAA AAATTTAACG 4050  
CGAATTTTAA CAAAATATTA ACGTTTACAA TTTTATGGTG CACTCTCAGT 4100  
10 ACAATCTGCT CTGATGCCGC ATAGTTAAGC CAACTCCGCT ATCGCTACGT 4150  
GACTGGGTCA TGGCTGCGCC CCGACACCCG CCAACACCCG CTGACGCGCC 4200  
15 CTGACGGGCT TGTCTGCTCC CGGCATCCGC TTACAGACAA GCTGTGACCG 4250  
TCTCCGGGAG CTGCATGTGT CAGAGGTTTT CACCGTCATC ACCGAAACGC 4300  
20 GCGAGGCAGT ATTCTTGAAG ACGAAAGGGC CTCGTGATAC GCCTATTTTT 4350  
ATAGGTTAAT GTCATGATAA TAATGGTTTC TTAGACGTCA GGTGGCACTT 4400  
TTCGGGGAAA TGTGCGCGGA ACCCCTATTT GTTTATTTTT CTAAATACAT 4450  
30 TCAAATATGT ATCCGCTCAT GAGACAATAA CCCTGATAAA TGCTTCAATA 4500  
ATATTGAAAA AGGAAGAGTA TGAGTATTCA ACATTTCCGT GTCGCCCTTA 4550  
35 TTCCCTTTTT TCGGCATTT TGCCTTCCTG TTTTGTCTCA CCCAGAAACG 4600  
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CATCGAACTG GATCTCAACA GCGGTAAGAT CCTTGAGAGT TTTCGCCCCG 4700  
45 AAGAACGTTT TCCAATGATG AGCACTTTTA AAGTTCTGCT ATGTGGCGCG 4750  
GTATTATCCC GTGATGACGC CGGGCAAGAG CAACTCGGTC GCCGCATACA 4800  
50 CTATTCTCAG AATGACTTGG TTGAGTACTC ACCAGTCACA GAAAAGCATC 4850  
TTACGGATGG CATGACAGTA AGAGAATTAT GCAGTGCTGC CATAACCATG 4900  
AGTGATAACA CTGCGGCCAA CTACTTCTG ACAACGATCG GAGGACCGAA 4950  
60 GGAGCTAACC GCTTTTTTGC ACAACATGGG GGATCATGTA ACTCGCCTTG 5000  
ATCGTTGGGA ACCGGAGCTG AATGAAGCCA TACCAAACGA CGAGCGTGAC 5050  
65 ACCACGATGC CAGCAGCAAT GGCAACAACG TTGCGCAAAC TATTAAGTGG 5100

CGAACTACTT ACTCTAGCTT CCCGGCAACA ATTAATAGAC TGGATGGAGG 5150

5 CGGATAAAGT TGCAGGACCA CTTCTGCGCT CGGCCCTTCC GGCTGGCTGG 5200

TTTATTGCTG ATAAATCTGG AGCCGGTGAG CGTGGGTCTC GCGGTATCAT 5250

10 TGCAGCACTG GGGCCAGATG GTAAGCCCTC CCGTATCGTA GTTATCTACA 5300

CGACGGGGAG TCAGGCAACT ATGGATGAAC GAAATAGACA GATCGCTGAG 5350

15 ATAGGTGCCT CACTGATTAA GCATTGGTAA CTGTCAGACC AAGTTTACTC 5400

ATATATACTT TAGATTGATT TAAAACTTCA TTTTAAATTT AAAAGGATCT 5450

20 AGGTGAAGAT CCTTTTGTAT AATCTCATGA CCAAATCCC TTAACGTGAG 5500

25 TTTTCGTTCC ACTGAGCGTC AGACCCCGTA GAAAAGATCA AAGGATCTTC 5550

TTGAGATCCT TTTTTTCTGC GCGTAATCTG CTGCTTGCAA ACAAAAAAAC 5600

30 CACCGCTACC AGCGGTGGTT TGTTTGCCCG ATCAAGAGCT ACCAACTCTT 5650

TTTCCGAAGG TAACTGGCTT CAGCAGAGCG CAGATACCAA ATACTGTCCT 5700

35 TCTAGTGTAG CCGTAGTTAG GCCACCACTT CAAGAACTCT GTAGCACCGC 5750

40 CTACATACCT CGCTCTGCTA ATCCTGTTAC CAGTGGCTGC TGCCAGTGGC 5800

GATAAGTCGT GTCTTACCGG GTTGGACTCA AGACGATAGT TACCGGATAA 5850

45 GCGCGAGCGG TCGGGCTGAA CGGGGGGTTC GTGCACACAG CCCAGCTTGG 5900

AGCGAACGAC CTACACCGAA CTGAGATACC TACAGCGTGA GCATTGAGAA 5950

50 AGCGCCACGC TTCCCGAAGG GAGAAAGGCG GACAGGTATC CGGTAAGCGG 6000

55 CAGGGTCGGA ACAGGAGAGC GCACGAGGGA GCTTCCAGGG GGAAACGCCT 6050

GGTATCTTTA TAGTCCTGTC GGGTTTCGCC ACCTCTGACT TGAGCGTCGA 6100

60 TTTTGTGAT GCTCGTCAGG GGGGCGGAGC CTATGGAAAA ACGCCAGCAA 6150

CGCGGCCTTT TTACGGTTCC TGGCCTTTTG CTGGCCTTTT GCTCACATGT 6200

65 TCTTCCTGCT GTTATCCCCT GATTCTGTGG ATAACCGTAT TACCGCCTTT 6250

GAGTGAGCTG ATACCGCTCG CCGCAGCCGA ACGACCGAGC GCAGCGAGTC 6300  
AGTGAGCGAG GAAGCGGAAG AGCGCCCAAT ACGCAAACCG CCTCTCCCCG 6350  
5 CGCGTTGGCC GATTCATTAA TCCAGCTGGC ACGACAGGTT TCCCGACTGG 6400  
AAAGCGGGCA GTGAGCGCAA CGCAATTAAT GTGAGTTACC TCACTCATT 6450  
10 GGCACCCAG GCTTTACACT TTATGCTTCC GGCTCGTATG TTGTGTGGAA 6500  
GGCTCGTATG TTGTGTGGAA 6500  
15 TTGTGAGCGG ATAACAATTT CACACAGGAA ACAGCTATGA CCATGATTAC 6550  
GAATTAA 6557  
20

## (2) INFORMATION FOR SEQ ID NO:4:

25 (i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 7305 bases  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: double  
(D) TOPOLOGY: linear

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

TTCGAGCTCG CCCGACATTG ATTATTGACT AGTTATTAAT AGTAATCAAT 50  
35 TACGGGGTCA TTAGTTCATA GCCCATATAT GGAGTTCCGC GTTACATAAC 100  
TTACGGTAAA TGGCCCGCCT GGCTGACCGC CCAACGACCC CCGCCCATTG 150  
40 ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA 200  
ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA 200  
45 TTGACGTCAA TGGGTGGAGT ATTTACGGTA AACTGCCCCAC TTGGCAGTAC 250  
ATCAAGTGTA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT 300  
50 AAATGGCCCCG CCTGGCATTG TGGCCAGTAC ATGACCTTAT GGGACTTTCC 350  
TACTTGGCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTGATGC 400  
GGTTTGGCA GTACATCAAT GGGCGTGGAT AGCGGTTTGA CTCACGGGGA 450  
60 TTTCCAAGTC TCCACCCCAT TGACGTCAAT GGGAGTTTGT TTTGGCACCA 500  
AAATCAACGG GACTTTCCAA AATGTCGTAA CAACTCCGCC CCATTGACGC 550  
65 AAATGGGCGG TAGGCGTGTA CGGTGGGAGG TCTATATAAG CAGAGCTCGT 600

TTAGTGAACC GTCAGATCGC CTGGAGACGC CATCCACGCT GTTTTGACCT 650

5 CCATAGAAGA CACCGGGACC GATCCAGCCT CCGCGGCCGG GAACGGTGCA 700

TTGGAACGCG GATTCCCCGT GCCAAGAGTG ACGTAAGTAC CGCCTATAGA 750

10 GTCTATAGGC CCACCCCTT GGCTTCGTTA GAACGCGGCT ACAATTAATA 800

CATAACCTTA TGTATCATAC ACATACGATT TAGGTGACAC TATAGAATAA 850

15 CATCCACTTT GCCTTTCTCT CCACAGGTGT CCACTCCCAG GTCCAACTGC 900

ACCTCGGTTT TAAGCTTATC GATATGAAA AGCCTGAACT CACCGCGACG 950

20 TCTGTCGAGA AGTTTCTGAT CGAAAAGTTC GACAGCGTCT CCGACCTGAT 1000

25 GCAGCTCTCG GAGGGCGAAG AATCTCGTGC TTTCAGCTTC GATGTAGGAG 1050

GGCGTGGATA TGTCTGCGG GTAAATAGCT GCGCCGATGG TTTCTACAAA 1100

30 GATCGTTATG TTTATCGGCA CTTTGCATCG GCCGCGCTCC CGATTCCGGA 1150

AGTGCTTGAC ATTGGGGAAT TCAGCGAGAG CCTGACCTAT TGCATCTCCC 1200

35 GCCGTGCACA GGGTGTACG TTGCAACACC TGCCTGAAAC CGAACTGCCC 1250

40 GCTGTTCTGC AGCCGGTTCG GGAGGCCATG GATGCGATCG CTGCGGCCGA 1300

TCTTAGCCAG ACGAGCGGGT TCGGCCCATC CGGACCGCAA GGAATCGGTC 1350

45 AATACACTAC ATGGCGTGAT TTCATATGCG CGATTGCTGA TCCCCATGTG 1400

TATCACTGGC AAACGTGAT GGACGACACC GTCAGTCCGT CCGTCGCGCA 1450

50 GGCTCTCGAT GAGCTGATGC TTTGGGCCGA GGAAGTCCGC 1500

55 ACCTCGTGCA CGCGGATTC GGCTCCAACA ATGTCCTGAC GGACAATGGC 1550

CGCATAACAG CGGTCATTGA CTGGAGCGAG GCGATGTTTG GGGATTCCCA 1600

60 ATACGAGGTC GCCAACATCT TCTTCTGGAG GCCGTGGTTG GCTTGATATG 1650

AGCAGCAGAC GTACTTCGAG CGGAGGCATC CGGAGCTTGC AGGATCGCCG 1700

65 CGGCTCCGGG CGTATATGCT CCGCATGGT CTTGACCAAC TCTATCAGAG 1750

CTTGGTTGAC GGCAATTTTCG ATGATGCAGC TTGGGCGCAG GGTTCGATGCG 1800  
ACGCAATCGT CCGATCCGGA GCCGGGACTG TCGGGCGTAC ACAAATCGCC 1850  
5 CGCAGAAGCG CGGCCGTCTG GACCGATGGC TGTGTAGAAG TACTCGCCGA 1900  
TAGTGGA AAC CGACGCCCCA GCACTCGTCC GAGGGCAAAG GAATAGAGTA 1950  
10 GATGCCGACC GAAGGATCCC CGGGGAATTC AATCGATGGC CGCCATGGCC 2000  
CAACTTGTTT ATTGCAGCTT ATAATGGTTA CAAATAAAGC AATAGCATCA 2050  
15 CAAATTTTAC AAATAAAGCA TTTTTCAC TGCATTCTAG TTGTGGTTTG 2100  
TCCAACTCA TCAATGTATC TTATCATGTC TGGATCGATC GGAATTAAT 2150  
20 TCGGCGCAGC ACCATGGCCT GAAATAACCT CTGAAAGAGG AACTTGGTTA 2200  
GGTACCTTCT GAGGCGGAAA GAACCAGCTG TGAATGTGT GTCAGTTAGG 2250  
30 GTGTGGAAG TCCCCAGGCT CCCCAGCAGG CAGAAGTATG CAAAGCATGC 2300  
ATCTCAATTA GTCAGCAACC AGGTGTGGAA AGTCCCCAGG CTCCCCAGCA 2350  
GGCAGAAGTA TGCAAAGCAT GCATCTCAAT TAGTCAGCAA CCATAGTCCC 2400  
40 GCCCCTAACT CCGCCCATCC CGCCCTAAC TCCGCCAGT TCCGCCATT 2450  
CTCCGCCCCA TGGCTGACTA ATTTTCTTA TTTATGCAGA GGCCGAGGCC 2500  
45 GCCTCGGCCT CTGAGCTATT CCAGAAGTAG TGAGGAGGCT TTTTGGAGG 2550  
CCTAGGCTTT TGCAAAAAGC TAGCTTATCC GGCCGGGAAC GGTGCATTGG 2600  
50 AACGCGGATT CCCCCTGCCA AGAGTCAGGT AAGTACCGCC TATAGAGTCT 2650  
ATAGGCCAC CCCCTGGCT TCGTTAGAAC GCGGCTACAA TTAATACATA 2700  
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60 CACAGGTGTC CACTCCCAGG TCCAACTGCA CCTCGGTTTCG CGAAGCTAGC 2800  
TTGGGCTGCA TCGATTGAAT TCCACCATGG GATGGTCATG TATCATCCTT 2850  
65 TTTCTAGTAG CAACTGCAAC TGGAGTACAT TCAGATATCC AGCTGACCCA 2900



GTCCCCGAGC TCCCTGTCCG CCTCTGTGGG CGATAGGGTC ACCATCACCT 2950

5 GCCGTGCCAG TCAGAGCGTC GATTACGATG GTGATAGCTA CATGAACTGG 3000

TATCAACAGA AACCAGGAAA AGCTCCGAAA CTACTGATTT ACGCGGCCTC 3050

10 GTACCTGGAG TCTGGAGTCC CTTCTCGCTT CTCTGGATCC GGTTCCTGGA 3100

CGGATTTCAC TCTGACCATC AGCAGTCTGC AGCCGGAAGA CTTGCAACT 3150

15 TATTACTGTC AGCAAAGTCA CGAGGATCCG TACACATTG GACAGGGTAC 3200

CAAGGTGGAG ATCAAACGAA CTGTGGCTGC ACCATCTGTC TTCATCTTCC 3250

20 CGCCATCTGA TGAGCAGTTG AAATCTGGAA CTGCCTCTGT TGTGTGCCTG 3300

CTGAATAACT TCTATCCCAG AGAGGCCAAA GTACAGTGA AGGTGGATAA 3350

CGCCCTCCAA TCGGGTAACT CCCAGGAGAG TGTCACAGAG CAGGACAGCA 3400

30 AGGACAGCAC CTACAGCCTC AGCAGCACCC TGACGCTGAG CAAAGCAGAC 3450

TACGAGAAAC ACAAAGTCTA CGCCTGCGAA GTCACCCATC AGGGCCTGAG 3500

35 CTCGCCCCGTC ACAAAGAGCT TCAACAGGGG AGAGTGTTAA GCTTCGATGG 3550

40 CCGCCATGGC CCAACTTGTT TATTGCAGCT TATAATGGTT ACAAATAAAG 3600

CAATAGCATC ACAAATTTC AATAAAGC ATTTTTTTCA CTGCATTCTA 3650

45 GTTGTGGTTT GTCCAACTC ATCAATGTAT CTTATCATGT CTGGATCGAT 3700

CGGGAATTAA TTCGGCGCAG CACCATGGCC TGAAATAACC TCTGAAAGAG 3750

50 GAACTTGGTT AGGTACCTC TGAGGCGGAA AGAACCAGCT GTGGAATGTG 3800

TGTCAGTTAG GGTGTGAAAA GTCCCCAGGC TCCCCAGCAG GCAGAAGTAT 3850

GCAAAGCATG CATCTCAATT AGTCAGCAAC CAGGTGTGGA AAGTCCCCAG 3900

60 GCTCCCCAGC AGGCAGAAGT ATGCAAAGCA TGCATCTCAA TTAGTCAGCA 3950

ACCATAGTCC CGCCCCTAAC TCCGCCCATC CCGCCCCTAA CTCCGCCAG 4000

65 TTCCGCCCAT TCTCCGCCCC ATGGCTGACT AATTTTTTTT ATTTATGCAG 4050

AGGCCGAGGC CGCCTCGGCC TCTGAGCTAT TCCAGAAGTA GTGAGGAGGC 4100  
TTTTTTGGAG GCCTAGGCTT TTGCAAAAAG CTGTAAACAG CTTGGCACTG 4150  
5 GCCGTCGTTT TACAACGTCG TGA CTGGGAA AACCTGGCG TTACCCAACT 4200  
TAATCGCCTT GCAGCACATC CCCCCTTCGC CAGCTGGCGT AATAGCGAAG 4250  
10 AGGCCCGCAC CGATCGCCCT TCCCAACAGT TGCGTAGCCT GAATGGCGAA 4300  
TGGCGCCTGA TGGGTATTT TCTCCTTACG CATCTGTGCG GTATTTTACA 4350  
15 CCGCATACGT CAAAGCAACC ATAGTACGCG CCCTGTAGCG GCGCATTAAG 4400  
CGCGGCGGGT GTGGTGGTTA CGCGCAGCGT GACCGCTACA CTTGCCAGCG 4450  
20 CCCTAGCGCC CGCTCCTTTC GCTTTCTTCC CTTCTTTCT CGCCACGTTT 4500  
GCCGCTTTC CCCGTCAAGC TCTAAATCGG GGGCTCCCTT TAGGGTTCCG 4550  
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35 TTGGAGTCCA CGTTCTTTAA TAGTGGACTC TTGTTCCAAA CTGGAACAAC 4700  
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45 AATTTTAACA AAATATTAC GTTTACAATT TTATGGTGCA CTCTCAGTAC 4850  
AATCTGCTCT GATGCCGCAT AGTTAAGCCA ACTCCGCTAT CGCTACGTGA 4900  
50 CTGGGTCATG GCTGCGCCCC GACACCCGCC AACACCCGCT GACGCGCCCT 4950  
GACGGGCTTG TCTGCTCCCG GCATCCGCTT ACAGACAAGC TGTGACCGTC 5000  
TCCGGGAGCT GCATGTGTCA GAGGTTTTCA CCGTCATCAC CGAAACGCGC 5050  
60 GAGGCAGTAT TCTTGAAGAC GAAAGGGCCT CGTGATACGC CTATTTTTAT 5100  
AGGTTAATGT CATGATAATA ATGGTTTCTT AGACGTCAGG TGGCACTTTT 5150  
65 CGGGGAAATG TGCGCGGAAC CCCTATTTGT TTATTTTCT AAATACATTC 5200

AAATATGTAT CCGCTCATGA GACAATAACC CTGATAAATG CTTCAATAAT 5250

ATTGAAAAAG GAAGAGTATG AGTATTCAAC ATTTCCGTGT CGCCCTTATT 5300

5 CCCTTTTTTG CGGCATTTTG CCTTCCTGTT TTTGCTCACC CAGAAACGCT 5350

GGTGAAAGTA AAAGATGCTG AAGATCAGTT GGGTGACGA GTGGGTTACA 5400

TCGAACTGGA TCTCAACAGC GGTAAGATCC TTGAGAGTTT TCGCCCCGAA 5450

15 GAACGTTTTTC CAATGATGAG CACTTTTAAA GTTCTGCTAT GTGGCGCGGT 5500

ATTATCCCGT GATGACGCCG GGCAAGAGCA ACTCGGTCGC CGCATACACT 5550

20 ATTCTCAGAA TGACTTGGTT GAGTACTCAC CAGTCACAGA AAAGCATCTT 5600

ACGGATGGCA TGACAGTAAG AGAATTATGC AGTGCTGCCA TAACCATGAG 5650

TGATAACACT GCGGCCAACT TACTTCTGAC AACGATCGGA GGACCGAAGG 5700

30 AGCTAACCGC TTTTTTGAC AACATGGGGG ATCATGTAAC TCGCCTTGAT 5750

CGTTGGGAAC CGGAGCTGAA TGAAGCCATA CCAACGACG AGCGTGACAC 5800

35 CACGATGCCA GCAGCAATGG CAACAACGTT GCGCAAATA TTAAGTGGCG 5850

AACTACTTAC TCTAGCTTCC CGGCAACAAT TAATAGACTG GATGGAGGCG 5900

GATAAAGTTG CAGGACCACT TCTGCGCTCG GCCCTTCCGG CTGGCTGGTT 5950

45 TATTGCTGAT AAATCTGGAG CCGGTGAGCG TGGGTCTCGC GGTATCATTG 6000

CAGCACTGGG GCCAGATGGT AAGCCCTCCC GTATCGTAGT TATCTACACG 6050

50 ACGGGGAGTC AGGCAACTAT GGATGAACGA AATAGACAGA TCGCTGAGAT 6100

AGGTGCCTCA CTGATTAAGC ATTGGTAACT GTCAGACCAA GTTTACTCAT 6150

ATATACTTTA GATTGATTTA AAACCTCATT TTTAATTAA AAGGATCTAG 6200

60 GTGAAGATCC TTTTGTGATA TCTCATGACC AAAATCCCTT AACGTGAGTT 6250

TTCGTTCCAC TGAGCGTCAG ACCCGTAGA AAAGATCAAA GGATCTTCTT 6300

65 GAGATCCTTT TTTTCTGCGC GTAATCTGCT GCTTGCAAAC AAAAAACCA 6350

CCGCTACCAG CGGTGGTTTG TTTGCCGGAT CAAGAGCTAC CAACTCTTTT 6400  
5 TCCGAAGGTA ACTGGCTTCA GCAGAGCGCA GATACCAAAT ACTGTCCTTC 6450  
TAGTGTAGCC GTAGTTAGGC CACCACTTCA AGAACTCTGT AGCACC GCCT 6500  
10 ACATACCTCG CTCTGCTAAT CCTGTTACCA GTGGCTGCTG CCAGTGGCGA 6550  
TAAGTCGTGT CTTACCGGGT TGGACTCAAG ACGATAGTTA CCGGATAAGG 6600  
15 CGCAGCGGTC GGGCTGAACG GGGGGTTCGT GCACACAGCC CAGCTTGGAG 6650  
CGAACGACCT ACACCGAACT GAGATACCTA CAGCGTGAGC ATTGAGAAAG 6700  
20 CGCCACGCTT CCCGAAGGGA GAAAGGCGGA CAGGTATCCG GTAAGCGGCA 6750  
GGGTCGGAAC AGGAGAGCGC ACGAGGGAGC TTCCAGGGGG AAACGCCTGG 6800  
TATCTTTATA GTCCTGTCGG GTTTCGCCAC CTCTGACTTG AGCGTCGATT 6850  
30 TTTGTGATGC TCGTCAGGGG GCGGAGCCT ATGGAAAAAC GCCAGCAACG 6900  
CGGCCTTTTT ACGGTTCTCTG GCCTTTTGCT GGCCTTTTGC TCACATGTTT 6950  
35 TTTCTGCGT TATCCCCTGA TTCTGTGGAT AACC GTATTA CCGCCTTTGA 7000  
GTGAGCTGAT ACCGCTCGCC GCAGCCGAAC GACCGAGCGC AGCGAGTCAG 7050  
TGAGCGAGGA AGCGGAAGAG CGCCCAATAC GCAAACCGCC TCTCCCCGCG 7100  
45 CGTTGGCCGA TTCATTAATC CAGCTGGCAC GACAGGTTTC CCGACTGGAA 7150  
AGCGGGCAGT GAGCGCAACG CAATTAATGT GAGTTACCTC ACTCATTAGG 7200  
50 CACCCCAGGC TTTACACTTT ATGCTTCCGG CTCGTATGTT GTGTGGAATT 7250  
GTGAGCGGAT AACAAATTCA CACAGGAAAC AGCTATGACC ATGATTACGA 7300  
55 ATTA 7305  
60

CLAIMS

1. A DNA construct comprising a transcriptional initiation site, a transcriptional termination site, a selectable gene, a product gene  
5 provided 3' to the selectable gene, a transcriptional regulatory region regulating transcription of both the selectable gene and the product gene, the selectable gene being positioned within an intron having a splice donor site 5' of the intron, which splice donor site regulates expression of the product gene using the transcriptional  
10 regulatory region.
2. The DNA construct of claim 1 wherein the splice donor site comprises an efficient splice donor sequence.
- 15 3. The DNA construct of claim 2 wherein the splice donor site comprises a consensus splice donor sequence.
4. The DNA construct of claim 2 wherein the splice donor site comprises the sequence GACGTAAGT.  
20
5. The DNA construct of claim 1 wherein the selectable gene is an amplifiable gene.
6. The DNA construct of claim 5 wherein the amplifiable gene is DHFR.  
25
7. The DNA construct of claim 1 wherein the transcriptional regulatory region comprises a promoter and an enhancer.
8. A vector comprising the DNA construct of claim 1.  
30
9. The vector of claim 8 wherein the selectable gene of the DNA construct is an amplifiable gene.
10. The vector of claim 8 that is capable of replication in a eukaryotic  
35 host.
11. A eukaryotic host cell comprising the vector of claim 10.
12. A eukaryotic host cell comprising the DNA construct of claim 5.  
40
13. The host cell of claim 11 wherein the vector is introduced into the host cell by electroporation.
14. A eukaryotic host cell comprising the DNA construct of claim 1  
45 integrated into a chromosome of the host cell.

15. The host cell of claim 14 that is a mammalian cell.
16. A method for producing a product of interest comprising culturing the host cell of claim 11 so as to express the product gene and recovering the product from the host cell culture.
17. The method of claim 16 further comprising recovering the product from the culture medium.
18. The method of claim 16 wherein the selectable gene is an amplifiable gene and the splice donor site comprises an efficient splice donor sequence.
19. A method for producing a product of interest comprising culturing the host cell of claim 12 so as to express the product gene in a selective medium comprising an amplifying agent for sufficient time to allow amplification to occur, and recovering the product.
20. A method for producing eukaryotic cells having multiple copies of a product gene comprising transforming eukaryotic cells with the DNA construct of claim 5, growing the cells in a selective medium comprising an amplifying agent for a sufficient time for amplification to occur, and selecting cells having multiple copies of the product gene.
21. The method of claim 20 further comprising recovering from the selected cells the product of interest.

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FIG. 1A

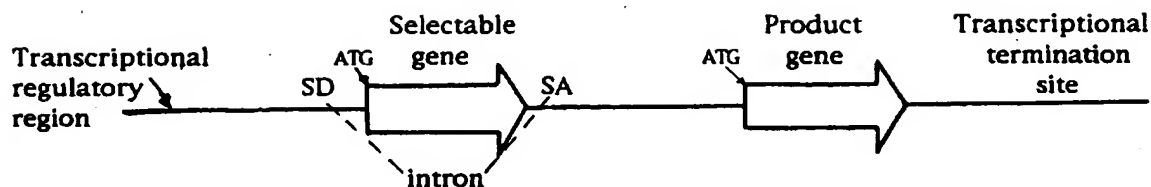


FIG. 1B

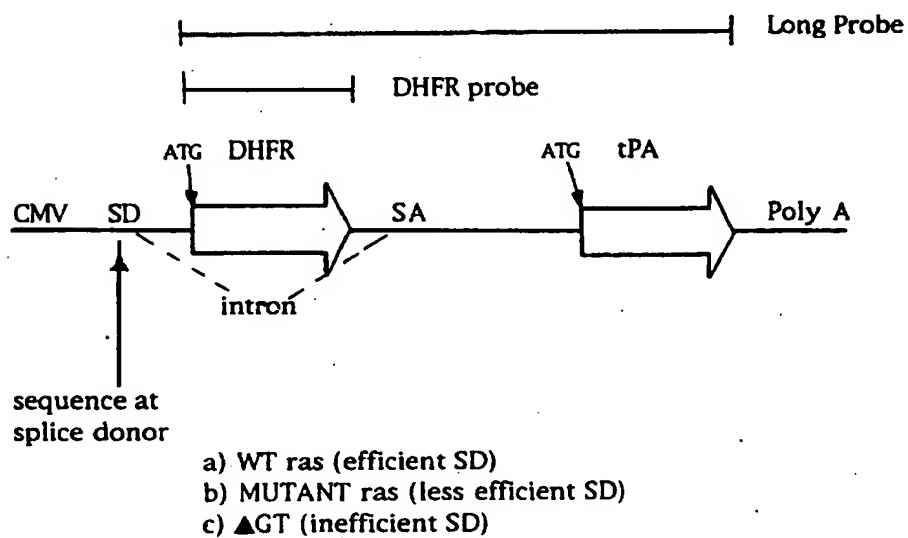
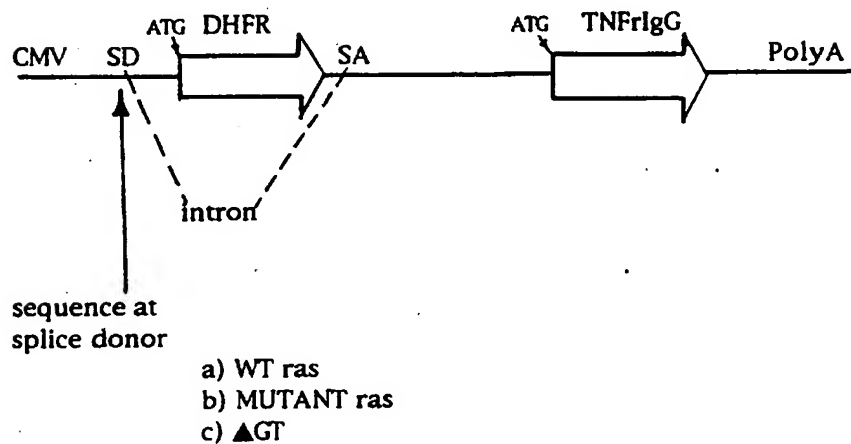


FIG. 1C



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FIG. 1D

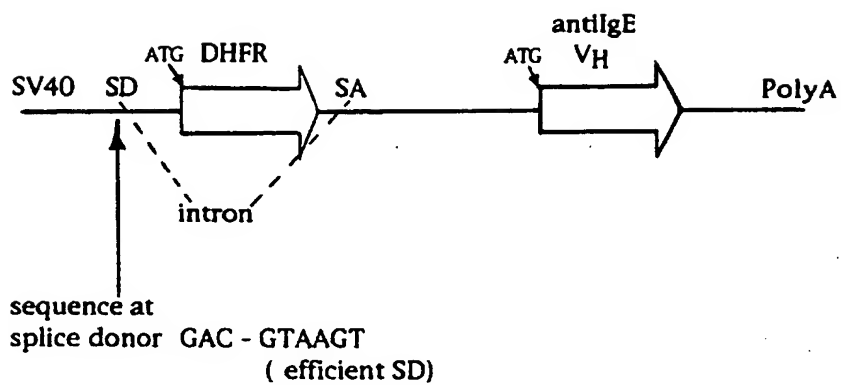
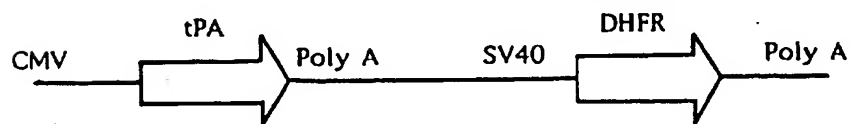


FIG. 2





[illegible]

**FIG. 3B**

401 GGTGTTGGCA GTACATCAAT GGGCGTGGAT AGCGGTTTGA CTCACGGGGA TTCCCAAGTC TCCACCCCAT TGACGTCAAT GGGAGTTTGT TTTGGCAGCA  
CCAAAACCGT CATGTAGTTA CCGGCACCTA TCGCCAAACT GAGTGCCCTT AGAGTTTCA AGTGGGTA ACTGCAGTTA CCTCAACA AAACCGTGGT

501 AAATCAACGG GACTTTCCAA AATGTCGTAA CAATCGGCC CCATGACGC AAATGGCGG TAGGCGTGA CGTGGGAGG TCTATATAAG CAGAGCTCGT  
TTAGTTGCC CTGAAAGGTT TTACAGCATT GTTGGGCGG GGTAACTGCG TTACCCGCC ATCCGCACAT GCCACCCCTC AGATATATTC GTCTCGAGCA

601 TTAGTGAACC GTCAGATCGC CTGGAGACGC CATCCACGCT GTTTGACCT CCATAGAGA CACCGGGACC GATCCAGCT CCGCGCGCGG GAACGGTGCA  
AATCACTTGG CAGTCTAGCG GACCTCTGCG GTAGGTGCGA CAAACTGGA GGTATCTTCT GTGGCCCTGG CTAGGTGCGA GCGCGCGGCC CTGCGCACGT

[illegible]

**FIG. 3D**

1101 CAAGTAAAGT AGACATGGTT TGGATAGTCG GAGGCGATTG TGGTATACCA GAGGCCATGA ATCAACCAGG CCACCTTAGA CTCTTGTGA CAAGGATCAT GTTCATTCA TCTGTACCAA ACCTATCAGC CTCGGTCAAG ACAATGGTC CTTCGGTACT TAGTTGGTCC GGTCGAATCT GAGAACACT GTTCCTAGTA

1201 GCAGGAATTT GAAAGTGACA CGTTTTCCTC AGAATTGAT TTGGGGAAT ATAAACCTCT CCCAGAAATAC CCAGCGTCC TCTCTGAGGT CCAGGAGAA CGTCCTTAAA CTTCACACTGT GCAAAAAGGG TCCTTAACTA AACCCCTTA TATTGGAGA GGGTCTTATG GGTCGCGAGG AGAGACTCCA GGTCCTCTCT

1301 AAAGGCATCA AGTATAAGTT TGAAGTCTAC GAGAAGAAAG ACTAACAGG AGATGCTTTC AAGTTCTCTG CTCCCCTCCT AAAGCTATGC ATTTTATAA TTTCGGTAGT TCATATTCAA ACTTCAGATG CTCTTCTTTC TGATTGCTCT TCTACGAAAG TTCAAGAGAC GAGGCGAGGA TTTCGATACG TAAAAATATT

1401 GACCATGGA CTTTGCTGG CTTTAGACCC CTTTGGCTTC GTTAGAACG CGCTACAATT AATACATAAC CTTATGTATC ATACACATAG ATTTAGGTGA CTGGTACCCT GAAAACGACC GAAATCTGG GGAACCGAAG CAATCTTGG CCGATGTAA TTATGTATTG GAATACATAG TATGTGTATC TAAATCCACT

**FIG. 3E**

[illegible]

SUBSTITUTE SHEET (RULE 26)

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[illegible]





**FIG. 31**

[illegible]

**FIG. 3J**

[illegible]

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## FIG. 3K

```

styI
aciI
fnu4HI sau96I
bglI nlaIIV
sfii ncoI haeIII/palI
haeIII/palI
eaeI dsal asuI
cfrI bsaJI
3601 GATGGCCGCC ATGCCCCAAC TTGTTTATTG CAGCTTATAA TGGTTACAAA TAAAGCAATA GCATCACAAA TTTCACAAAT AAAGCATTIT TTTCAGTGCA
CTACCGGCGG TACCGGGTGG AACAAATAAC GTCGAATAAT ACCAATGTTT ATTTCGTTAT CGTAGTGTIT AAAGTGTTTA TTTCGTAAAA AAAGTGACGT
bsmI

sau3AI
mboI/ndeII(dam-)
dpmI(dam+)
dpmII(dam-)
pvuI/bspCI
mcrI
taqI(dam-) tru9I
clal/bsp106(dam-)
sau3AI mseI
mboI/ndeII(dam-)
dpmI(dam+) xmnI
dpmII(dam-) aseI/asnI/vspI bsaJI
nlaIII alwI(dam-) asp700 hhaI/cfoI nlaIII
3701 TTCTAGTTGT GGTGTGTTCCA AACTCATCAA TGTATCTTAT CATGTCTGGA TCGATCGGGA ATTAATTCCG CGCAGCACCA TGGCCTGAAA TAACCTCTGA
AAGATCAACA CCAACAGGT TTGAGTAGTT ACATAGAATA GTACAGACCT AGCTAGCCCT TAATTAAGCC GCGTCGTGCT ACCGACTTT ATTGGAGACT
mnlI
maeI
nlaIIV
rsal
csp6I
nlaIIV
kpnI
hgiCI
bani
asp718 mnlI
acc65I ddeI aciI
3801 AAGAGGAACT TGGTTAGGTA CCTTCTGAGG CGGAAGAAGC CAGCTGTGGA ATGTGTGTCA GTTAGGGTGT GGAAGTCCC CAGGCTCCCC AGCAGGCGCA
TTCTCCTTGA ACCAATCCAT GGAAGACTCC GCCTTCTTGT GTGACACCT TACACACAGT CAATCCACA CCTTTCAGGG GTCCGAGGGG TCGTCCGTCT
bsaJI
nlaIIV
scrFI
mvaI
ecorII
dsav
bstNI
apyI(dcm+)
bsaJI

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[illegible]

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## FIG. 3M

hinPI  
 hhai/cfoI  
 nlaIV  
 nari  
 kasi  
 hinII/acyI  
 hgiCI  
 haeII  
 bani  
 sfaNI  
 acil  
 sfaNI  
 acil  
 bglI  
 sau3AI  
 sau96I  
 mboI/ndeII[dam-]  
 haeIII/pali  
 asuI  
 mnII  
 mboII  
 aciI  
 earI/ksp632I  
 mcrI  
 4301 CGAAGAGGCC CGCACCAGTC GCCCTTCCCA ACAGTTGCGT AGCCTGAATG GCGAATGGCG CTGATGCGG TATTTCTCC TTACGCATCT GTGCGGTATT  
 GCTTCTCCGG CGGTGGCTAG CGGGAAGGGT TGTCAACGCA TCGGACTTAC CGCTTACCGC GGACTACGCC ATAAAGAGG AATGCGTAGA CAGCCATAA  
 aciI  
 fnu4HI  
 hinPI  
 thai  
 fnuDII/mvni  
 bstUI  
 bsh1236I  
 rsal  
 hhai/cfoI  
 fnu4HI  
 tru9I  
 aciI  
 msei  
 bsh1236I  
 4401 TCACACCGCA ATGCCTAAAG CAACCATAGT AGCGCCCTG TAGCGCGCA TTAAGCGCG CGGTGTGTG GTTACGCG AGCGTGACCG CTACACTTGC  
 AGTGTGGCGT ATGCAGTTTC GTTGTATCA TCGCGGGGAC ATCGCGCGGT AATTCGCGC GCCACACCA CCAATGCGG TCGCACTGGC GATGTGAACG  
 hinPI  
 hhai/cfoI  
 rmaI  
 hinPI  
 haeII  
 hhai/cfoI  
 bsrBI  
 haeII  
 maeI  
 aciI  
 mboII  
 4501 CAGCGCCTA CGCGCGGCTC CTTTCGCTT CTTCCCTTCC TTTCGCGCA CGTTCGCGG CTTTCCCGT CAAGCTCTAA ATCGGGGGCT CCCTTTAGG  
 GTCGGGGAT CGCGCGGCGAG GAAAGCGAAA GAAGGAAGG AAGAGCGGT GCAAGCGGC GAAAGGGGCA GTTCGAGATT TAGCCCCGA GGGNAATCCC  
 nlaIV  
 hgiCI  
 taqI  
 bani  
 mnII  
 hphI  
 maeII  
 haeIII/pali  
 draIII  
 sau96I  
 bsalI  
 asuI  
 4601 TTCCGANTTA GTGCTTTACG GCACCTCGAC CCCAAAAAC TTGATTGGG TGATGGTTCA CGTAGTGGC CATGCCCTG ATAGACGTT TTTCGCCCTT  
 AAGGCTAAAT CACGAAATGC CGTGGAGCTG GGGTTTTTG AACTAAACCC ACTACCAAGT GCATCACCCG GTAGCGGAC TATCTGCCA AAGCGGGAA  
 maeII  
 pleI  
 tru9I  
 msei  
 hinfi  
 4701 TGACGTGGA GTCCAGTTC TTTAATAGTG GACTCTGTT CCAACTGGA ACAACACTCA ACCCTATCTC GGGCTATTCT TTTGATTTAT AAGGATTTT  
 ACTGCAACCT CAGGTGCAAG AAATTATCAC CTGAGACAA GGTGTGACCT TGTGTGAGT TGGGATAGAG CCCGATAGA AACTAATAA TTCCCTAAA



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## FIG. 30

```

nlaIV
aciI
thai
fnuDI1/mvni
bstUI
bsh1236I
hinPI
hhaI/cfoI

5201 CTTTTCGGG AAATGTGGC GGAACCCCTA TTTGTTTATT TTTCTAATA CATTCAATA TGTATCCGCT CATGAGCAA TAACCTGAT AAATGCTTCA
GAAAAGCCCC TTACACGCG CCTTGGGAT AAACAATAA AAAGATTTAT GTAAGTTTAT ACATAGCGCA GTACTCTGTT ATTGGGACTA TTACGAAAGT

rcaI
bspHI
bstBI bsmAI
aciI nlaIII

fnu4HI
aciI

5301 ATAATATTGA AAAAGGAAGA GTATGAGTAT TCAACATTTC CGTGTCGCC TTATTCCTT TTTTGGGCA TTTTGCCTC CTGTTTTGC TCACCCAGAA
TATTATAACT TTTTCTTCT CATACTCATA AGTTGTAAAG GCACAGCGG AATAAGGAA AAAACGCCGT AAAACGGAAG GACAAAACG AGTGGGTCTT

hgiAI/aspHI
bsp1286
sau3AI bsiHKAI
mboI/ndeII(dam-)
dpmI(dam+) bmyI
dpmII(dam-)
eco57I
apaLI/snoI

hphI
sfaNI mboII(dam-) alw4I/snoI maeIII taqI alwI(dam-) aciI bstYI/xhoII
5401 ACGCTGGTGA AAGTAAAGA TGCTGAAGAT CAGTTGGGTG CACGAGTGG TTACATCGAA CTGGATCTCA ACACGGTAA GATCCTTGAG AGTTTTGCGC
TGGGACCACT TTCATTTTCT ACGACTTCTA GTCAACCCAC GTGCTCACCC AATGTAGCTT GACCTAGAGT TGTCGCCAT TGTCGCCAT TCAGGAAC TCAGGAACGGG

scrFI
nciI
mspi
hpaII
dsav
hinII/acyI
hgaI cauli
ahaII/bsaHI
bcgI mcrI fnu4HI
aciI

5501 CCGAAGAAG TTTTCCAATG ATGAGCACTT TTAAGTTCT GCTATGTGC GCGGTATTAT CCCGTGATGA CGCCGGGCAA GAGCAACTCG GTCCGCGCAT
GGCTTCTTG AAAAGGTTAC TACTCGTGAA AATTCAAGA CGATACACCG CGCCATAATA GGCCTACTT CCGGCCGTT CTGCTTGAGC CAGCCGCGTA

maeII
psp1406I
xmnI
asp700
mboII

rsai
csp6I bsri
scaI hphI maeIII
sfaNI foki nlaIII
fnu4HI
bbvI

5601 ACACATATTCT CAGAATGACT TGGTTGAGTA CTCACCAAGT ACAGAAAAGC ATCTTACGGA TGGCATGACA GTAAGAGAAAT TATGCACTGC TGCCATAACC
TGTGATAAGA GTCTTACTGA ACCAACTCAT GAGTGGTTCAG TGTCTTTTCG TAGAATGCGT ACCGTACTGT CATTCTCTTA ATAGTCAGC ACGTATTGG

```

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## FIG. 3P

sau96I  
 avaiI  
 sau3AI asuI  
 mboI/ndeII(dam-)  
 dpnI(dam+)  
 dpnII(dam-)  
 pvuI/bspCI  
 mcrI mnlI  
 aciI  
 haeIII/palI  
 eaeI  
 cfrI  
 fnu4HI  
 aluI  
 sau3AI nlaIV  
 mboI/ndeII(dam-) aluI  
 dpnI(dam+) hpaII  
 dpnII(dam-) bsaWI  
 5701 ATGAGTGATA AACTGGCGC CAACTTACTT CTGACAACGA TCGGAGGACC GAAGGAGCTA ACCGCTTTTT TGCACAACAT GGGGGATCAT GTAACTCGCC  
 TACTCACTAT TGTGACGCCG GTTGAATGAA GACTGTGTCT AGCCTCTCTGG CTTCCTCGAT TGGCGAAAAA ACGTGTTGTA CCCCCTAGTA CATTGAGCGG  
 mspI  
 sau3AI nlaIV  
 mboI/ndeII(dam-) aluI  
 dpnI(dam+) hpaII  
 dpnII(dam-) bsaWI  
 5801 TTGATCGTTG GGAACCGGAG CTGAATGAAG CCATACCAA CGACGAGCGT GACACCACGA TGCCAGCAGC AATGGCAACA AGTTGCGCA AACTATTAAAC  
 AACTAGCAAC CCTTGCCTC GACTTACTTC GGTATGGTTT GCTGTCTGCA CTGTGGTGCT ACGTCTGCTG TTACCGTTGT TGCAACGGCT TTGATAATTG  
 mspI  
 hpaII  
 scrFI  
 aluI nciI  
 rnaI dsav  
 maeI cauII  
 5901 TGGCGAACTA CTTACTCTAG CTTCCCGGCA ACAATTAATA GACTGGATGG AGCGGATAA AGTTGCAGGA CCACCTCTGC GCTCGGCCCT TCCGGCTGGC  
 ACCGCTTGAT GAATGAGATC GAAGGCGCGT TGTTAATTAT CTGACCTACC TCCGCTATT TCAACGTCCT GGTGAAGACG CGAGCCGGA AGGCCGACCG  
 mspI  
 hpaII  
 cfr10I  
 nlaIV hphI  
 gsuI/bpmI  
 6001 TGGTTTATTG CTGATAAATC TGGAGCCGGT GAGCGTGGGT CTGCGGGTAT CATTCAGCA CTGGGCCAG ATGGTAAGCC CTCCCGTATC GTAGTATCT  
 ACCAAATAAC GACTATTTAG ACCTCGGCA CTGCGACCCA GAGCCGATA GTAACGTCGT GACCCCGTC TACCATTCCG GAGGGCATAG CATCAATAGA  
 mspI  
 hpaII  
 cfr10I  
 nlaIV hphI  
 gsuI/bpmI  
 6101 ACACGACGGG GAGTCAGGCA ACTATGGATG AACGAATAG ACAGATCGGT GAGATAGGTG CCTCACTGAT TAAGCATTGG TAACTGTGAC ACCAAGTTTA  
 TGTGCTGCC CTCAGTCCGT TGATACCTAC TTGCTTTATC TGTCTAGCGA CTCATCCAC GGAGTGACTA ATTCGTAACC ATTGACAGTC TGGTTCAAAT



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## FIG. 3Q

```

rmaI      sau3AI
sau3AI hphI mboI/ndeII(dam-)
mboI/ndeII(dam-)
dpmI(dam+) dpmI(dam+)
dpmII(dam-) dpmII(dam-)
tru9I bstYI/xhoII alwI/dam- nlaIII maeII
mseI alwI(dam-) bstYI/xhoII rcaI tru9I
ahaIII/draI mseI mboII(dam-) bspHI mseI
6201 CTCATATATA CTTTATGATTG ATTTAAAGGA TCTAGGTGAA GATCCTTTT GATAATCTCA TGACCAAAAT CCTTAACGT
GAGTATATAT GAAATCTAAC TAAATTTTGA AGTAAATTTT AGATCCACTT CTAGGAAAAA CTATTAGAGT ACTGGTTTTTA GGGAAATTGCA

sau3AI
mboI/ndeII(dam-)
dpmI(dam+) sau3AI
dpmII(dam-) mboI/ndeII(dam-) thaI fnuDII/mvnI
bstYI/xhoII dpmI(dam+) bstUI
sau3AI alwI(dam-) dpmII(dam-) bsh1236I
mboI/ndeII(dam-) alwI(dam-) hinPI fnu4HI
dpmI(dam+) mboII(dam-) bstYI/xhoII hhaI/cfoI bbvI
dpmII(dam-)
6301 GAGTTTTCGT TCCACTGAGC GTCAGACCCC GTAGAAAGA TCAAGAGATC TTCTTTGAGAT CCTTTTTC TGCGCGTAAT CTGCTGCTTG CAAACAAAAA
CTCAAAAGCA AGGTGACTCG CAGTCTGGG CATCTTTTCT AGTTTCTTAG AAGAACTCTA GGAATAAAAG ACGCCGATTA GACGACGAAC GTTGTGTTTTT

sau3AI
mboI/ndeII(dam-)
dpmI(dam+)
dpmII(dam-)
alwI(dam-)
mspI
aciI nspBII hpaII aluI bsrI hinPI
aciI AACCAACCGT ACCAGCGGTG GTTGTGTTGC CGGATCAAGA GCTACCAACT CTTTTTCCGA AGGTAACTGG CTTCAGCAGA GCGCAGATAC CAAATACTGT
TTGGTGCGCA TGGTCGCCAC CAAACAAACG GCCTAGTTCT CGATGGTTGA GAAAAGGCT TCCATTGACC GAAGTCGTCT CGCGTCTATG GTTTATGACA
6401

haeIII/palI
rmaI haeI bslI
maeI
6501 CCTTCTAGTG TAGCCGTAGT TAGGCCACCA CTTCAAGAAC TCTGTAGCAC CGCTACATA CCTCGCTCTG CTAATCTGTG TACCACTGGC TGCTGCCAGT
GGAAGATCAC ATCGGCATCA ATCCGGTGGT GAAGTTCTTG AGACATCGTG GCGGATCTAT GGAGCGAGAC GATTAGGACA ATGCTCACCG ACGACGGTCA

```

**FIG. 3R**

66601	GGCGATAAGT CGTGTCTTAC CGGGTTGGAC TCAAGACGAT AGTTACCGGA TAAGCGCGAG CGGTGGGCT GAACGGGGG TTCTGTGCACA CAGCCAGCT CCGCTATTCA GCACAGAATG GCCCAACCTG AGTTCTGCTA TCAATGGCTT TCAATGGCTT GCCAGCCCGA CTTGCCCCCC GTTGGGTCTG	acii nspBII fnu4HI bbvI hinPI mcrI hhaI/cfoI maeIII pleI hinfi cauII	mspi hpaII bsaWI maeIII bsaWI maeIII pleI hinfi cauII	apalI/snoI alw44I/snoI	mspi hpaII bsli bsaWI aciI bsaWI aciI	fnu4HI bsli bsaWI aciI	
6701	TGGAGCGAAC GACCTACACC GAACCTAGAT ACCTACAGCG TGAGCATTTGA GAAGCGCCA CGCTTCCGA AGGAGAAAG CCGACAGGT ATCCGCTAAG ACCTCGCTTG CTGGATGTTG CTGTGACTCTA TGGATGTGCG ACTCGTAAT CTTCGCGGT GCGAAGGGCT TCCCTCTTTC CCGCTGTCCA TAGGCCATTC	ddeI scfi mval ecoRII dsav bstNI bsaJI hinPI mnlI hhaI/cfoI aluI apyI[dcM+] apYI[dcM+] CCTGTGATCT TTATAGTCTT GTCCGGTTTC GCCACCTCTG ACTTCAGCGT	mspi hpaII bsaWI maeIII bsaWI maeIII pleI hinfi cauII	mspi hpaII bsli bsaWI aciI bsaWI aciI	apalI/snoI alw44I/snoI	mspi hpaII bsli bsaWI aciI bsaWI aciI	fnu4HI bsli bsaWI aciI
6801	CGGCAGGGTC GGAACAGGAG AGCGACGAG GGAGCTTCCA GGGGAAACG CCTGTGATCT TTATAGTCTT GTCCGGTTTC GCCACCTCTG ACTTCAGCGT GCGGTCCCAG CCTTGTCTTC CTCTGAGGCT CCTCGAAGGT CCCCCTTTTC GGACCATAGA AATATCAGGA CAGCCCAAAG CCGTGGAGAC TGAACTCGCA	scrFI mval ecoRII dsav bstNI bsaJI hinPI mnlI hhaI/cfoI aluI apyI[dcM+] apYI[dcM+] CCTGTGATCT TTATAGTCTT GTCCGGTTTC GCCACCTCTG ACTTCAGCGT	mspi hpaII bsaWI maeIII bsaWI maeIII pleI hinfi cauII	mspi hpaII bsli bsaWI aciI bsaWI aciI	apalI/snoI alw44I/snoI	mspi hpaII bsli bsaWI aciI bsaWI aciI	fnu4HI bsli bsaWI aciI
6901	CGATTTTGT GATGCTCGTC AGGGGGCGCG AGCCTATGGA AAAACGCCAG CAACGGGCG CCTTTTACGGT TCCTGGCCCT TTGCTGGCTT TTTGCTCACA GCTAAAAACA CTACGAGCAG TCCCGCCGCG GTTGGCGGTC TCGGATACCT TTTTGGCGTC GGTGGCGGAA AGGACCCGGA AACGAGTGT	sfaNI aciI nlaIV bsaJI hinPI mnlI hhaI/cfoI aluI apyI[dcM+] apYI[dcM+] CCTGTGATCT TTATAGTCTT GTCCGGTTTC GCCACCTCTG ACTTCAGCGT	mspi hpaII bsaWI maeIII bsaWI maeIII pleI hinfi cauII	mspi hpaII bsli bsaWI aciI bsaWI aciI	apalI/snoI alw44I/snoI	mspi hpaII bsli bsaWI aciI bsaWI aciI	fnu4HI bsli bsaWI aciI
7001	TGTTCTTTCC TCGGTTATCC CCTGATCTG TGGATAACCG TATTACCGCC TTGAGTAGAG CTGATACCGC TCGCCGCGAG CGAACGACCG AGCGCAGCA ACAAGAAAG ACGCAATAGG GGACTAAGAC ACCTATTGCG ATAAATGGCG AAATCACTC GACTATGGCG AGCGCGTGC GTTGTCTGCG TCGGCTCGT	scrFI mval ecoRII dsav bstNI bsaJI hinPI mnlI hhaI/cfoI aluI apyI[dcM+] apYI[dcM+] CCTGTGATCT TTATAGTCTT GTCCGGTTTC GCCACCTCTG ACTTCAGCGT	mspi hpaII bsaWI maeIII bsaWI maeIII pleI hinfi cauII	mspi hpaII bsli bsaWI aciI bsaWI aciI	apalI/snoI alw44I/snoI	mspi hpaII bsli bsaWI aciI bsaWI aciI	fnu4HI bsli bsaWI aciI



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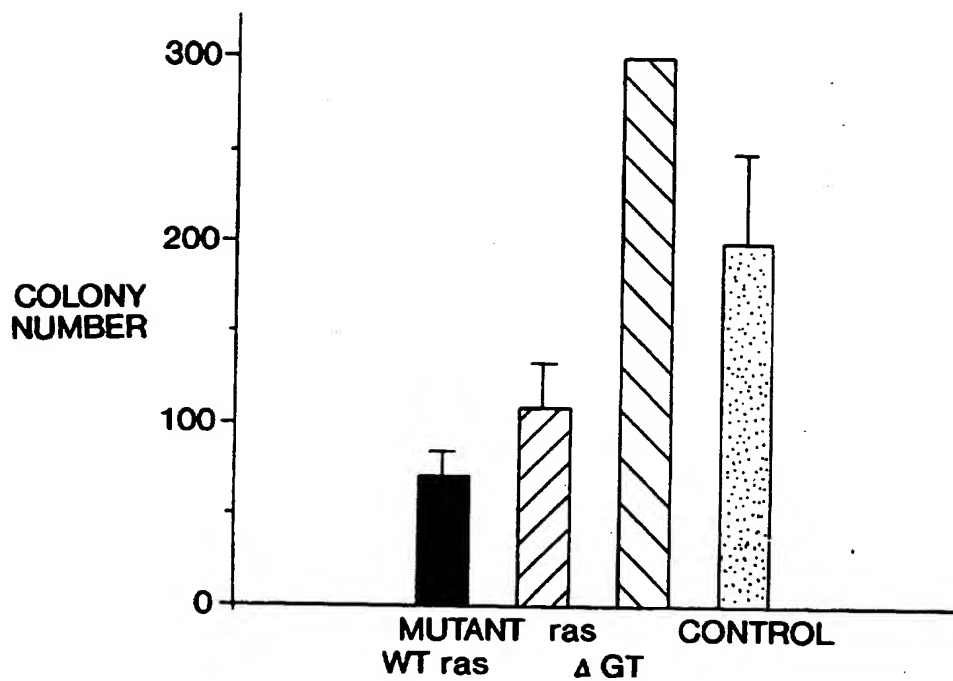


FIG. 4

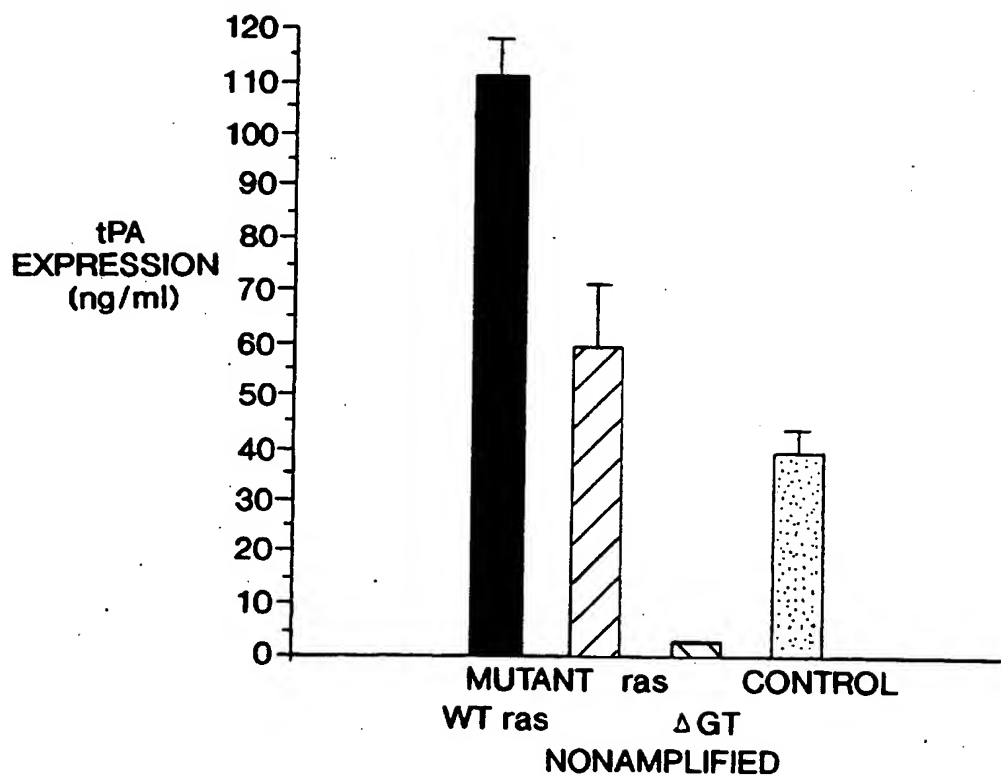
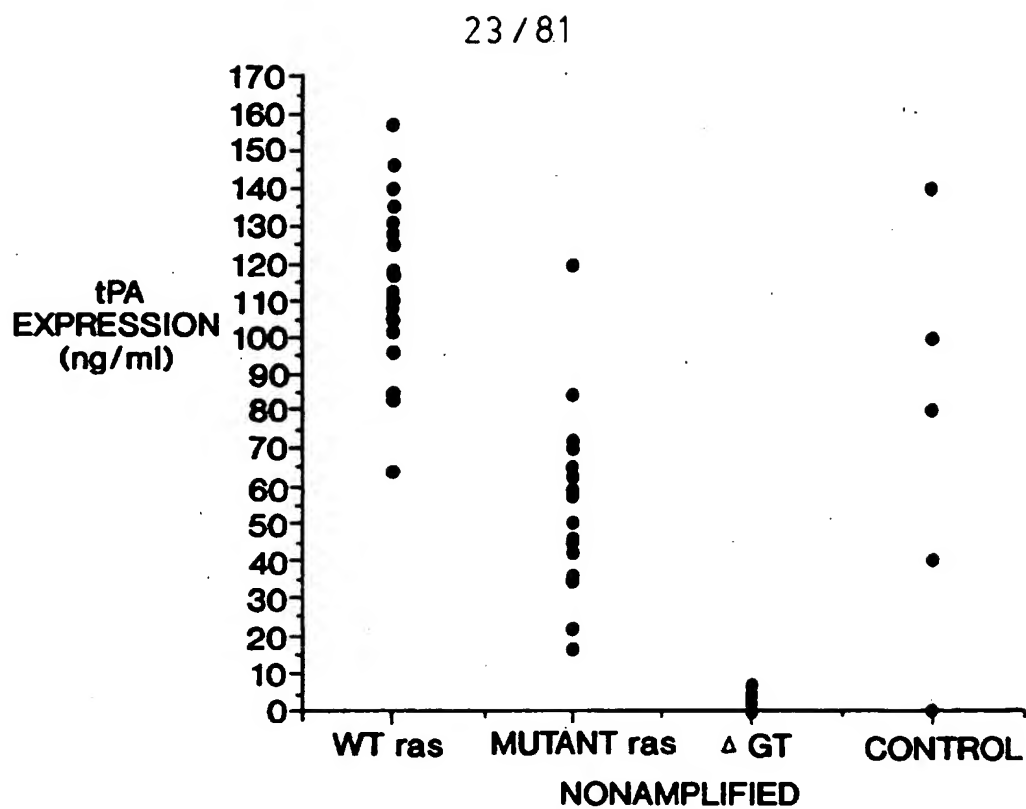
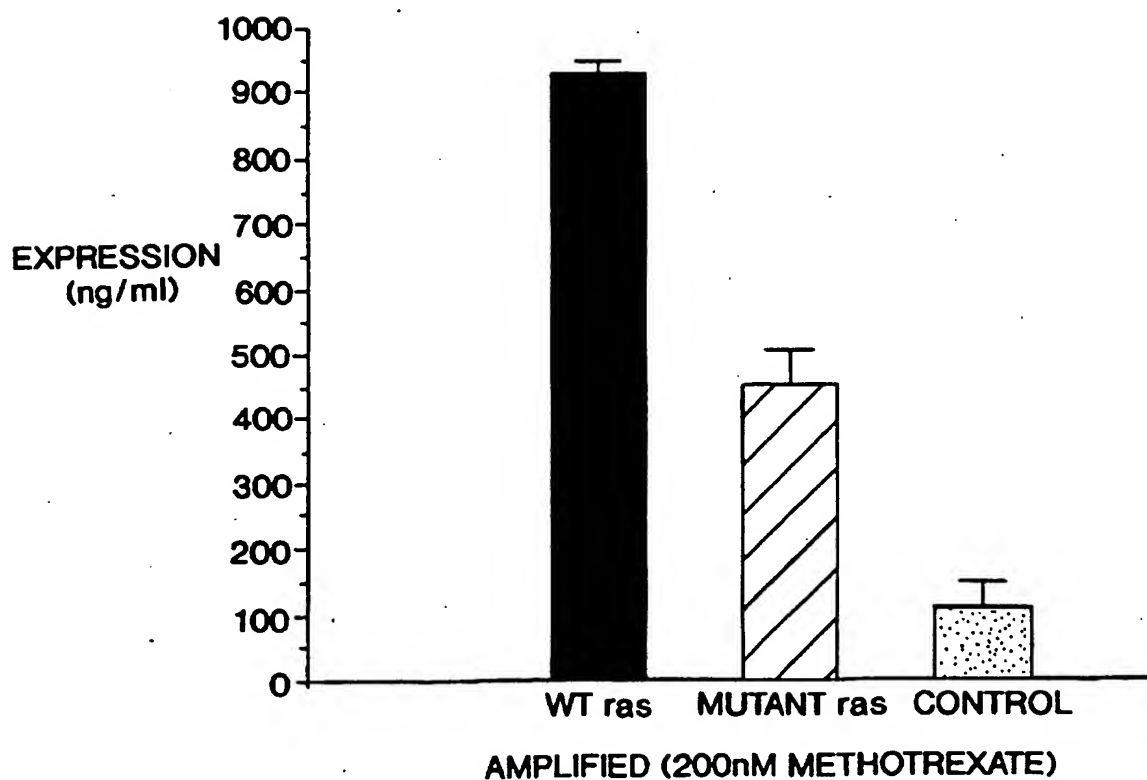


FIG. 5A

**FIG. 5B****FIG. 5C**

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## FIG. 6A

```

alul
sstI
sacI
hgiJII
hgiAI/asphi
ecII136II
bsp1286
bsiHKA1
bmyI
banII
taqI
1 TTCGAGCTCG CCGGACATTG ATTATTGACT AGTTATTAAAT AGTAATCAAT TACGGGGTCA TTAGTTTATA GCCCATATAT GGAGTTCGGC GTTACATAAC
AAGTCGAGC GGGCTGTAA C TAATACTGA TCATAAATTA TCATTAGTTA ATGCCCCAGT AATCAAGTAT CGGTATATA CTCAAGGCG CAATGTATTG

          rmaI   tru9I
          maeI   mseI
          speI   asel/asnI/vspI
          bslI
          acII maeIII
          bsh1236I
          fnuDII/mvnI
          bstUI
          thaI

          scrFI
          mvaI
          ecorII
          dsav
          aciI
          bglI bstNI
          sau96I
          haeIII/palI aciI
          asuI apyI[dcM+]
101 TTACGGTAA TGGCCCGCCT GGCTGACCG CCAAGGACCC CGGCCCATTTG ACCTCAATAA TGACGTATGT TCCCATAGTA AGCCATAG AGACTTTCCA
AATGCCATT ACCGGCGGA CCGACTGCG GGTGCTGG GCGGGTAA C TGCAGTTATT ACTGCATACA AGGTATCAT TCGGTATC CCGAAAGGT

          maeII
          hinII/acyI
          ahaII/bsaHI
          aatII
          maeIII
          maeII
          hinII/acyI
          ahaII/bsaHI
          aatII
          rsaI
          csp6I
          ndeI
          bglI
          rsaI
          csp6I
201 TTACGTCAA TGGGTGGAGT ATTACGTA AACTGCCAC TTGGCAGTAC ATCAAGTGA TCATATGCCA AGTACGCCC CTATTGACGT CAATGACGCT
AACTGAGTT ACCACCTCA TAAATGCCAT TTGACGGGTG AACCGTCATG TAGTTCACAT AGTATACGCT TCATGCCGGG GATACTGCA GTTACTGCCA

          scrFI
          mvaI
          ecorII
          aciI
          bglI dsav
          sau96I bstNI
          haeIII/palI
          asuI apyI[dcM+] bsrI nlaIII
          rsaI
          csp6I
          maeII
          snbI
          bsaI
          csp6I
          nlaIII
          styI
          ncoI
          dsaI hphI aciI
          bsaJI sfaNI
301 AAATGGCCC CCTGGCATT TGGCCAGTAC ATGACCTTAT GGGACTTTCC TACTTGGCAG TACATCTAGC TATTAGTCAT CGCTATTACC ATGGTGATGC
TTTACCGGC GACCCGTAAT ACGGTTCATG ACGGAAAGG ATGAACCGTC ATGTAGATGC ATAATCAGTA GCGATATGG TACCACCTAGC

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## FIG. 6B

401 GGTTTTGGCA GTACATCAAT GGGCGTGGAT AGCGGTTTGA CTCACGGGGA TTTCCAAGTC TCCACCCCAT TGAGTCAAT GGGAGTTTGT TTTGGCACCA  
 CCAAAACCGT CATGTAGTTA CCGGCACCTA TCGCCAAACT GAGTCCCCCT AAAGTTTCAG AGGTGGGTA ACTGCAGTTA CCTCAACA AAACCGTGT

501 AAATCAACGG GACITTTCCAA AATGTGTA CAACCTCCGCC CCAITGACGC AAATGGCGG TAGGCGTGT TAATGGGAGG TCTATATAAG CAGAGCTCGT  
 TTTAGTTGCC CTGAAGGTT TTACAGCAAT GTTGAGGCGG GGTAACTGCG TTTACCCGCC ATCCGCACAT GCCACCCTCC AGATATATTC GTCTCGACCA

601 TTAGTGAACC GTCAGATCC CTGGAGACGC CATCCACGCT GTTTTGACCT CCATAGAAGA CACCGGACCT GATCCAGCCT CCGCGGCCGG GAACGGTGCA  
 AATCACTTGG CAGCTAGG GACCTCTGCG GTAGGTGCGA CAAACTGGA GGTATCTTCT GTGGCCCTGG CTAGGTGCGA GGGCCCGGCC CTGCCCACGT

Restriction Enzymes and Sites:  
 rsal csp6I maeIII acII acII hgaI hgaI acII acII csp6I mmlI  
 pleI hinfI acII hinfI bsmAI  
 nlaIV nlaIV nlaIV nlaIV nlaIV nlaIV nlaIV nlaIV nlaIV nlaIV  
 hgiII hgiII hgiII hgiII hgiII hgiII hgiII hgiII hgiII hgiII  
 eelIII eelIII eelIII eelIII eelIII eelIII eelIII eelIII eelIII eelIII  
 bsp1286 bsp1286 bsp1286 bsp1286 bsp1286 bsp1286 bsp1286 bsp1286 bsp1286 bsp1286  
 bsiHRAI bsiHRAI bsiHRAI bsiHRAI bsiHRAI bsiHRAI bsiHRAI bsiHRAI bsiHRAI bsiHRAI  
 bmyI bmyI bmyI bmyI bmyI bmyI bmyI bmyI bmyI bmyI  
 banII banII banII banII banII banII banII banII banII banII  
 haeIII/palI haeIII/palI haeIII/palI haeIII/palI haeIII/palI haeIII/palI haeIII/palI haeIII/palI haeIII/palI haeIII/palI  
 mcrI mcrI mcrI mcrI mcrI mcrI mcrI mcrI mcrI mcrI  
 eaeI eaeI eaeI eaeI eaeI eaeI eaeI eaeI eaeI eaeI  
 cfrI cfrI cfrI cfrI cfrI cfrI cfrI cfrI cfrI cfrI  
 fnu4HI fnu4HI fnu4HI fnu4HI fnu4HI fnu4HI fnu4HI fnu4HI fnu4HI fnu4HI  
 acII acII acII acII acII acII acII acII acII acII  
 tbaI tbaI tbaI tbaI tbaI tbaI tbaI tbaI tbaI tbaI  
 fnuDII/mvnI fnuDII/mvnI fnuDII/mvnI fnuDII/mvnI fnuDII/mvnI fnuDII/mvnI fnuDII/mvnI fnuDII/mvnI fnuDII/mvnI fnuDII/mvnI  
 sacII/sstII sacII/sstII sacII/sstII sacII/sstII sacII/sstII sacII/sstII sacII/sstII sacII/sstII sacII/sstII sacII/sstII  
 nspBII nspBII nspBII nspBII nspBII nspBII nspBII nspBII nspBII nspBII  
 kspI kspI kspI kspI kspI kspI kspI kspI kspI kspI  
 dsal dsal dsal dsal dsal dsal dsal dsal dsal dsal  
 bgII bgII bgII bgII bgII bgII bgII bgII bgII bgII  
 sau3AI sau3AI sau3AI sau3AI sau3AI sau3AI sau3AI sau3AI sau3AI sau3AI  
 mboI mboI mboI mboI mboI mboI mboI mboI mboI mboI  
 dpmI dpmI dpmI dpmI dpmI dpmI dpmI dpmI dpmI dpmI  
 dpmI dpmI dpmI dpmI dpmI dpmI dpmI dpmI dpmI dpmI  
 alwI alwI alwI alwI alwI alwI alwI alwI alwI alwI  
 cauII cauII cauII cauII cauII cauII cauII cauII cauII cauII  
 sau3AI sau3AI sau3AI sau3AI sau3AI sau3AI sau3AI sau3AI sau3AI sau3AI  
 mboI mboI mboI mboI mboI mboI mboI mboI mboI mboI  
 dpmI dpmI dpmI dpmI dpmI dpmI dpmI dpmI dpmI dpmI  
 dpmI dpmI dpmI dpmI dpmI dpmI dpmI dpmI dpmI dpmI  
 alwI alwI alwI alwI alwI alwI alwI alwI alwI alwI  
 cauII cauII cauII cauII cauII cauII cauII cauII cauII cauII  
 sau3AI sau3AI sau3AI sau3AI sau3AI sau3AI sau3AI sau3AI sau3AI sau3AI  
 mboI mboI mboI mboI mboI mboI mboI mboI mboI mboI  
 dpmI dpmI dpmI dpmI dpmI dpmI dpmI dpmI dpmI dpmI  
 dpmI dpmI dpmI dpmI dpmI dpmI dpmI dpmI dpmI dpmI  
 alwI alwI alwI alwI alwI alwI alwI alwI alwI alwI  
 cauII cauII cauII cauII cauII cauII cauII cauII cauII cauII

## FIG. 6C

```

      tfil
      acil
      thal hinfI
      fnuDII/mvml
      bstUI
      bsh1236I
701 TTGGAACGCG GATTCCCGT GCCAAGAGTG CTGTAAAGTAC CGCCTATAGA CGGATAAGAG GATTTATCC CGCTGCCAT CATGGTTCGA CCATTGAAC
      AACCTTGCG CTAAGGGCA CGGTCTCAC GACATTATG CGGATAATCT CGCTATTCTC CTAAATAGG CCGCAGCGTA GTACCAAGCT GGTAACCTGA
      fnu4HI
      bbvI
      nspBII
      acil
      nlaIII
      taqI
      thal
      fnuDII/mvml
      bstUI
      bsh1236I
      mlui
      barBI
      afIII
      rsal
      csp6I
      acil
      xmiI
      ssp700
      scaI
801 GCATCGTCCG CGTGTCCTCA AATATGGGA TTGGCAAGAA CGGAGACCTA CCTGCCCTC CGCTCAGAA CGGTTCAAG TACTTCCAA GAATGACCAC
      CGTAGCAGCG GCACAGGCTT TTATACCCCT AACCGTTCTT GCCTCTGGAT GCGACGGGAG CCGAGTCTT CCGCAAGTTC ATGAAGGTTT CTACTGGTG
      pflMI
      bslI
      bsmAI
      bsal
      mnlI
      ddel
      asp700
      scaI
      scrFI
      mvaI
      ecorII
      dsav
      bstNI
      apyI(dcm+)
      sexAI
      tfil
      hinfI
      hphI
      alwNI
      mboII
      taqI
      msel
      tru9I
      msel
      asel/asnI/vspI
901 AACCTCTTCA GTGGAAGTA AACAGAATCT GGTGATTATG GTAGGAAA CCTGTTCTC CATTCCTGAG AAGAATCGAC CTTAAAGGA CAGAATTAT
      TTGGAGAAGT CACCTTCCAT TTGTCTTAGA CCACTAATAC CCATCTTTT GGACCAAGAG GTAAGGACTC TTCTAGCTG GAAATTTCTT GTCTTAATTA
      aluI
      sstI
      sacI
      hgiJII
      hgiAI/asphI
      eclI36II
      bsp1286
      bsiHKA
      bmyI
      banII
      mnlI
      bslI
      ddel
      bstXI
      foki
      sfaNI
      msel
      afIII/bfrI
      bsaWI
      mspl
      hpaII
1001 ATAGTTCTCA GTAGAGAAGT CAAAGAACCA CCACGAGGAG CTCATTTTCT TGCCAAAAGT TTGGATGATG CCTTAAGACT TATTGAACAA CCGGAATTGG
      TATCAAGAGT CATCTCTTGA GTTCTTGGT GGTGCTCTC GAGTAAAGA ACGGTTTTCA AACCTACTAC GGAATCTGA ATACTTGT GGCCTTAACC

```



## FIG. 6D

FIG. 6D

1101 CAAGTAAAGT AGACATGGTT TGGATAGTCG GAGGCAGTTC TGTTACCAG GAAGCCATGA ATCAACCAGG CCACCTTAGA CTCTTTGTA CAAGGATCAT GTTCATTCA TCTGTACCA ACCTATCAGC CTCCGTCAAG ACAATGGTC CTTCGGTACT TAGTTGGTCC GGTGGAATCT GAGAACACT GTTCCTAGTA

11201 GCAGGAATTT GAAGTGACA CGTTTTTCCC AGAAATGAT TTGGGGAAT ATAACTCTT CCCAGAATAC CCAGGCGTCC TCTCTGAGGT CCAGGAGGAA CGTCCTTAAA CTTTCACTGT GCAAAAAGGG TCTTTAACTA AACCCTTTA TATTTGGAGA GGGTCTTATG GGTCCGACG AGAGACTCCA GGTCTCTCTT

1401 GACCATGGGA CTTTGTCTGG CTTTAGACCC CTTTGGCTTC GTTAGAACGC GGCTACAAAT AATACATAAC CTTATGTATC ATACACATAG ATTTAGGTGA CTGGTACCCT GAAAACGACC GAATCTGGG GGAACCGAAG CAATCTTGG CCGATGTTAA TTATGTATTG GAATACATAG TATGTATC TAATCCACT

Restriction Enzymes:

haeIII/palI  
haeI  
scrFI  
mvaI  
ecoRII  
dsav  
bstNI  
nlaIII  
mnlI  
accI  
nlaIII  
apoI  
maeIII  
aflIII  
maeII  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
fnu4HI  
acII  
thaI  
fnuDII/mvnI  
tru9I  
bstUI  
msei  
bsaJI  
bsh1236I  
aseI/asnI/vspI  
maeIII  
hphI  
maeIII

Restriction Enzymes:

scrFI  
mvaI  
ecoRII  
dsav  
bstNI  
nlaIII  
mnlI  
accI  
nlaIII  
apoI  
maeIII  
aflIII  
maeII  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
fnu4HI  
acII  
thaI  
fnuDII/mvnI  
tru9I  
bstUI  
msei  
bsaJI  
bsh1236I  
aseI/asnI/vspI  
maeIII  
hphI  
maeIII

Restriction Enzymes:

haeIII/palI  
haeI  
scrFI  
mvaI  
ecoRII  
dsav  
bstNI  
nlaIII  
mnlI  
accI  
nlaIII  
apoI  
maeIII  
aflIII  
maeII  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
fnu4HI  
acII  
thaI  
fnuDII/mvnI  
tru9I  
bstUI  
msei  
bsaJI  
bsh1236I  
aseI/asnI/vspI  
maeIII  
hphI  
maeIII

Restriction Enzymes:

haeIII/palI  
haeI  
scrFI  
mvaI  
ecoRII  
dsav  
bstNI  
nlaIII  
mnlI  
accI  
nlaIII  
apoI  
maeIII  
aflIII  
maeII  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
fnu4HI  
acII  
thaI  
fnuDII/mvnI  
tru9I  
bstUI  
msei  
bsaJI  
bsh1236I  
aseI/asnI/vspI  
maeIII  
hphI  
maeIII

Restriction Enzymes:

haeIII/palI  
haeI  
scrFI  
mvaI  
ecoRII  
dsav  
bstNI  
nlaIII  
mnlI  
accI  
nlaIII  
apoI  
maeIII  
aflIII  
maeII  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
fnu4HI  
acII  
thaI  
fnuDII/mvnI  
tru9I  
bstUI  
msei  
bsaJI  
bsh1236I  
aseI/asnI/vspI  
maeIII  
hphI  
maeIII

Restriction Enzymes:

haeIII/palI  
haeI  
scrFI  
mvaI  
ecoRII  
dsav  
bstNI  
nlaIII  
mnlI  
accI  
nlaIII  
apoI  
maeIII  
aflIII  
maeII  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
fnu4HI  
acII  
thaI  
fnuDII/mvnI  
tru9I  
bstUI  
msei  
bsaJI  
bsh1236I  
aseI/asnI/vspI  
maeIII  
hphI  
maeIII

Restriction Enzymes:

haeIII/palI  
haeI  
scrFI  
mvaI  
ecoRII  
dsav  
bstNI  
nlaIII  
mnlI  
accI  
nlaIII  
apoI  
maeIII  
aflIII  
maeII  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
fnu4HI  
acII  
thaI  
fnuDII/mvnI  
tru9I  
bstUI  
msei  
bsaJI  
bsh1236I  
aseI/asnI/vspI  
maeIII  
hphI  
maeIII

Restriction Enzymes:

haeIII/palI  
haeI  
scrFI  
mvaI  
ecoRII  
dsav  
bstNI  
nlaIII  
mnlI  
accI  
nlaIII  
apoI  
maeIII  
aflIII  
maeII  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
fnu4HI  
acII  
thaI  
fnuDII/mvnI  
tru9I  
bstUI  
msei  
bsaJI  
bsh1236I  
aseI/asnI/vspI  
maeIII  
hphI  
maeIII

Restriction Enzymes:

haeIII/palI  
haeI  
scrFI  
mvaI  
ecoRII  
dsav  
bstNI  
nlaIII  
mnlI  
accI  
nlaIII  
apoI  
maeIII  
aflIII  
maeII  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
fnu4HI  
acII  
thaI  
fnuDII/mvnI  
tru9I  
bstUI  
msei  
bsaJI  
bsh1236I  
aseI/asnI/vspI  
maeIII  
hphI  
maeIII

Restriction Enzymes:

haeIII/palI  
haeI  
scrFI  
mvaI  
ecoRII  
dsav  
bstNI  
nlaIII  
mnlI  
accI  
nlaIII  
apoI  
maeIII  
aflIII  
maeII  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
fnu4HI  
acII  
thaI  
fnuDII/mvnI  
tru9I  
bstUI  
msei  
bsaJI  
bsh1236I  
aseI/asnI/vspI  
maeIII  
hphI  
maeIII

Restriction Enzymes:

haeIII/palI  
haeI  
scrFI  
mvaI  
ecoRII  
dsav  
bstNI  
nlaIII  
mnlI  
accI  
nlaIII  
apoI  
maeIII  
aflIII  
maeII  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
fnu4HI  
acII  
thaI  
fnuDII/mvnI  
tru9I  
bstUI  
msei  
bsaJI  
bsh1236I  
aseI/asnI/vspI  
maeIII  
hphI  
maeIII

Restriction Enzymes:

haeIII/palI  
haeI  
scrFI  
mvaI  
ecoRII  
dsav  
bstNI  
nlaIII  
mnlI  
accI  
nlaIII  
apoI  
maeIII  
aflIII  
maeII  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
fnu4HI  
acII  
thaI  
fnuDII/mvnI  
tru9I  
bstUI  
msei  
bsaJI  
bsh1236I  
aseI/asnI/vspI  
maeIII  
hphI  
maeIII

Restriction Enzymes:

haeIII/palI  
haeI  
scrFI  
mvaI  
ecoRII  
dsav  
bstNI  
nlaIII  
mnlI  
accI  
nlaIII  
apoI  
maeIII  
aflIII  
maeII  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
fnu4HI  
acII  
thaI  
fnuDII/mvnI  
tru9I  
bstUI  
msei  
bsaJI  
bsh1236I  
aseI/asnI/vspI  
maeIII  
hphI  
maeIII

Restriction Enzymes:

haeIII/palI  
haeI  
scrFI  
mvaI  
ecoRII  
dsav  
bstNI  
nlaIII  
mnlI  
accI  
nlaIII  
apoI  
maeIII  
aflIII  
maeII  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
fnu4HI  
acII  
thaI  
fnuDII/mvnI  
tru9I  
bstUI  
msei  
bsaJI  
bsh1236I  
aseI/asnI/vspI  
maeIII  
hphI  
maeIII

Restriction Enzymes:

haeIII/palI  
haeI  
scrFI  
mvaI  
ecoRII  
dsav  
bstNI  
nlaIII  
mnlI  
accI  
nlaIII  
apoI  
maeIII  
aflIII  
maeII  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
fnu4HI  
acII  
thaI  
fnuDII/mvnI  
tru9I  
bstUI  
msei  
bsaJI  
bsh1236I  
aseI/asnI/vspI  
maeIII  
hphI  
maeIII

Restriction Enzymes:

haeIII/palI  
haeI  
scrFI  
mvaI  
ecoRII  
dsav  
bstNI  
nlaIII  
mnlI  
accI  
nlaIII  
apoI  
maeIII  
aflIII  
maeII  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
fnu4HI  
acII  
thaI  
fnuDII/mvnI  
tru9I  
bstUI  
msei  
bsaJI  
bsh1236I  
aseI/asnI/vspI  
maeIII  
hphI  
maeIII

Restriction Enzymes:

haeIII/palI  
haeI  
scrFI  
mvaI  
ecoRII  
dsav  
bstNI  
nlaIII  
mnlI  
accI  
nlaIII  
apoI  
maeIII  
aflIII  
maeII  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
fnu4HI  
acII  
thaI  
fnuDII/mvnI  
tru9I  
bstUI  
msei  
bsaJI  
bsh1236I  
aseI/asnI/vspI  
maeIII  
hphI  
maeIII

Restriction Enzymes:

haeIII/palI  
haeI  
scrFI  
mvaI  
ecoRII  
dsav  
bstNI  
nlaIII  
mnlI  
accI  
nlaIII  
apoI  
maeIII  
aflIII  
maeII  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
nlaIII  
styI  
ncoI  
dsal  
bsaJI  
fnu4HI  
acII  
thaI  
fnuDII/mvnI  
tru9I  
bstUI  
msei  
bsaJI  
bsh1236I  
aseI/asnI/vspI  
maeIII  
hphI  
maeIII

Restriction Enzymes:

haeIII/palI  
haeI  
scrFI  
mvaI  
ecoRII  
dsav  
bstNI  
nlaIII  
mnlI  
accI

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**FIG. 6F**

FIG. 6F

**FIG. 6G**

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## FIG. 6H

```

          eam1105I
          sau96I
          scrFI
          mvaI    avaII
          ecorII
          dsav
          bstNI  asuI
          bsaJI  mnlI
          mboII mboII
          bpuAI earI/ksp632I
          bbsI mnlI
          styI
          bsaJI
          2401 CTTGTGACAC ACCTCCCCCA TGCCACCGGT GCCAGCACCC TGAACCTCCTG GGAGGACCGT CAGTCTTCCT CTTCCTCCCA AAACCCAAGG ATACCCCTTAT
              GAACACTGTG TGGAGGGGT ACGGTGCCA CCGGTGCTGG ACTTGAGGAC CCTCCTGGCA GTCAGAAGGA GAAGGGGGGT TTTGGGTTC TATGGGAATA

          sau96I
          nlaIV
          avaII
          asuI
          mspI
          hpaII
          scrFI
          nciI
          dsav
          cauII
          2501 GATTTCCCGG ACCCTGAGG TCACGTGCGT GGTGGTGGAC GTGAGCCACG AAGACCCGA GGTCAGTTC AAGTGTGACG TGGACGGCGT GGAGGTGCAT
              CTAAGGGCC TGGGGACTCC AGTGCACGCA CCACCACCTG CACTCGGTGC TTCTGGGGCT CCAGTCAAG TTCACCATGC ACCTGCCGCA CCTCCACGTA

          acil
          thal
          fnuDII/mvnI
          bstUI
          bsh1236I
          sacII/sstII
          nspBII
          kspI
          dsal
          bsaJI
          aciI
          fnu4HI mnlI
          2601 AATGCCAAGA CAAAGCCCGG GGAGGAGCAG TTCAACAGCA CGTTCCGTGT GGTACCGTTC CTCACCGTCC TGACCCAGGA CTGGCTGAAC GGCAAGGAGT
              TTACGGTCT GTTCGGCGC CCTCCTCGTC AAGTTGTGCT GCAAGGCACA CCAGTCCGAG GAGTGGCAGG ACGTGTCTCT GACCGACTTG CCGTTCCTCA

          scrFI
          mvaI bsaI
          ecorII
          dsav
          bstNI
          mnlI
          hpaI bslI
          apyI(dcm+)
          rsaI
          csp6I
          rsaI
          csp6I

```

## FIG. 6I

2701 ACAAGTCAA GGTCTCCAAC AAAGCCCTCC CAGCCCCCAT CGAGAAACC ATCTCCAAA CCAAAGACA GCCCGAGAA CCACAGGTGT ACACCCCTGCC  
 TGTTCACGTT CCAGAGGTTG TTTCGGGAGG GTCGGGGGTA GCTCTTTGG TAGAGGTTT GGTTCCTGT CGGGCTCTT GGTGTCCACA TGTGGGACGG  
 rsaI  
 csp6I  
 bspI407I  
 bsaI  
 bsmAI  
 mnlI  
 taqI  
 scrFI  
 nciI  
 mspI  
 hpaII  
 dsav  
 cauII  
 xmaI/pspAI  
 smaI  
 scrFI  
 nciI  
 dsav  
 cauII  
 bsaJI  
 foki  
 bslI  
 avaI  
 mnlI  
 CCAAGATGA CCAAGAACCA GGTGAGCTG ACCTGCTGG TCAAGGCTT CTACCCGAGC GACATGCGG TGGAGTGGGA GAGCAGCGG  
 GGTAGGCGC CTCTCTACT GGTCTTGT CCAGTCGAC TGGAGGACC AGTTCCGAA GATGGGTCG CTGTAGCGC ACCTCACCT CTCGTGCGCC  
 acil  
 dsal  
 bslI  
 bsaJI  
 nspBII  
 fnu4HI  
 fnu4HI  
 bbvI  
 bbvI  
 2801 CCATCCCGG GAGGAGTGA CCAAGAACCA GGTGAGCTG ACCTGCTGG TCAAGGCTT CTACCCGAGC GACATGCGG TGGAGTGGGA GAGCAGCGG  
 GGTAGGCGC CTCTCTACT GGTCTTGT CCAGTCGAC TGGAGGACC AGTTCCGAA GATGGGTCG CTGTAGCGC ACCTCACCT CTCGTGCGCC  
 dsal  
 hphI  
 mnlI  
 mboII  
 nlaIV  
 mboII  
 scrFI  
 aluI  
 bsaJI  
 bspMI  
 bbvI  
 2901 CAGCCGAGGA ACAACTACAA CACCACGCT CCCATGCTG ACTCCGACGG CTCCTTCTTC CTCTACAGCA AGCTCACCGT GGACAAGAGC AGGTGGCAGC  
 GTCGGCTCT TGTGATGT GTGGTGCGA GGTACGACC TGGAGTGGC GAGATGCTG TCGAGTGGCA CCTGTTCTCG TCCACCGTCG  
 mspI  
 hpaII  
 pleI  
 mnlI  
 nlaIII  
 hinFI  
 nlaIV  
 mboII  
 scrFI  
 aluI  
 bsaJI  
 bspMI  
 bbvI  
 3001 AGGGGAACAT CTCTCATGC TCCGTGATGC ATGAGGCTCT GCACAACCGC TTCACGAGA AGAGCTCTC CTGTCTCG GGTAAATGAG TCGCAGCGCC  
 TCCCTTGTGA GAAGAGTAG AGGCACTACG TACTCCGAGA GGTGTTGGC AAGTGGTCT TCTCGGAGAG GGACAGAGC CCATTACTC ACGTGGCGG  
 xmnI  
 mboII  
 asp700  
 nlaIII  
 sfaNI  
 mnlI  
 ppu10I  
 nsiI/avaIII  
 scrFI  
 nciI  
 mspI  
 hpaII  
 dsav  
 bsmAI  
 mnlI  
 earI/ksp632I  
 bslI  
 cauII  
 mcrI  
 eagi/xnaIII/ecfXI  
 eaeI  
 cfrI

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## FIG. 6J

rmaI  
 mnlI  
 sau3AI  
 mboI/ndeII(dam-)  
 dpnI(dam+)  
 dpnII(dam-)  
 alwI(dam-)  
 nlaIV maeI hincII/hindII  
 bstYI/xhoII accI pstI  
 bamHI xbaI pIeI bsgI aluI bspMI  
 alwI(dam-) hinfI bspMI  
 3101 GGGGATCCTC TAGAGTCGAC CTCGAGAAGC TTGGCCGCCA TGGCCCAACT TGTATTATGC AGCTTATAAT GGTACAAAT AAAGCAATAG CATCACAAAT  
 CCCCTAGGAG ATCTCAGCTG GACGTCTTCG AACCGGCGGT ACCGGGTGA ACAATAACG TCGAATATTA CCAATGTTTA TTTCGTTATC GTAGTGTTA  
 rmaI  
 maeI  
 bsmI  
 3201 TTCACAAATA AAGCAATTTT TTCAGTCGAT TCTAGTTGTG GTTGTGCCAA ACTCATCAAT GTATCTTATC ATGTCTGAT CGATCGGAA TTAATTCGOC  
 AAGTGTAT TTCTGTAATAA AAGTCAGGTA AGATCAACAC CAACAGGTT TGAGTAGTTA CATAGATAG TACAGACCTA GCTAGCCCTT AATTAAGCCG  
 haeI  
 styI  
 ncoI  
 dsal haeIII/palI  
 fnu4HI nlaIII  
 bbvI bsaJI  
 3301 GCAGACCAT GGCCTGAAAT AACCTCTGAA AGAGGAACTT GGTAGGTAC CTTCTGAGGC GGAAGAACC AGCTGTGAA TGTGTGTGAG TTAGGGTGTG  
 CGTCGTGGTA CCGGACTTTA TTGGAGACTT TCTCCTTGAA CCAATCCATG GAAGACTCCG CCTTCTTGG TCGACACCTT ACACACAGTC AATCCACAC

styI  
 aciI  
 fnu4HI sau96I  
 sfii ncoI haeIII/palI  
 haeIII/palI  
 eaeI dsal asuI  
 aluI  
 fnu4HI  
 bbvI  
 maeIII  
 sfanI apol  
 3101 GGGGATCCTC TAGAGTCGAC CTCGAGAAGC TTGGCCGCCA TGGCCCAACT TGTATTATGC AGCTTATAAT GGTACAAAT AAAGCAATAG CATCACAAAT  
 CCCCTAGGAG ATCTCAGCTG GACGTCTTCG AACCGGCGGT ACCGGGTGA ACAATAACG TCGAATATTA CCAATGTTTA TTTCGTTATC GTAGTGTTA  
 sau3AI  
 mboI/ndeII(dam-)  
 dpnI(dam+)  
 dpnII(dam-)  
 pvuI/bspCI  
 mcrI  
 taqI(dam-) tru9I  
 clalI/bsp106(dam-)  
 sau3AI mseI  
 mboI/ndeII(dam-)  
 dpnI(dam+) xmnI  
 hinPI  
 dpnII(dam-) asel/asnI/vspI  
 alwI(dam-) asp700 hhaI/cfoI  
 3201 TTCACAAATA AAGCAATTTT TTCAGTCGAT TCTAGTTGTG GTTGTGCCAA ACTCATCAAT GTATCTTATC ATGTCTGAT CGATCGGAA TTAATTCGOC  
 AAGTGTAT TTCTGTAATAA AAGTCAGGTA AGATCAACAC CAACAGGTT TGAGTAGTTA CATAGATAG TACAGACCTA GCTAGCCCTT AATTAAGCCG  
 haeI  
 styI  
 ncoI  
 dsal haeIII/palI  
 fnu4HI nlaIII  
 bbvI bsaJI  
 3301 GCAGACCAT GGCCTGAAAT AACCTCTGAA AGAGGAACTT GGTAGGTAC CTTCTGAGGC GGAAGAACC AGCTGTGAA TGTGTGTGAG TTAGGGTGTG  
 CGTCGTGGTA CCGGACTTTA TTGGAGACTT TCTCCTTGAA CCAATCCATG GAAGACTCCG CCTTCTTGG TCGACACCTT ACACACAGTC AATCCACAC

[illegible]



**FIG. 6L**

[illegible]

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## FIG. 6M

4201 ATCGCCCTGA TAGACGGTTT TTGCCCCTTT TCGCCCTTT GACGTGGAG TCCACGTTCT TTAATAGTGG ACTCTTGTTC CAACTGGAA CAACACTCAA CCCTATCTCG  
TAGCGGGACT ATCTGCCAA AGCGGGAAA CTGCAACCTC AGGTGCAAGA AATTATCACC TGAGAACAAAG GTTTGACCTT GTTGAGTT GGGATAGAC

4301 GGCTATTCTT TTGATTATA AGGATTTTG CCGATTTCG CCTATTGGT AAAAATGAG CTGATTAAAC AAAAATTTAA CGCGAATTTT AACAAATAT  
CCGATAAGAA AACTAAATAT TCCTAATAAC GGCTAAAGCC GGATAACCAA TTTTACTC GACTAAATG TTTTAAATT GGCCTTAAA TTGTTTTATA

4401 TAACGTTTAC AATTATTAG TGCACTCTCA GTACAATCTG CTCTGATGCC GCATAGTTAA GCCAACTCCG CTATCGCTAC GTGACTGGT CATGCGTCCG  
ATTGCAATG TTAAATACC ACGTGAGAGT CATGTTAGAC GAGACTACCG CGTATCAATT CGTTGAGGC GATAGCGATG CACTGACCCA GTACCGACGC

4501 CCCGACACC CGCCAACACC CGCTGACCG CCCTGACGG CTGTCTGTCT CCCGGATCC GCTTACAGAC AAGCTGTGAC CGTCTCCGG AGTCTCATGT  
GGGCTGTGG GCGGTGTGG GCGACTGCG GCGACTGCCC GAACAGACGA GGGCCGTAGG CGAATGCTG TTCGACACTG GCAGAGGCC TCGACGTACA

4601 GTCAGAGTT TTCACCGTCA TCACCGAAAC CGCGGAGCA GTATTCTTGA AGACGAAAG CGCTCGTAT ACAGCTATTT TTATAGGTTA ATGTCATGAT  
CAGTCTCAA AAGTGGCAGT AGTGGCTTTG CGCGTCCGT CATAAGAACT TCTGCTTTCC CGGAGCACTA TCGGATAAA AATATCCAAT TACAGTACTA

## FIG. 6N

hinII/acyI  
 ahaII/bsaHI  
 aatII  
 ddeI maeII  
 4701 AATAATGGTT TCTTAGACGT CAGGTGGCAC TTTTCGGGGA AATGTGGCGG GAACCCCTAT TTGTTTATTT TTCTAATATC ATTCAATAT GTATCCGCTC  
 TTATTACCAA AGAATCTGCA GTCCACCGTG AAAAGCCCT TTACACGCGC CTGGGGATA AACAAATAAA AAGATTATG TAAGTTTATA CATAGCGGAG  
 rcaI  
 bspHI  
 bsrBI  
 aciI nlaIII  
 fnuDII/mvnI  
 bstUI  
 bsh1236I  
 hinPI  
 hhaI/cfoI  
 mboII  
 earI/ksp632I  
 4801 ATGAGACAAT AACCTGATA AATGCTTCAA AATGATGAA AAAGGAAGAG TATGAGTATT CAACATTTC GTGTGGCCTT TATTCCTTT TTTGGGCGAT  
 TACTCTGTTA TTGGGACTAT TTACGAAGTT ATTATAACTT TTTCCCTTCT ATACTCATTA GTTGTAAGG CACAGCGGA ATAAGGAAA AACGCCGTA  
 bsmAI  
 sspI  
 hgiAI/aspHI  
 bsp1286  
 sau3AI  
 mboI/ndeII(dam-)  
 dpmI(dam+) bmyI  
 dpmII(dam-)  
 eco57I  
 apaLI/snoI  
 sfaNI mboII(dam-) alw44I/snoI maeIII taqI alwI(dam-)  
 4901 TTTGGCTTCC TGTTTTGCT CACCCAGAAA CGCTGGTGAA AGTAAAGAT GCTGAAGATC AGTTGGGTGC ACAGTGGGT TACATCGAAC TGGATCTCAA  
 AAACGAAGG ACAAAACGA GTGGGTCTTT GCGACCACCT TCAATTTCTA CGACTTCTAG TCAACCCACG TGCTCACCCA ATGTAGCTTG ACCTAGAGTT  
 hphI  
 hphI  
 maeII  
 psp1406I  
 xmnI  
 asp700  
 mboII  
 CGAAGACGT TTTTCCAAATGA TGAGCACTTT TAAAGTTCTG CTATGTGGCG CGGTATTATC CCGTGTATGAC  
 GTCGCCATTC TAGGAACCTCT CAAAAGCGGG GCTTCTTGCA AAAGTTACT ACTCGTGAAA ATTTCAAGAC GATACACCGC GCCATAATAG GGCACACTAG  
 sau3AI  
 mboI/ndeII(dam-)  
 dpmI(dam+)  
 dpmII(dam-)  
 aciI alwI(dam-)  
 nspBII bstYI/xhoII  
 5001 CAGCGGTAAG ATCCTTGAGA GTTTCGCCG GTTTCGCCG CGAAGACGT TTTTCCAAATGA TGAGCACTTT TAAAGTTCTG CTATGTGGCG CGGTATTATC CCGTGTATGAC  
 GTCGCCATTC TAGGAACCTCT CAAAAGCGGG GCTTCTTGCA AAAGTTACT ACTCGTGAAA ATTTCAAGAC GATACACCGC GCCATAATAG GGCACACTAG  
 scrFI  
 nciI  
 mspI  
 hpaII  
 dsav  
 caulI bclI mcrI fnu4HI  
 5101 GCCGGGCAAG AGCAACTCGG TCGCCGCGATA CACTATTCTC AGAATGACTT GGTGAGTAC TCACCACTCA CAGAAAAGCA TCTTACGGAT GGCATGACAG  
 CGGCCGCTC TCGTTGAGCC AGCGGCGTAT GTGATAAGAG TCTTACTGAA CCAACTCATG AGTGGTCAGT GTCCTTTCTG AGAATGCCTA CCGTACTGTC  
 foki  
 nlaIII  
 rsaI  
 csp6I bsrI  
 scaI hphI maeIII  
 sfaNI  
 foki  
 nlaIII  
 fnuDII/mvnI  
 bstUI  
 bsh1236I  
 hinPI  
 hhaI/cfoI  
 ahaII/bsaHI  
 bsrBI  
 aciI nlaIII  
 fnu4HI  
 aciI  
 sau3AI  
 mboI/ndeII(dam-)  
 dpmI(dam+)  
 dpmII(dam-)  
 aciI alwI(dam-)  
 nspBII bstYI/xhoII  
 5001 CAGCGGTAAG ATCCTTGAGA GTTTCGCCG GTTTCGCCG CGAAGACGT TTTTCCAAATGA TGAGCACTTT TAAAGTTCTG CTATGTGGCG CGGTATTATC CCGTGTATGAC  
 GTCGCCATTC TAGGAACCTCT CAAAAGCGGG GCTTCTTGCA AAAGTTACT ACTCGTGAAA ATTTCAAGAC GATACACCGC GCCATAATAG GGCACACTAG  
 scrFI  
 nciI  
 mspI  
 hpaII  
 dsav  
 caulI bclI mcrI fnu4HI  
 5101 GCCGGGCAAG AGCAACTCGG TCGCCGCGATA CACTATTCTC AGAATGACTT GGTGAGTAC TCACCACTCA CAGAAAAGCA TCTTACGGAT GGCATGACAG  
 CGGCCGCTC TCGTTGAGCC AGCGGCGTAT GTGATAAGAG TCTTACTGAA CCAACTCATG AGTGGTCAGT GTCCTTTCTG AGAATGCCTA CCGTACTGTC  
 foki  
 nlaIII

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## FIG. 6P

```

maeIII
5701 AAGCATGGT AACTGTCAGA CCAAGTTTAC TCATATATAC TTTAGATTGA TTTAAACTT CATTTTAAAT TTAAGGAT CTAGGTGAAG ATCTTTTTCG
TTQGTAAACA TTGACAGTCT GGTTCAAATG AGTATATATG AAATCTAACT AAATTTTGA GTAAATAATTA AATTTCTTA GATCCACTTC TAGGAAAAAC

      rnaI      sau3AI
sau3AI hphI mboI/ndeII(dam-)
mboI/ndeII(dam-)
dpnI(dam+) dpnI(dam+)
dpnII(dam-) dpnII(dam-)
      shaIII/draI maeI
tru9I tru9I bstYI/xhoII alwI/dam-
mseI mseI mseI alwI(dam-) mboII(dam-)
      ahaIII/draI
5801 ATAATCTCAT GACCAAAATC CCTTAACGTG AGTTTTCGTT CCACTGAGCG TCAGACCCCG TAGAAAGAT CAAGGATCT TCTTGAGATC CTTTTCCTTCT
TATTAGAGTA CTGGTTTTCG GGAATTGCAC TCAAAAGCAA GGTGACTGCG AGTCTGGGCG ATCTTTTCTA GTTTCCTAGA AGAACTCTAG GAAAAAAGA

      nlaIII      maeII      hgaI
xcaI      tru9I      ddeI
bspHI      mseI
      sau3AI
mboI/ndeII(dam-)
dpnI(dam+) sau3AI
dpnII(dam-) mboI/ndeII(dam-)
bstYI/xhoII dpnI(dam+)
sau3AI alwI(dam-) dpnII(dam-)
mboI/ndeII(dam-) alwI(dam-)
dpnI(dam+) mboII(dam-)
dpnII(dam-) bstYI/xhoII

      sau3AI
mboI/ndeII(dam-)
dpnI(dam+)
dpnII(dam-)
alwI(dam-)
      mspI      aciI      nspBII      hpaII      aluI      bsrI      maeIII      eco57I
5901 GCGCGTAATC TGCTGCTTGC AAACAAAAA ACCACCGCTA CCAGCGGTGG TTTGTTTGGC GGATCAAGAG CTACCAACTC TTTTCCGAA GGTAAGTGGC
CGCGCAATTAG ACGACGAACG TTTGTTTTTT TGGTGGCGAT GGTGCGCACC AAACAAACGG CTAAGTTCTC GATGTTGAG AAAAAGGCTT CCATTGACCG

      hinPI      rnaI      haeIII/palI
hinPI      maeI      bslI      haeI      scfI      aciI      mnlI
      hhaI/cfoI
6001 TTCAGCAGAG CGCAGATACC AAATACTGTC CTTCTAGTGT AGCCGTAGTT AGGCCACCAC TTCAAGAACT CTGTAGCACC GCCTACATAC CTCGCTCTGC
AAGTCGCTC GCGTCTATGG TTTATGACAG GAAGATCACA TCGGCATCAA TCCGGTGGTG AAGTCTTGA GACATCGTGG CGGATGTATG GAGCGAGACG

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## FIG. 6Q

```

        scrFI      acil      nspBII      fnu4HI      mspI      hpaII      bsaWI      maeIII      hhaI/cfoI
        nciI      mspI      hpaII      dsav      cauII      hinfi      pleI      hinfi      maeIII      hhaI/cfoI
        fnu4HI      bsvI      bsrI      fnu4HI      bsvI      bsrI      fnu4HI      bsvI      bsrI      fnu4HI      bsvI      bsrI
        alwNI      bsrI      fnu4HI      bsvI      bsrI      fnu4HI      bsvI      bsrI      fnu4HI      bsvI      bsrI
        maeIII      bsrI      fnu4HI      bsvI      bsrI      fnu4HI      bsvI      bsrI      fnu4HI      bsvI      bsrI
        6101 TAATCCTGTT ACCAGTGGCT GCTGCCAGTG GCGATAAGTC GTGTCTTACC GGCTTGACT CAAGACGATA GTTACCGGAT AAGCGGCAGC GGTCCGGCTG
        ATTAGGACAA TGGTCACCGA CGACTATTCAG CGCTATTCAG CACAGAATGG CCAACCTGA GTTCTGCTAT CAATGGCCTA TTCCGGCTG CCAGCCGAC
        hgiAI/aspHI      bspI286      bsiHKAI      bmyI      apaLI/snoI      alw44I/snoI      aluI      ddeI      scfI      hhaI/cfoI
        6201 AACGGGGGGT TCGTGACAC AGCCAGCTT GGAGCGAAG ACCTACACCG AACTGAGATA CCTACAGGT GAGCATTTAG AAAGGCCAC GCTTCCGAA
        TTGCCCCCA AGCAGGTG TCGGTGAA CCGCTGCTG TGGATGCG CTTGCTGCTT TGGATGCG TGGATGCG TGGATGCG TGGATGCG TGGATGCG
        scrFI      mvaI      ecorII      dsav      bstNI      bsaJI      aluI      apyI(dcm+)      apyI(dcm+)
        mspI      hpaII      bsaWI      fnu4HI      bsaWI      fnu4HI      bsaWI      fnu4HI      bsaWI      fnu4HI      bsaWI
        6301 GGGAGAAAG CGGACAGTA TCCGTAAGC GCGAGGTG GAACAGGAGA GCGACGAGG GAGCTTCAG GGGGAAAGC CTGGTATCTT TATAGTCTG
        CCTCTTTCC GCTGTCCAT AGGCCATTCG CGTCCCAGC CTTGTCTCT CCGTGTCTC CCGTGTCTC CCGTGTCTC CCGTGTCTC CCGTGTCTC
        haeIII/palI      fnu4HI      acil      thalI      bslI      fnuDII/mvni      bstUI      bsh1236I      nlaIV
        mnlI      drdI      hgaI      taqI      nlaIV      acil      bsh1236I      nlaIV
        6401 TCGGGTTTCG CCACCTCTGA CTTGAGCGTC GATTTTGTG ATGCTGTC AAGCGGCGGA GCCTATGAA AAAGGCCAGC AAGCGGCTT TTTACGGTT
        AGCCCAAAGC GGTGGAGACT GAACGTCAG CTAAGAACAC TACGAGCAGT CCCCCCGCT CGATACCTT TTTGGGCTG TTGCGCCGA AAATGCCAA

```

**SUBSTITUTE SHEET (RULE 26)**

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>length: 6889
```

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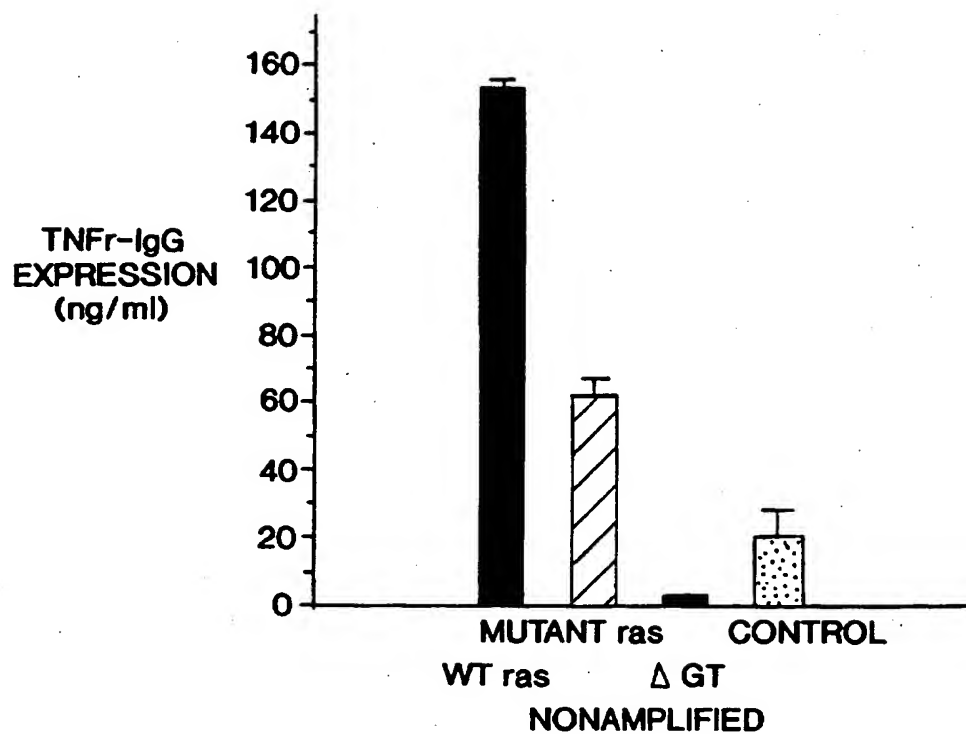


FIG. 7A

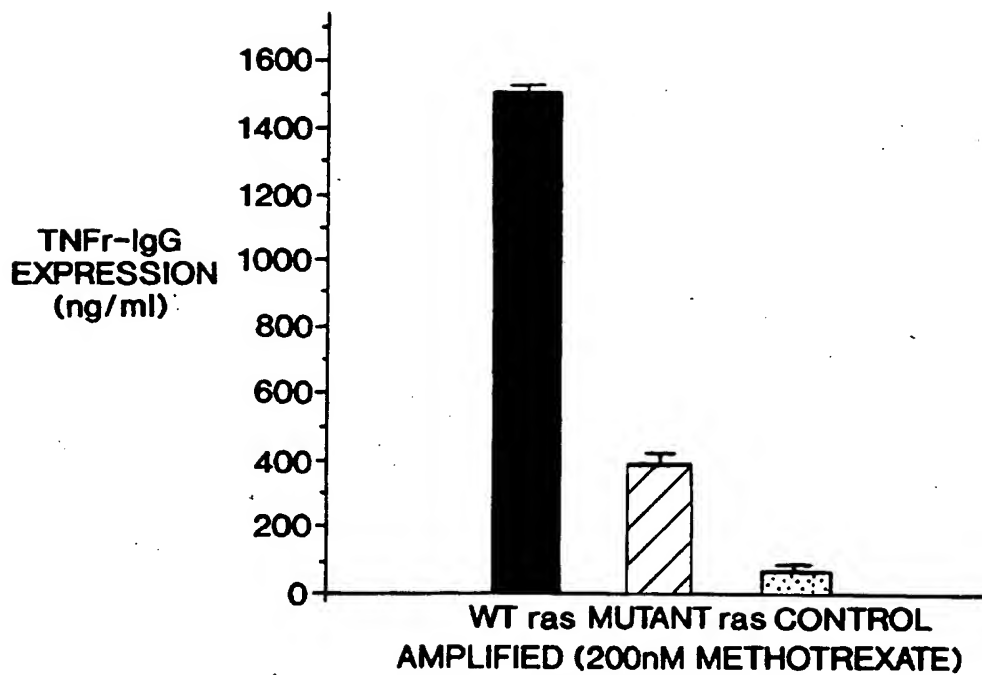


FIG. 7B



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FIG. 8

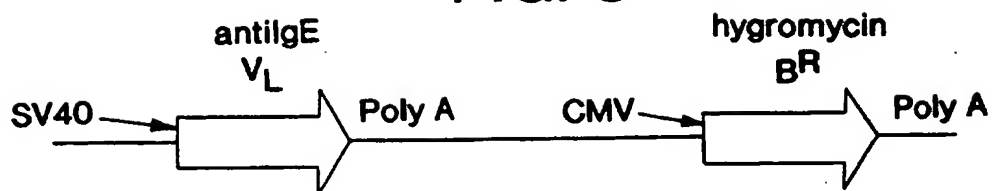
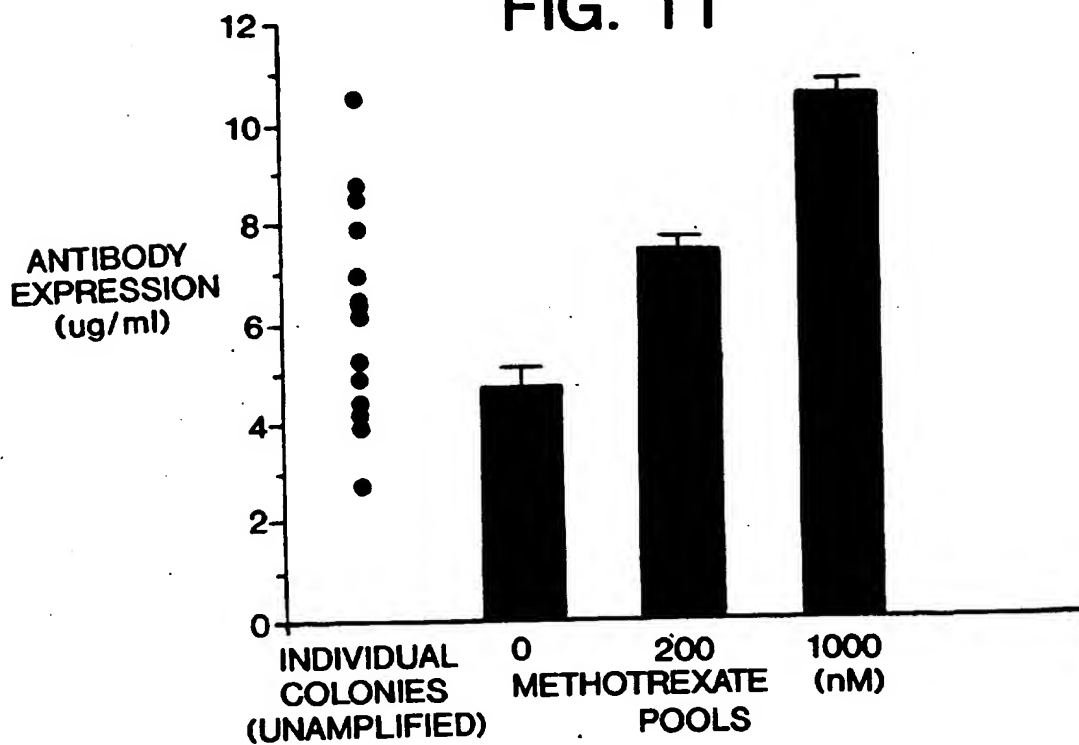


FIG. 11



[illegible]

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## FIG. 9B

```

scrFI      tfII      hinfI      fnu4HI      bbvI      nspBII      nlaIII.
ncII       aciI      thal      fnuDII/mvni  bstUI      bsh1236I
mspI       hpaII     dsav      cauII      CGCGGAAACGG  TGCATTGGAA  CGCGGATTCC  CCGTGCCAAG  AGTGACGTAA  GTACCGCTA  TAGACGATA  AGAGGATTTT  ATCCCGCTG  CCATCATGGT
401 dsav      cauII      GCGCCTAGG  GGCACGGTTC  TCACTGCATT  CATGCCGAT  ATCTGCTAT  TCTCTAAAA  TAGGGCGGAC  GGATGACCA
GGCCCTTGGC  ACGTAACCTT  GCGCCTAAGG  GGCACGGTTC  TCACTGCATT  CATGCCGAT  ATCTGCTAT  TCTCTAAAA  TAGGGCGGAC  GGATGACCA

          haeIII/palI
          haeI
scrFI      mvaI      bsrBI
mvaI      ecorII
dsav      bstNI      aciI      xmnI      rsaI      csp6I
          apyI[dcM+]  bsajI  mmlI  ddeI  asp700  scaI
          bsmAI      bsaI      bsaJI  mmlI  ddeI  asp700  scaI
          pflMI      bsmAI      bsaI      bsaJI  mmlI  ddeI  asp700  scaI
          bsII      bsaI      bsaJI  mmlI  ddeI  asp700  scaI
          taqI      sfanI      pflMI      bsmAI      bsaI      bsaJI  mmlI  ddeI  asp700  scaI
501 TCGACCATTC  AACTGCATCG  TCGCCGTGTC  CCAAAATATG  GGGATTGGCA  AGAAGGAGA  CCTACCTGG  CCTCCCTCA  GGAACGAGTT  CAAGTACTTC
AGCTGGTAAC  TTGACGTAGC  AGCGGCACAG  GGTTTTATAC  CCTAACCGT  TCTTGCTCT  GGATGGACC  GGAGGCGAGT  CCTTGCTCAA  GTTCATGAAG

          scrFI      mvaI      ecorII      dsav      bstNI      apyI[dcM+]  sexAI      trfI      hinfI      msel      tru9I
          mboII      earI/ksp632I  mmlI      alwNI      hphI      ddeI  mboII  taqI  ahaII/draI
601 CAAAGAATGA  CCACAACCTC  TTCAGTGGAA  GGTAAACAGA  ATCTGGTGAT  TATGGGTAGG  AAAACCTGGT  TCTCCATTCC  TGAGAAGAT  CGACCTTTAA
GTTCTTACT  GGTGTTGGAG  AAGTCACCTT  CCATTGTCT  TAGACCACCTA  ATACCCTCC  TTTTGACCA  AGAGGTAAGG  ACTCTTCTTA  GCTGGAATT

          sstI      sacI      hgiII      hgiAI/aspHI  ecl136II  bsp1286  bsiHRAI  bmyI      banII      bali      mmlI  aluI      bstXI      foki  sfanI  msel
          tru9I      msel      aseI/asnI/vspI      ddeI      ahlII/bfrI
701 AGGACAGAAT  TAATATAGTT  CTCAGTAGAG  AACTCAAGA  ACCACACGA  GGAGCTCAT  TTCTGCCAA  AAGTTGGAT  GATGCCTTAA  GACTTATGA
TCTGTCTTA  ATTATATCAA  GAGTCATCTC  TTGAGTTCT  TGGTGGTCT  CCTCGAGTAA  AAGAAGGTT  TTCAAGCTA  CTACGGAAT  CTGAATAACT

```

## FIG. 9C

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801 ACAACCGGAA TTGGCAAGTA AAGTAGACAT GGTTCGGATA GTCCGAGGCA GTTCGTGTTA CCAGGAGCC ATGATCAAC CAGGCCACCT TAGACTCTTT  
 TGTGGCCTT AACCGTTCAT TTCATCTGTA CCAACCTAT CAGCCTCGT CAGACAAAT GGTCCTCGG TACTAGTTG GTCCGTGGA ATCTGAGAAA  
 mspI hpaII bsaBI accI nlaIII mnlI  
 scrFI mvaI ecorII dsav bstNI nlaIII bstNI ddeI pleI  
 haeIII/palI haeI  
 901 GTGACAAGGA TCAGCAGGA ATTGAAAGT GACACGTTT TCCAGAAAT TGATTGGGG AAATAAAGC CTCGCCAGA ATACCCAGGC GTCTCTCTG  
 CACTGTCTCT AGTAGCTCT TAACTTTCA CTGTGCAAAA AGGTCTTTA ACTAAACCC TTTATATTG GAGAGGTCT TATGGGTCCG CAGGAGAGAC  
 nlaIII sau3AI mboI/ndeII[dam-] maeII  
 dpmI[dam+] afIII  
 maeIII alwI[dam-] apoI maeIII  
 1001 AGGTCCAGGA GGAAAAGGC ATCAAGTATA AGTTGAAGT CTACGAGAAG AAGACTAAC AGGAGATGC TTCAAGTTC TCTGCTCCC TCTAAAGCT  
 TCCAGGTCT CCTTTTCCG TAGTTCATAT TCAAACTTCA GATGCTCTC TTCTGATTG TCCTTCTACG AAGTTCAAG AGACGAGGG AGGATTTGGA  
 scrFI mvaI ecorII dsav bstNI apyI[dcm+] sau96I  
 avall asuI mnlI sfaNI accI mboII mboII mnlI aluI  
 1101 ATGCAATTTT ATAAGACCAT GGGACTTTTG CTGCTTTTAG ATCCCTTGG CTTCGTTAGA AGCAGACTAC AATTAATACA TAACCTTATG TATCATACAC  
 TACGTAAAAA TATTCTGGTA CCTGTGGAAC GACCGAAATC TAGGGAAACC GAAGCAATCT TGCCTCGATG TTAATTATGT ATTGGAATAC ATAGTATGCG  
 ppulOI nsiI/avaIII bstNI xhoII  
 nlaIII styI ncoI dsal bsaJI  
 sau3AI mboI/ndeII[dam-] dpmI[dam+] dpmII[dam-] aluI tru9I  
 bstNI xhoII bsaJI  
 fnu4HI mseI  
 bsvI aseI/asnI/vspI

**FIG. 9D**

[illegible]

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**FIG. 9E**

FIG. 9E

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## FIG. 9F

```

scrFI      scrFI
mvaI       mvaI
ecorII     ecorII
dsav       dsav
bstNI      bstNI
apyI(dcm+) apyI(dcm+)
hinPI      hinPI
hhaI/cfoI  hhaI/cfoI
nlaIV      nlaIV
nari       nari
kasi       kasi
hinII/acyI hinII/acyI
hgiCI      hgiCI
haeII      haeII
bani       bani
ahaII/bsaHI ahaII/bsaHI
1601 ACTGCAGAT GAACAGCCTG CGTGCTGAGG AACTGCGGT CTATTATTGT GCTCGAGGCA GCCACTATTG CCGCGCTGG CACTTCCCG TGTGGGTCA
TGGACGTCTA CTTGTCCGAC GCAGACTCC TGTGACGCA GATAATAACA CGAGCTCCGT CGTGATATAA GCCGCGGACC GTGAGCGGC ACACCCCGT

scfI       scfI
pstI       pstI
bsgI       bsgI
bspMI      bspMI
mmII       mmII
ddeI drdI  ddeI drdI
1601 ACTGCAGAT GAACAGCCTG CGTGCTGAGG AACTGCGGT CTATTATTGT GCTCGAGGCA GCCACTATTG CCGCGCTGG CACTTCCCG TGTGGGTCA
TGGACGTCTA CTTGTCCGAC GCAGACTCC TGTGACGCA GATAATAACA CGAGCTCCGT CGTGATATAA GCCGCGGACC GTGAGCGGC ACACCCCGT

sau96I     sau96I
haeIII/palI haeIII/palI
sau96I     sau96I
nlaIV      nlaIV
hgiJII     hgiJII
bsp1286    bsp1286
bmyI       bmyI
banII      banII
asui       asui
scrFI      scrFI
mvaI       mvaI
ecorII     ecorII
dsav       dsav
bstNI hphi bstNI hphi
apyI(dcm+) bsmAI haeIII/palI eco109I/draII apaI mboII dsav hgiAI/aspHI bsp1286 fnu4HI
bsaJI maeIII mmII nlaIV bstEII esp3I bsaJI mmII bpuAI apyI(dcm+) mmII bsiHKAI bsp1286 acilI apyI(dcm+)
nlaIV bsteII esp3I bsaJI mmII bbsI bsaJI mmII bmyI mmII bmyI nspBII bsaJI bmyI nspBII bsaJI
1701 AGGAACCCCTG GTCACCTCT CCGCGGCTC CACCAAGGC CCATCGGTCT TCCCTGGC ACCCTCTCC AGAGCACT CTGGGGGCAC AGCGCCCTG
TCCTGGGAC CAGTGGCAGA GGAGCCGAG GTGTTTCCC GTGAGCCAGA AGGGGACCG TGGGAGGAG TTCTCTGGA GACCCCGTG TCGCCGGAC

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**FIG. 9G**

[illegible]



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## FIG. 9H

```

sau96I      sau96I      sau96I      sau96I      sau96I      sau96I      sau96I      sau96I      sau96I      sau96I
nlaIV       nlaIV       nlaIV       nlaIV       nlaIV       nlaIV       nlaIV       nlaIV       nlaIV       nlaIV
avaII       avaII       avaII       avaII       avaII       avaII       avaII       avaII       avaII       avaII
mspI        mspI        mspI        mspI        mspI        mspI        mspI        mspI        mspI        mspI
scrFI       scrFI       scrFI       scrFI       scrFI       scrFI       scrFI       scrFI       scrFI       scrFI
ncII        ncII        ncII        ncII        ncII        ncII        ncII        ncII        ncII        ncII
sau3AI      hpaII      hpaII      hpaII      hpaII      hpaII      hpaII      hpaII      hpaII      hpaII
mboI/ndeII(dam-)
dpmI(dam+)  nlaIII      dsav      dsav      dsav      dsav      dsav      dsav      dsav      dsav
nlaIII      dsav      dsav      dsav      dsav      dsav      dsav      dsav      dsav      dsav
rcal        cauli      cauli      cauli      cauli      cauli      cauli      cauli      cauli      cauli
bspHI(dam-) asuI      eco8II     nspHI      nspHI      nspHI      nspHI      nspHI      nspHI      nspHI
mnlI      dpmII(dam-) bsu36I/mstII/sauI maeII      maeII      maeII      maeII      maeII      maeII      maeII
styI        bsaJI        bsaJI        bsaJI        bsaJI        bsaJI        bsaJI        bsaJI        bsaJI        bsaJI
2101 TTCCCCCCAA AACCCACGGA CACCTCATG ATCTCCCGGA CCCCTGAGGT CACATGCCGT GTGGTGGACG TGAGCCACGA AGACCTTGAG GTCAAGTTCA
AAGGGGGGTT TTGGGTTTCT GTGGGAGTAC TAGAGGGCCT GGGGACTCCA GTGTACGCAC CACCACCTGC ACTGGGTGCT TCTGGGACTC CAGTTCAAGT
2201 ACTGGTACGT GGACGGCCGT GAGGTGCATA ATGCCACAGC AAGCCCGCGG GAGGAGCAGT ACACAGCAC GTACCGTGTG GTACCGTGC TCACCGTCT
TGACCATGCA CTTGCCGCAC CTCCACGTAT TACGGTTCTG TTTCGGCGCC CTCTCGTCA TGTGTGCTG CATGGCACAC CAGTGGCAGG AGTGGCAGGA
2301 GCACCCAGGAC TGGCTGAATG GCAAGGAGTA CAAGTGCAGG GTCTCCAACA AAGCCCTCCC AGCCCCCATC GAGAAACCA TCTCCAAAGC CAAAGGCAG
CGTGGTCTCTG ACCGACTTAC CGTTCCTCAT GTTCACGTTT CAGAGTTGT TTCCGGAGGG TCGGGGGTAG CTCTTTTGGT AGAGGTTTCG GTTTCCTGTC

```

SUBSTITUTE SHEET (RULE 26)

taqI[dam-]  
clai/bsp106[dam-]  
sau3AI  
mboI/ndeII[dam-]  
dpnI[dam+]  
dpnII[dam-]  
lwi[dam-]

bsmi  
rmai

rsai  
cgn61

SCIFI .

aciI

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## FIG. 9K

```

nlaIII      fnu4HI
styI        sfiI mnlI
ncoI        haeIII/pali
dsal        bsaJI bglI
bslI        haeIII/pali bsaJI mnlI aluI
aciI        mnlI mnlI aciI haeIII/pali mnlI
bsaJI       CGATTCTCG CCATCTCG CCCATGGCT GACTAATTTT TTTTATTTAT GCAGAGCG AGCGCGCTC GGCTCTGTAG CTATTCCAGA AGTAGTGAGG
GGTCAAGGCG GGTAGAGCG GGGTACCGA CTGATTAAAA AAATAAATA CGTCTCCGC TCCGGCGGAG CCGAGACTC GATAAGTCT TCATCACTCC

3201

          fnu4HI      hinPI
          mcrI       hhaI/cfoI
          eagI/xmaIII/ecI XI  thal
          eaeI       fnuDII/mvnl
          bsrBI      tru9I bstOI      bspMI
          xhoI notI   tru9I   bsh1236I   scfI
          pae7I haeIII/pali   hinPI   tru9I   pstI
          avai fnu4HI   paci   hhaI/cfoI mseI   bsgI
          maeIII taqI cfrI mseI   bsshII ahaIII/draI   maeIII
          aluI mnlI aciI mseI asci swaI sse8387I aluI bsrI
          AGGCTTTT GGAGGCTAG GCTTTTGCA AAAGCTGTTA CCTCGAGCG CGGTTAAT AGGCGCGC ATTAAATCC TGCAGGTAAC AGCTGGCAC
          TCCGAAAAA CCTCCGATC CGAAACGTT TTTCGACAT GGAGCTCGC GCGAATTA TTCCGCGCG TAAATTTAG AGTCCATTG TCGAACCGTG

3301

          scrFI
          mvaI
          ecorII
          dsav
          bstNI
          apyI(dcm+)
          bsaJI maeIII
          bsrI
          maeII maeIII
          TGGCCGTGCT TTTACACGT GTGACTGG AAAACCTGG CGTACCACAA CTTAATCGC TTGCAGACA TCCCCCTTC GCCAGTGGC GTAATAGCA
          ACCGGCAGCA AATGTTGCA GCACTGACC TTTTGGACC GCAATGGCT GAATTAGCG AACGTGCTGT AGGGGGAG CGTCCGACG CATTATCGT

3401

          haeIII/pali
          eaeI
          cfrI
          sau96I
          haeIII/pali
          asuI
          mnlI aciI
          mcrI
          sau3AI
          mboI/ndeII(dam-)
          dpnI(dam+)
          dpnII(dam-)
          haeIII/pali
          asuI
          pvul/bspCI
          mnlI aciI
          mcrI
          hinPI
          hhaI/cfoI
          nlaIV
          narI
          kasi
          hinII/acyI
          hgiCI
          haeII
          bani
          sfaNI
          bglI
          ahaII/bsaHI
          sfani aciI
          AGAGGCCCGC ACCGATCGC CTTCGACCA GTTGGTAGC CTGAATGGCG AATGGCGCT GATCGGTAT TTTCTCCTTA CGCATCTGT GGTATTCTCA
          TCTCCGGGCG TGGCTAGCG GAAGGTTGT CAACGCATCG GACTTACCG GACTTACCG TTAGCCGGA CTACCCATA AAGAGGAT CGGTAGACAC GCCATAAAGT

3501

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## FIG. 9L

hinPI                      fnu4HI                      hinPI                      hhaI/cfoI                      hinPI  
 hhaI/cfoI                      aciI                      hhaI/cfoI                      hhaI/cfoI                      hhaI/cfoI  
 thai                      fnuDII/mvni                      fnuDII/mvni                      fnuDII/mvni                      fnuDII/mvni  
 bstUI                      bstUI                      bstUI                      bstUI                      bstUI  
 bsh1236I                      bsh1236I                      bsh1236I                      bsh1236I                      bsh1236I  
 rsaI                      scfI                      fnu4HI                      hhaI/cfoI                      hhaI/cfoI  
 csp6I                      bslI                      aciI                      msel                      hhaI/cfoI  
 3601 CACCGCATAC GTCAAAGCAA CCATAGTAG CGCCCTGTAG CGCGCATTA AGCGCGCGG GTGTGTGTGT TACGCGCAG GTACCGCTA CACTTGCCAG  
 GTGGCGTATG CAGTTTCGTT GGTATCATGC GCGGACATC GCCGCGTAAT TCGCGCGGCC CACACCACCA ATCGCGCTCG CACTGGCGAT GTGAACGGTC  
 aciI                      maeII                      hinPI                      hhaI/cfoI                      haeII  
 hinPI                      hhaI/cfoI                      haeII                      hhaI/cfoI                      haeII  
 rnaI                      bsrBI                      maeII                      cfr10I                      aluI                      nlaIV  
 maeI                      aciI                      maeII                      cfr10I                      aluI                      nlaIV  
 3701 CGCCCTAGCG CCGCTCCTT TCGCTTCTT CCGCTCCTT CTGCGCAGT TCGCGGCTT TCCCGTCAA GCTCTAAATC GGGGGCTCCC TTAGGGGTTC  
 GCGGATCGC GGGCGAGGAA AGCGAAGAA AGCGGAGGAA GAGCGGTGCA AGCGGCGGCA AGCGGCGGCA AGGGCGAGT CGAGATTAG CCCCCGAGG AAATCCCAAG  
 nlaIV                      hgiII                      hgiII                      bsp1286                      bmyI                      banII                      nlaIV  
 hgiII                      hgiII                      hgiII                      bsp1286                      bmyI                      banII                      nlaIV  
 3801 CGATTAGTG CTTAAGGCA CCGGACCC AAAAAGCTG ATTGGGTGA TGGTTCAGT AGTGGGCGAT CGCCCTGATA GACGGTTTTT CGCCCTTTGA  
 GCTAAATCAC GAAATGCCGT GGAGCTGGG TTTTGAAC TAAACCACT ACCAAGTGA TCACCCGTA CGCGGACTAT CTGCCAAAA GCGGAAACT  
 pleI                      tru9I                      pleI                      hinfI                      hinfI                      bsrI                      bslI                      bslI                      aval  
 hinfI                      maeII                      maeII                      hinfI                      hinfI                      bsrI                      bslI                      bslI                      aval  
 3901 CGTTGGATC CAGTTCTTT AATAGTGAC TCTGTGTTCA AACTGGAACA AACTCAACC CTATCTCGG CTATCTTTT GATTATAAG GGATTTTGCC  
 GCAACCTCAG GTGCAAGAA TTATCACCTG AGAACAAAGT TTGACCTGT TTGAGTTGG GATAGAGGCC GATAAGAAAA CTAAATATTC CCTAAACCG  
 pleI                      tru9I                      pleI                      hinfI                      hinfI                      bsrI                      bslI                      bslI                      aval  
 hinfI                      maeII                      maeII                      hinfI                      hinfI                      bsrI                      bslI                      bslI                      aval  
 4001 GATTTCGCC TATTGGTTAA AAAATGAGCT GATTTAACAA AAATTTAAG CGAATTTAA CAAATATTA ACCTTTACAA TTTTATGCTG CACTTCTCAGT  
 CTAAAGCGG ATAACCAATT TTTTACTCGA CTAATTTGTT TTTAATTC GCCTAAATTC GTTATATAT TCGCAATGTT AAATACCAC GTGAGAGTCA  
 hgiII/aspHI                      bsp1286                      bsiHKA1                      bmyI                      ddeI                      apaLI/snoI                      rsaI  
 hgiII/aspHI                      bsp1286                      bsiHKA1                      bmyI                      ddeI                      apaLI/snoI                      rsaI  
 4001 GATTTCGCC TATTGGTTAA AAAATGAGCT GATTTAACAA AAATTTAAG CGAATTTAA CAAATATTA ACCTTTACAA TTTTATGCTG CACTTCTCAGT  
 CTAAAGCGG ATAACCAATT TTTTACTCGA CTAATTTGTT TTTAATTC GCCTAAATTC GTTATATAT TCGCAATGTT AAATACCAC GTGAGAGTCA

**FIG. 9M**

[illegible]

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## FIG. 9P

```

sau3AI
mboI/ndeII(dam-)
dpmI(dam+) sau3AI          thal
dpmII(dam-) mboI/ndeII(dam-)
bstYI/xhoII dpmI(dam+) fnuDII/mvnI
sau3AI alwI(dam-) dpmII(dam-) bstUI
mboI/ndeII(dam-) alwI(dam-) bsh1236I
dpmI(dam+) mboII(dam-) hinPI fnu4HI
dpmII(dam-) bstYI/xhoII hhaI/cfoI bbvI
5501 TTTTCGTTCC ACTGAGGTC AGACCCCGTA GAAAGATCA AAGGATCTTC TTGAGATCCT TTTTTCCTGC GCGTATCTG CTGCTTGCAA ACAAATAAAC
AAAAACAAGG TGAATCGCAG TCTGGGGCAT TTTTCTAGT TTCTAGTAG AACTCTAGGA AAAAAAGACG CGCATTAGAC GACCAACGTT TGTTTTTTTG

sau3AI
mboI/ndeII(dam-)
dpmI(dam+)
dpmII(dam-)
alwI(dam-)
mspI
aciI nspBII hpaII aluI          hinPI          hhaI/cfoI
5601 CACCGCTACC AGCGTGTT TGTTCGCGG ATCAAGAGCT ACCAATCTTT TTTCCGAAGG TAACTGGCTT CAGCAGAGCG CAGATACCAA ATACTGTCTT
GTGGCGATGG TGGCCACCAA ACAAACGCC TAGTTCTCGA TGGTTGAGAA AAAGGCTCC ATTGACCGAA GTGCTCTGCG GTCTATGTT TATGACAGGA

rmaI          haeIII/palI          scfI          aciI          mmlI          maeIII          bbvI          bsrI
maeI          bslI          haeI          CCGTAGTTAG GCCACCACTT CAAGAATCTT GTAGCACCGC CTACATACCT CGCTCTGCTA ATCTGTGTAC CAGTGGCTGC TGCCAGTGGC
AGATCACATC GGCATCAATC CGGTGGTGAA GTTCTTGAGA CATCGTGGCG GATGTATGGA GCGAGACGAT TAGGACAATG GTACCCGACG ACGTCAACG

scrFI          nciI          mspI          hpaII          dsav          cauII          pleI          hinfi          maeIII          hhaI/cfoI          hinPI          mcrI          fnu4HI          aspHI
5701 GTAAAGTCTG GTCTTACCG GTTGACTCA AGACGATAGT TACCGGATAA GGCGACGCG TGCGGGTGA CGGGGGGTTT GTCCACACAG CCCAGCTTGG
CTATTACGA CAGATGGCC CAACCTGAGT TCTGTCTATCA ATGGCTATT CCGCGTGGCC AGCCCGACTT GCGCCCAAG CAGGTGTGTC GGTGCAACC

5801

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## FIG. 9Q

5901 AGCGAAGCAG CTACACCGAA CTGAGATACC TACAGCGTGA GCATTGAGAA AGCGCCAGCG TTCCCGAAGG GAGAAAGCG GACAGGTATC CGGTAAGCGG  
 TCGCTTGCTG GATGTGGCTT GACTCTATGG ATGTGCACT CGTAACCTCTT TCGCGGTGCG AAGGCTTCC CTCTTTCCGC CTGTCCATAG GCCATTCCGC  
 nspl hinpi hpaII fnu4HI  
 bslI acil  
 bsaWI  
 6001 CAGGTGCGA ACAGGAGAGC GCACGAGGA GCTTCAGG GGAACGCCT GGTATCTTTA TAGTCTGTC GGGTTTCGCC ACCTCTGACT TGAGCGTGA  
 GTCCAGCCT TGCTCTCTCG CGTCTCTCCT CGAAGTCCC CCTTGCGGA CCATAGAAAT ATCAGGACAG CCCAAGCGG TGGAGACTGA ACTCGCAGCT  
 hinpi mmlI aluI apyI[dcM+] apyI[dcM+] mmlI drdI hgaI  
 hhaI/cfoI  
 6101 TTTTGTGAT GCTCGTCAGG GGGCGGAGC CTATGMAAA ACGCCAGCAA CGCGGCTTT TTACGGTTCC TGGCCTTTG CTGCGCTTTT GCTCAGATGT  
 AAAACACTA CGAGCAGTCC CCGCGCTCG GATACCTTTT TCGGTCGTT GCGCGGAAA ATGCCAAG ACCCGAAA GACCGAAA CGAGGTACA  
 sfaNI nlaIV acil nlaIII  
 haeIII/palI  
 6201 TCTTTCCTGC GTTATCCCTT GATTCTGTGG ATAACCGTAT TACCGCTTTT GAGTGAGCTG ATACCGCTCG CGCAGCCGA ACAGCGAGC GCAGCGAGTC  
 AGAAGGAGC CAATAGGGA CTAAGACACC TATTGGCATA ATGCGGAAA CTCACTCGAC TATGGGAGC GCGCTCGCTT TGTGCTCG CGTCTCTCAG  
 tflI fnu4HI  
 hinfI bsvI pleI  
 hinpi hhaI/cfoI  
 mcrI

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## FIG. 9R

```

        thal
        fnuDII/mvnI
        bstUI
        bsh1236I
        hinPI
        hhaI/cfoI
        thal
        fnuDII/mvnI
        bstUI
        bsh1236I.haeIII/palI
        pvuII
        bslI
        mnlI
        acII
        cfrI
        hinPI
        mseI
        nspBII
        bsrI
        6301 AGTGAGCGAG GAAGCGGAG AGGCCCAAT AGCMAACCG CCTCTCCCCG CGCGTTGGCC GATTCATTAA TCCAGCTGGC ACGACAGGTT TCCCGACTGG
        TCACTCGCTC CTTCGCCCTC TCGCGGTTA TCGGTTTGGC GGAGAGGGG GCGCAACCG CTAAGTAATT AGGTGACCG TGCTGTCCAA AGGGCTGACC

        scrFI
        mvaI
        ecorII
        dsav
        nlaIV bstNI
        hgiCI apyI(dcm+)
        banI bsaJI
        mspI
        hpaII
        6401 AAAGCGGCGA GTGAGCGCAA CGCAATTAAT GTGAGTTACC TCACATCATTG GGCACCCGAG GCTTTACACT TTATGCTTCC GGCTGGTATG TTGTGTGCAA
        TTTGCCCCGT CACTCGCGTT GCGTTAATTA CACTCAATGG AGTGAGTAAT CGGTGGGCTC CGAATGTGA AATACGAAGG CCGAGCATAC AACACACCTT

        tru9I
        mseI
        aseI/asnI/vspI
        xmnI
        asp700
        6501 TTGTGAGCGG ATAACAATTT CACACAGGAA ACAGCTATGA CCATGATTAC GAATTAA
        AACACTCGCC TATTGTTAA GTGTGCTCTT TGTGATACT GGTACTAATG CTTAATT

        aciI
        bsrBI
        nlaIII
        aluI
        nlaIII
        6501 TTGTGAGCGG ATAACAATTT CACACAGGAA ACAGCTATGA CCATGATTAC GAATTAA
        AACACTCGCC TATTGTTAA GTGTGCTCTT TGTGATACT GGTACTAATG CTTAATT

        >length: 6557

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## FIG. 10A

aluI  
 sstI  
 sacI  
 hgiJII  
 hgiAI/aspHI  
 ecli36II  
 bspI286  
 bsiHKA  
 bmyI  
 banII  
 taqI  
 1 TCGAGCTCG CCGGACATG ATTATTGACT AGTTATTAAAT AGTAATCAAT TAGGGGGTCA TTAGTTCATA GCCCATATAT GGAGTTCCGC GTTACATAAC  
 AAGCTCGAGC GGGCTGTAAC TAATAACTGA TCATACTGA TCATTAGTGA ATGCCCCAGT AATCAAGTAT CGGGTATATA CCTCAAGCGC CAATGTATTG

rmaI tru9I  
 maeI mseI  
 speI aseI/asnI/vspI  
 bslI  
 aciI maeIII  
 thaI  
 fnuDII/mvnI  
 bstUI  
 bsh1236I

scrFI  
 mvaI  
 ecorII  
 dsav  
 aciI  
 bglI bstNI  
 sau96I  
 haeIII/palI aciI  
 asuI apyI(dcm+)

maeII  
 hinII/acyI  
 ahaII/bsaHI  
 aatII  
 101 TTACGGTAA TGGCCGCCT GCGTGACCG CCAACGACCC CCGCCCATG ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA  
 AATGCCATT ACCGGCGGA CCGACTGGG GGTTCGTGG GCGGGGTAAC TGCAGTTATT ACTGCATACA AGGTATCAT TCGGGTTATC CCTGAAAGGT

maeII  
 hinII/acyI  
 ahaII/bsaHI  
 aatII  
 201 TTGACGTCAA TGGGTGGAGT ATTTACGTA AACTGCCAC TTGGCAGTAC ATCAAGTGA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT  
 AACTGCAGT ACCACCTCA TAAATGCCAT TTGACGGGTG AACGGTCATG TAGTTCACAT AGTATACGGT TCATGCGGG GATNACTGCA GTTACTGCCA

scrFI  
 mvaI  
 ecorII  
 aciI  
 bglI dsav  
 sau96I bstNI  
 haeIII/palI  
 asuI apyI(dcm+) bsrI nlaIII  
 301 AAATGGCCG CCTGGCATT TCGCCAGTAC ATGACCTTAT GGCACCTTCC TACTGGCAG TACATCTACG TATTAGTCAAT CGCTATTACC ATGGTGATGC  
 TTTACCGGCG GACCGTAAT ACGGGTCATG TACTGGAATA CCTGAAAGG ATGAACCGTC ATGTAGATGC ATATCAGTA GCGATAATGG TACCACTACG

nlaIII  
 styI  
 ncoI  
 dsai hphI aciI  
 bsaJI sfaNI

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## FIG. 10C

tfii  
 acii  
 thai  
 fnuDII/mvni  
 bstUI  
 bsh1236I  
 701 TTGGAACGG GATTCCCGT GCCAAGAGTG ACGTAAGTAC CGCCTATAGA GTCTATAGGC CCACCCCTT GGGTTGGTTA GNAOCCGGCT ACAATTATATA  
 AACCTTGGC CTAAGGGCA CGGTTCTCAC TGCATTTCATG GGGATATCT CAGATATCCG GGTGGGGAA CCGAAGCAAT CTTCGCCGA TGTTAATTAT

fnu4HI  
 acii  
 thai  
 fnuDII/mvni tru9I  
 bstUI msel  
 bsh1236I asel/asni/vspI

sau96I  
 avaiI  
 asuI  
 scrFI  
 mval  
 ecorII  
 dsav  
 bstNI  
 apyI[dcn+]

maeIII  
 hphI scfI foki  
 801 CATAACCTTA TGTATCATAC ACATACGATT TAGGTGACAC TATAGAATAA CATCCACTTT GCCTTCTCT CCACAGGTGT CCACCTCCAG GTCCAACTGC  
 GTATTGGAAT ACATAGTATG TGTATGCTAA ATCCACTGTG ATATCTTATT GTAGTGAAA CCGAAGAGA GGTCTCCACA GGTGAGGTC CAGTTGACG

hinII/acyI  
 ahaiI/bsaHI  
 aatII  
 thai  
 fnuDII/mvni  
 bstUI  
 acII maeII  
 hphI bsh1236I taqI  
 901 ACCTCGGTTT TAAGCTTATC GATATGAAAA AGCCTGAAT CACCGGACG TGTGTCGAGA AGTTCTGAT CGAAAGTTC GACAGGCTCT CCGACCTGAT  
 TCGAGCCAAG ATTGGAATAG CTATACTTTT TCGGACTTGA GTGGCGCTGC AGACAGCTCT TCAAGACTA GCTTTTCAAG CTGTCCGAGA GGCTGGACTA

aluI taqI  
 hindIII  
 ddeI claiI/bsp106  
 bsaJI  
 1001 GCAGCTCTCG GAGGGCGAAG AATCTCGTGC TTTCAGCTTC GATGTAGGATA GGCTGGATA TGCTTCGGG GTAATAGCT GCGCCGATGG TTTCTACAAA  
 CGTCGAGAGC CTCCCGCTTC TTAGAGCAGC AAGTCTGAAG CTACATCTC CCGACCTAT ACAGGACGCC CATTTATCGA CCGGCTACC AAGATGTTT

aluI  
 fnu4HI  
 bbvI  
 mnlI mboII  
 tfii  
 hinfi  
 taqI  
 aluI  
 mnlI  
 acii  
 aluI hhaiI/cfoI  
 bbvI  
 fnu4HI  
 hinPI

## FIG. 10D

hinPI mspI  
 hhaI/cfoI hpaII  
 thal mroI  
 acII  
 haeIII/palI bspMII  
 mcrI fnuDII/mvni bspEI  
 eagi/xmaIII/ecI XI bsaWI  
 eaeI bstUI tfil  
 cfrI bsh1236I hinfI  
 sfanI fnu4HI bslI accIII  
 1101 GATCGTTATG TTTATCGGCA CTTTGCATCG GCCGGCTCC CGATTCCGGA AGTGCCTTGAC ATTGGGGAAT TCAGCGAGAG CCTGACCTAT TGCATCTCCC  
 CTAGCAATAC AAATAGCCGT GAAACGTAGC CGCGCGGAGG GCTAAGGCCT TCACGAACCTG TAACCCCTTA AGTCGTCTC GGACTGGATA ACGTAGAGGG  
 sau3AI  
 mboI/ndeII(dam-) sfanI acII  
 dpnII(dam+) apol  
 dpnII(dam-) ecorI  
 1201 GATCGTTATG TTTATCGGCA CTTTGCATCG GCCGGCTCC CGATTCCGGA AGTGCCTTGAC ATTGGGGAAT TCAGCGAGAG CCTGACCTAT TGCATCTCCC  
 CTAGCAATAC AAATAGCCGT GAAACGTAGC CGCGCGGAGG GCTAAGGCCT TCACGAACCTG TAACCCCTTA AGTCGTCTC GGACTGGATA ACGTAGAGGG  
 hgiAI/asphi  
 bsp1286  
 bsiHKAI  
 bmyI  
 apaLI/snoI maeII  
 alw44I/snoI maeIII  
 bslI draIII maeIII  
 1201 GCCGTGCACA GGGGTACAG TTGCAACACC TGCCTGAAC CGAATGCCC GCTGTTCTGC AGCGGTGCG GGAGGCCATG GATCGCATCG CTGCGGCGGA  
 CGGCACGTGT CCCACAGTGC AACGTTGTG ACGGACTTTG CTTTACCGG CGACAAGAGC TCGGCCAGCG CTCTCGGTAC CTACGCTAGC GACGCCGCT  
 sau96I  
 avall  
 asul  
 sau96I rsrII/cspi  
 haeIII/palI acII tfil  
 acII asul cpoI hinfI  
 bsrBI  
 ddel  
 1301 TCTTAGCCAG ACAGACGGGT TCGGCCCAAT CGGACCGCAA GGAATCGGTC AATACACTAC ATGGCGTAT TTCATATGCG CGATTGCTGA TCCCATGTTG  
 AGAATCGGTC TGCTGCCCCA AGCCGGGTAA GCCTGGCGTT CCTTAGCCAG TTATGTGATG TACCGCACTA AAGTATACGC GCTAAGCACT AGGGGTACAC  
 hinPI  
 hhaI/cfoI  
 thal  
 fnuDII/mvni  
 bstUI  
 bsh1236I  
 taqI  
 aluI  
 sfanI  
 sau96I  
 haeIII/palI  
 bsaJI  
 dralII  
 nlaIV  
 hgiCI  
 bani  
 mspI  
 bslI  
 hpaII  
 1401 TATCACTGGC AAACGTGAT GGACGACACC GTCAGTGCGT CCGTCGCGCA GGTCTCGAT GAGCTGATGC TTTGGGCGGA GGACTGCCCC GAAGTCCGCG  
 ATAGTGACC GCTGCTGTGG CAGTCACGCA GGCAGCGGT CCGAGAGCTA CTCGACTACG AAACCGGCT CTGACGGG GTTACGGCGG

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## FIG. 10F

nlaIV  
 mspI  
 hpaII scrFI  
 bslI nciI  
 mroI mspI  
 bspMI hpaII  
 bspEI[dam-]  
 bsaWI dsav  
 accIII[dam-]  
 sau3AI caulI  
 mboI/ndeII[dam-]  
 dpnI[dam+]  
 dpnII[dam-]  
 alwI[dam-]  
 1801 ACGCAATCGT CCGATCCGA GCCGGGACTG TCGGGCGTAC ACAAAATGCC CGCAGAAGCG CGGCCGTCTG GACCGATGGC TGTGTAGAAG TACTCGCCGA  
 TCGGTAGCA GGTAGGCT CGCCCTGAC AGCCCGCATG TGTTAGCG GGTCTTCCG CCGGCAGAC CTGGCTACCG ACACATCTTC ATGAGCGGCT

haeIII/palI  
 mcrI  
 eagI/xmaIII/ecI XI  
 eaeI  
 cfrI  
 fnu4HI  
 aciI  
 thaI  
 fnuDII/mvnl  
 bstUI  
 bsh1236I sau96I  
 hinPI avaiI  
 hhaI/cfoI asuI  
 rsaI  
 csp6I  
 scaI  
 1901 TAGTGGAAAC CGACGCCCA GCATCGTCC GAGGCAAG GAATAGAGTA GATGCCGACC GAAGGATCCC CGGGGAATTC AATCGATGCC CGCCATGGCC  
 ATCACCTTTG GCTGCGGGT CGTGAGCAGG CTCCGCTTC CTTATCTCAT CTACGGCTGG GTTCTAGCG GCCCCTTAAG TTAGCTACCG GCGGTACCG

scrFI  
 nciI  
 mspI  
 hpaII  
 dsav  
 xmaI/pspAI  
 smaI  
 scrFI  
 nciI  
 dsav  
 caulI  
 bsaJI  
 avaiI  
 bsaJI  
 sau3AI  
 mboI/ndeII[dam-]  
 dpnI[dam+]  
 dpnII[dam-]  
 alwI[dam-]  
 nlaIV caulI  
 bstVI/xhoII  
 bamHI bsaJI ecoRI  
 alwI[dam-] apoI  
 clai/bsp106 bsaJI  
 mcrI  
 bslI  
 sfaNI  
 mnuII  
 bsaJI  
 hgaI  
 hpaII/bsaHI  
 ahaII/bsaHI  
 1901 TAGTGGAAAC CGACGCCCA GCATCGTCC GAGGCAAG GAATAGAGTA GATGCCGACC GAAGGATCCC CGGGGAATTC AATCGATGCC CGCCATGGCC  
 ATCACCTTTG GCTGCGGGT CGTGAGCAGG CTCCGCTTC CTTATCTCAT CTACGGCTGG GTTCTAGCG GCCCCTTAAG TTAGCTACCG GCGGTACCG

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## FIG. 10I

```

sau3AI
mboI/ndeII(dam-)
dpmI(dam+)
dpmII(dam-)
alwI(dam-)
taqI(dam-)
clai/bsp106(dam-)
sau3AI
mboI/ndeII(dam-)
dpmI(dam+)
dpmII(dam-)
alwI(dam-)
fokI
2701 ACCTTTTGA TCATCCTAC TGACACTGAC ATCCACTTTT TCTTTTCTC CACAGGTGC CACTCCAGG TCCAAGTGA CCTCGGTTCG CGAAGCTAGC
TGGAACACCT AGTAGGATG ACTGTGACTG TAGGTGAAA AGAAAAAGAG GTGTCCACAG GTGAGGGTCC AGTTGACGT GGAGCCAAGC GCTTCGATCG

nlaIII
styI
pflMI
ncoI
sfanI ecorI
fnu4HI taqI apoI
bbvI clai/bsp106
bsaJI
2801 TTGGGCTGCA TCCATTGAAT TCCACCATTG GATGTCATG TATCATCCTT TTTCTAGTAG CAACTGCAAC TGGAGTACAT TCAGATATCC AGCTGACCCA
AACCAGACGT AGCTAACTTA AGGTGGTACC CTACCAGTAC ATAGTAGGAA AAAGATCATC GTTGACGTTG ACCTCATGTA AGTCTATAGG TCGACTGGGT

aluI
sstI
sacI
hgiJII
hgiAI/aspHI
ecII36II
bsp1286
bsiHKA1
bmyI
banII
nmlI
aciI
2901 GTCCCGAGC TCCTGTCCG CCTCTGTGGG CGATAGGGTC ACCATCACCT GCCGTGCCAG TCAGAGCGTC GATTACGATG GTGATAGCTA CATGAACCTGG
CAGGGGCTCG AGGACACAGC GGAGACACCC GCTATCCCCG TGGTAGTGGA CGGCACGGTC AGTCTCGCAG CTAATGCTAC CACTATCGAT GTACTTGACC

sau96I
avaII
asuI
scrFI
mvaI
ecoRII
dsav
bstNI
apyI(dcm+)
bsaJI
bslI
bsaJI
2701 ACCTTTTGA TCATCCTAC TGACACTGAC ATCCACTTTT TCTTTTCTC CACAGGTGC CACTCCAGG TCCAAGTGA CCTCGGTTCG CGAAGCTAGC
TGGAACACCT AGTAGGATG ACTGTGACTG TAGGTGAAA AGAAAAAGAG GTGTCCACAG GTGAGGGTCC AGTTGACGT GGAGCCAAGC GCTTCGATCG

thai maeI
fnuDII/mvni
bstUI nheI
mnlI bsh1236I aluI
bsaJI nrui aluI
mnlI bsh1236I aluI
rsal
gsuI/bpmI
pvuII tth111I/aspI
bsrI csp6I
ecoRV nspBII bsrI
2801 TTGGGCTGCA TCCATTGAAT TCCACCATTG GATGTCATG TATCATCCTT TTTCTAGTAG CAACTGCAAC TGGAGTACAT TCAGATATCC AGCTGACCCA
AACCAGACGT AGCTAACTTA AGGTGGTACC CTACCAGTAC ATAGTAGGAA AAAGATCATC GTTGACGTTG ACCTCATGTA AGTCTATAGG TCGACTGGGT

hphI
maeIII bspMI
bstEII hphI
bsrI
hphI aluI nlaIII
hphI aluI nlaIII
2901 GTCCCGAGC TCCTGTCCG CCTCTGTGGG CGATAGGGTC ACCATCACCT GCCGTGCCAG TCAGAGCGTC GATTACGATG GTGATAGCTA CATGAACCTGG
CAGGGGCTCG AGGACACAGC GGAGACACCC GCTATCCCCG TGGTAGTGGA CGGCACGGTC AGTCTCGCAG CTAATGCTAC CACTATCGAT GTACTTGACC

```

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## FIG. 10J

mspI  
 hpaII  
 bslI  
 bsaWI  
 gsuI/bpmI  
 scrFI  
 mvaI  
 haeIII/palI  
 fnu4HI  
 ecorII  
 acII  
 bstNI  
 fnuDII/mvnI  
 apyI[dcM+] pleI  
 bstUI  
 rsaI  
 pleI  
 gsuI/bpmI  
 bsh1236I  
 csp6I  
 hinFI  
 hinFI  
 sau3AI  
 mboI/ndeII[dam-]  
 dpnII[dam-]  
 alwI[dam-]  
 nlaIV  
 bstYI/xhoII  
 bamHI  
 alwI[dam-]  
 mboI/ndeII[dam-]  
 dpnII[dam+]  
 3001 TATCAACAGA AACCAGGAAA AGCTCCGAAA CTACTGATTT ATAGTGTCT TTGGTCTTT TCGAGGCTTT GATGACTAAA TCGCGCGGAG CATGGACCTC AGACCTCAGG GAAGAGCGAA GAGACCTAGG CCAAGACCTC  
 ATAGTGTCT TTGGTCTTT TCGAGGCTTT GATGACTAAA TCGCGCGGAG CATGGACCTC AGACCTCAGG GAAGAGCGAA GAGACCTAGG CCAAGACCTC  
 fnu4HI mboII  
 bbvI bpuAI  
 scfI mspI  
 pstI hpaII  
 bsgI  
 sau3AI  
 mboI/ndeII[dam-]  
 dpnII[dam+]  
 dpnII[dam-]  
 3101 CGGATTTTAC TCTGACCATC AGCAGTCTGC AGCCGGAAGA CTTCGCAACT TATTACTGTC AGCAAAGTCA CGAGGATCCG TACACATTTG GACAGGGTAC  
 GCCTAAAGTG AGACTGGTAG TCGTCAGACG TCGGCTTTCT GAAGCGTTGA ATAATGACAG TCGTTTCAGT GCTCCTAGGC ATGTGTAAAC CTGTCCCATG  
 fnu4HI mboII  
 bbvI bpuAI  
 scfI mspI  
 pstI hpaII  
 bsgI  
 sau3AI  
 mboI/ndeII[dam-]  
 dpnII[dam+]  
 dpnII[dam-]  
 3201 CAAGGTGGAG ATCAACGAA CTGTGGCTGC ACCATCTGTC TTCACTTCC CGGCATCTGA TGAGCAGTTG AAATCTGGAA CTGCTCTGT TGTGTGCTG  
 GTTCCACCTC TAGTTTGCTT GACACCGACG TGGTAGACAG AAGTAGAAGG GCGGTAGACT ACTCGTCAAC TTAGACCTT GACGAGACA ACACAGGAC  
 haeIII/palI  
 haeI  
 mnlI  
 rsaI  
 csp6I  
 xmnI  
 asp700  
 3301 CTGAATAACT TCTATCCAG AGAGGCCAAA GTACAGTGGA AGGTGGATAA CGCCTCCAA TCGGGTAACT CCCAGGAGAG TGTACAGAG CAGGACAGCA  
 GACTTATTGA AGATAGGCTC TCTCGGCTT CATGTCACT TCCACCTATT TCCACCTATT GCGGAGGTT AGCCCATTTA GGGTCTCTC ACAGTGTCTC GTCTGTCTC

## FIG. 10K

sstI  
 sacI  
 hgiJII  
 hgiAI/asphi  
 ecl136II  
 bsp1286  
 bsiHKA  
 bmyI  
 haeIII/pali  
 sau96I aluI  
 asuI banII  
 hphI eco0109I/draII  
 maeIII alwNI ddeI  
 accI  
 ddeI  
 celII/espI  
 bpul102I  
 hgaI  
 ddeI fnu4HI  
 scfI mnlI bbvI  
 3401 AGGACAGCAC CTACAGCCTC AGCAGCACCC TGACGCTGAG CAAGCAGAC TAGAGAAAC ACAAGTCTA CGCCTGGAA GTACCCCATC AGGGCCTGAG  
 TCCTGTCGTG GATGTCGGAG TCGTCGTGGG ACTGCGACTC GTTTCGTCTG ATGCTCTTTG TGTTCAGAT CGGAGCGCTT CAGTGGGTAG TCCCGGACTC  
 sau96I  
 nlaIII  
 acII haeIII/pali  
 fnu4HI asuI  
 bglI styI  
 sfiI ncoI  
 aluI  
 hindIII eaeI dsal  
 tru9I cfrI bsaJI  
 mseI taqI haeIII/pali  
 maeIII aluI  
 3501 CTCGCCCGTC ACAAGAGCT TCAACAGGGG AGAGTGTTAA GTTTCGATGG CGCCATGGC CCAACTTGT TATTGAGCT TATAATGGT ACAATTAAG  
 GAGCGGGCAG TGTTCCTCGA AGTTGTCCCC TCTCACAAT CGAAGCTACC GCGGTACCG GGTGAACAA ATAACCTGA ATATTACCA TGTATTATTC  
 sau3AI  
 mboI/ndeII(dam-)  
 dpnI(dam+)  
 dpnII(dam-)  
 pvuI/bspCI  
 mcrI  
 taqI(dam-)  
 claiI/bsp106(dam-)  
 sau3AI  
 mboI/ndeII(dam-)  
 dpnI(dam+)  
 dpnII(dam-)  
 nlaIII alwI(dam-)  
 rmaI  
 bsmI maeI  
 sfaNI apoI  
 3601 CAATAGCATC ACAATTTCA CAATAAAGC ATTTTTTCA CTGCATCTA GTTGTGGTT GTCCAACTC ATCAATGTAT CTTATCATGT CTGGATCAT  
 GTTATCGTAG TGTTTAAAGT GTTTATTTCG TAAAAAAGT GACGTAAGAT CAACACCAG CAGGTTTCAG TAGTTACATA GAATAGTACA GACCTAGCTA

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**FIG. 10M**

FIG. 10M

styI  
bsaJI  
blnI  
haeIII/palI  
stuI rmaI  
haeI maeI  
mmII avrII  
4101 TTTTGTGAG GCCTAGGCTT TTGCAAAAG CTGTAAACAG CTGTGCACTG GCCTGCTGT TACAACGTG TGACTGGGA AACCTGTGCG TTACCCAACT  
AAAAAACC TC CGGATCCGAA AACGTTTTTC GACAATTGTC GAACCGTGAC CGGCAGCAAA ATGTTGCAGC ACTGACCCCT TTGGGACCGC AATGGGTGGA

tru9I  
mseI  
hpaI aluI  
aluI hincII/hindII bsrI  
alul hincII/hindII bsrI  
4201 TAATCGGCTT GCAGCACATC CCCCCTTGC CAGCTGGCGT AATAGCGAAG AGGCCCGCAC CGATCGCCCT TCCCAACAGT TCGGTAGCCT GAATGGCGAA  
ATTAGCGAA CGTGTGTAG GGGGGAAGCG GTGACCGCA TTATCGCTTC TCCGGCGGTG GCTAGCGGA AGGTTGTCA ACGCATCGA CTTACCGCTT

hinPI  
hhaI/cfoI  
nlaIV  
nari  
kasi  
hinII/acyI  
hgiCI  
haeII  
bani sfaNI  
ahaII/bsaHI  
4301 TGGCGCTGA TGGCGTATTT TCTCCTTAG CATCTGTGCG GTATTTCACA CCGCATACGT CAAAGCAACC ATAGTACGCG CCTGTAGCG GCGCATTAAG  
ACCGCGGACT ACGCCATAAA AGAGGAATGC GTAGACACGC CATAAAGTGT GCGGTATGCA GTTTCGTGG TATCATGCGC GGGACATGCG CCGGTAAATC

acII  
fnu4HI  
aciI  
thai  
fnuDII/mvni  
bstUI  
bsh1236I  
maeIII bsvI maeIII  
4401 CGCGCGGCT GTGGTGGTTA CGCGACGCT GACCGCTACA CTGCGCAGCG CCTAGCGCC CGCTCCTTTC GCTTCTTTC CTTCTTCTCT CCGCAAGTTC  
CGCGCGCCA CACACCAAT GCGCGTGC GAACGCTGCA CTGCGCATGT GAACGCTGCG GCGATCGCG GCGAGGAAG CGAAGAAGG GAAGGAAGA GCGGTGCAAG

scrFI  
mvaI  
ecorII  
dsaV  
bstNI  
apyI[dcM+]  
bsaJI maeIII  
tru9I  
mseI  
maeIII  
bsrI  
maeII  
maeIII  
sau3AI  
mboI/ndeII[dam-]  
dpmI[dam+]  
sau96I  
haeIII/palI  
mmII aciI  
mmII aciI  
mboII asuI  
earI/ksp632I  
mcrl  
bglI  
hinPI  
hhaI/cfoI  
thai  
fnuDII/mvni  
bstUI  
bsh1236I  
rsal  
scfI fnu4HI  
tru9I  
mseI  
csp6I  
bslI  
aciI  
hinPI  
hhaI/cfoI  
maeII  
bstUI  
bsh1236I  
maeII  
hhaI/cfoI  
bsrBI  
maeII  
maeI  
aciI  
fnu4HI  
hinPI  
hhaI/cfoI  
thai  
fnuDII/mvni  
bstUI  
bsh1236I  
maeIII bsvI maeIII  
4401 CGCGCGGCT GTGGTGGTTA CGCGACGCT GACCGCTACA CTGCGCAGCG CCTAGCGCC CGCTCCTTTC GCTTCTTTC CTTCTTCTCT CCGCAAGTTC  
CGCGCGCCA CACACCAAT GCGCGTGC GAACGCTGCA CTGCGCATGT GAACGCTGCG GCGATCGCG GCGAGGAAG CGAAGAAGG GAAGGAAGA GCGGTGCAAG



**FIG. 10N**

[illegible]

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**FIG. 10P**

**SUBSTITUTE SHEET (RULE 26)**

[illegible]

**FIG. 10R**

[illegible]

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## FIG. 10S

6701	CGCCACGCTT	CCCGAAGGGA	GAAGGCGGA	CAGGTATCCG	GTAAGCGGA	GGGTGGAAC	AGGAGAGCGC	ACGAGGGAGC	TTCCAGGGGG	AAACGCCCTGG	scrFI mvaI ecoRII dsav bstNI bsaJI hinPI mnII hhaI/cfoI aluI scrFI mvaI ecoRII dsav bstNI bsaJI apyI(dcm+)
	CGGCTGCGAA	GGGCTTCCTT	CTTTCCGCT	GTCCATAGGC	CATTGCGCGT	CCAGCCTTG	TCCTCTCGCG	TGCTCCCTCG	AAGTCCCCC	TTTGGGAGCC	
6801	TATCTTTATA	GTCTGTGCGG	GTTTCGCCAC	CTCTGACTTG	AGCGTCGATT	TTTGTGATGC	TGCTCAGGGG	GGCGGAGCCT	ATGGAAAAAC	GCCAGCAACG	fnu4HI aciI thaI fnuDII/mvnI bstUI bsh1236I nlaIV aciI nlaIV aciI sfaNI taqI hgaI mnII drdI haeIII/palI scrFI mvaI ecoRII dsav bstNI bsaJI apyI(dcm+)
	ATAGAAATAT	CAGGACAGCC	CAAGCGGTG	GAGACTGAAC	TGCGAGCTAA	AAACACTACG	AGCAGTCCCQ	CCGCCTCGGA	TACCTTTTGG	CGGTGCTTGC	
6901	CGGCTTTTAT	ACGCTTCTG	GCCTTTGCT	GGCTTTTGG	TCACATGTC	TTTCTGCGGT	TATCCCCCTGA	TTCTGTGGAT	AACCGTATTA	CGGCTTTTGA	bslI haeIII/palI nlaIV haeI haeIII/palI nspI haeIII/palI nspHI aflIII tffI hinfI aciI bslI bstNI apyI(dcm+)
	CGCCGAAAAA	TGCCAAGGAC	CGGAAACGA	CCGAAACGA	AGTGATACAAG	AAAGGACGA	ATAGGGGACT	AGACACCTTA	TTGGCATAAT	GGCGGAAACT	

**FIG. 10T**

FIG. 10T

[illegible]

```
scrFI
mvaI
ecorII
dsav
bstNI
apyI(dcm+)
```

	apyl[dcn+]	mspi	aciI	xmnl
	bsauI	hpaII	bsrBI	alul
7201	CACCCCAAGC	TTTACACTTT	ATGCTTCGG	CTCGTATGTT
	GTGGGGTCCG	AAATGAGAAA	TACGAAGGCC	GAGCATACAA
			CACACCTTAA	CACACCTTAA
			CTGACGGGAT	AACAAATTCA
			ACCTATGACC	ATGATTACGA
			GTGTCCTTTG	TCGATACCTG
			TGTTTAAAGT	TACTAATGCT
			nlaiII	asp700

```

tru9I
mseI
aseI/asnI/vspI
7301 ATTAA
TAATT

```

>length: 7305

## INTERNATIONAL SEARCH REPORT

International Application No  
PC 1/US 95/09576

## A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C12N15/64 C12N15/67 C12N15/85 C12N9/72 C12N5/10

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	DNA CLONING, VOLUME III, EDITED BY D.M. GLOVER, 1987 IRL PRESS, OXFORD, GB;, pages 189-212, A.M.C. BROWN AND M.R.D. SCOTT 'Retroviral vectors'	1-3, 7, 8
Y	see page 192, line 7 - page 196, line 5; figures 2, 3  --- -/--	5, 6, 9-12, 16-21

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

## \* Special categories of cited documents:

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Date of the actual completion of the international search

23 November 1995

Date of mailing of the international search report

08.12.95

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Fax (+31-70) 340-3016

Authorized officer

Hornig, H



## INTERNATIONAL SEARCH REPORT

Int. Application No  
PCT/US 95/09576

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
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X	CELL, vol. 37, no. 3, July 1984 CELL PRESS, CAMBRIDGE, MA, US; pages 1053-1062, C.L. CEPKO ET AL. 'Construction and applications of a highly transmissible murine retrovirus shuttle vector' cited in the application	1-3,7,8
Y	pZIP-Neo SV(B)1 see figure 1	5,6, 9-12, 16-21
Y	--- MOL. CELL. BIOL., vol. 5, no. 3, March 1985 ASM WASHINGTON, DC, US, pages 431-437, A.D. MILLER ET AL. 'Generation of helper-free amphotrophic retroviruses that transduce a dominant-acting, methotrexate-resistant dihydrofolate reductase gene' see page 432, right column, line 5 - page 436, right column, line 7; figure 1	5,6, 9-12, 16-21
Y	WO,A,94 05784 (US) 17 March 1994  see the whole document	5,6, 9-12, 16-21
Y	--- EP,A,0 215 548 (ZYMOGENETICS INC ;UNIV WASHINGTON (US)) 25 March 1987  see the whole document	5,6, 9-12, 16-21
A	--- WO,A,92 17566 (GENENTECH INC) 15 October 1992 cited in the application see the whole document	1-21
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# INTERNATIONAL SEARCH REPORT

Int ional Application No  
PCT/US 95/09576

## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>PROC. NATL.ACAD SCI., vol. 86, February 1989 NATL. ACAD SCI., WASHINGTON, DC, US;, pages 1041-1045, M. VIVAUD ET AL. 'A 5' splice-region G-C mutation in exon 1 of the human beta-globin gene inhibits pre-mRNA splicing: A mechanism for beta+-thalassemia' see the whole document -----</p>	1-4

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Information on patent family members

International Application No

PCT/US 95/09576

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		FR-A- 2603899	18-03-88
		GB-A, B 2197321	18-05-88
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		AU-B- 5295890	30-08-90
		EP-A- 0385558	05-09-90
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		NO-B- 174934	25-04-94
		SG-A- 3994	10-06-94
		US-A- 4965199	23-10-90

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